eurofins | Environment Testing

2025

Analytical Method Summaries

ANALYTICAL METHOD SUMMARIES

Analyte	Method Summary	Reference Method
Organics Laboratory		
1,4-Dioxane	A water sample that has been dechlorinated and preserved with a microbial inhibitor is fortified with the isotopically labelled surrogate, 1,4-dioxane-d_8. The sample is extracted by one of two SPE options. In option 1, a 500-mL sample is passed through an SPE cartridge containing 2 g of coconut charcoal to extract the method analyte and 1,4-dioxane-d_8. In option 2, a 100-mL sample is extracted on a Waters AC-2 Sep-Pak or Supelco Supelclean ENVI-Carb Plus cartridge. In either option, the compounds are eluted from the solid phase with a small amount of dichloromethane (DCM), approximately 9 mL or 1.5 mL, respectively. The extract volume is adjusted, and the IS, tetrahydrofuran-d_8 (THF-d_8), is added. Finally, the extract is dried with anhydrous sodium sulfate. Analysis of the extract is performed by GC/MS. The data provided in this method were collected using splitless injection with a high-resolution fused silica capillary GC column that was interfaced to an MS operated in the SIM mode. The analyte, 1,4-dioxane-d_8 and IS are separated and identified by comparing the acquired mass spectra and retention times to reference spectra and retention times for calibration standards acquired under identical GC/MS conditions. The concentration of the analyte is determined by comparison to its response in calibration standards relative to the IS. Analyse samples, standards, blanks, and Quality Control samples by GC-MS using in-house method LTM-ORG-2270.	**US EPA Method 522 DETERMINATION OF 1,4-DIOXANE IN DRINKING WATER BY SOLID PHASE EXTRACTION (SPE) AND GAS CHROMATOGRAPHY MASS SPECTROMETRY (GC/MS) WITH SELECTED ION MONITORING (SIM): EPA/600/R-08/101**
Methamphetamine in Swabs	**TARGET ANALYTES**: • Ephedrine • Pseudoephedrine • Amphetamine • Methamphetamine • MDA • MDMA **SAMPLING**: 1. Using a new pair of nitrile gloves, remove the wipe from its protective package that is contained in the supplied kit. NOTE: Do not use vinyl gloves due to the potential for leaching of phthalate plasticisers and contamination of the samples.	**NIOSH Method 9111 - METHAMPHETAMINE on Wipes by Liquid Chromatography/Mass Spectrometry**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	2. Place the supplied 10 cm x 10 cm template over the area to be sampled (may tape in place along outside edge of template). Wipe the surface to be sampled with firm pressure, using vertical S-strokes. Fold the exposed side of the pad in and wipe the area with horizontal S-strokes. Fold the pad once more and wipe the area again with vertical S-strokes. 3. Fold the pad, exposed side in, and place in supplied shipping container and seal with cap. NOTE: Keep samples refrigerated (<6ºC). While nicotine and related compounds are stable on the recommended wipe media for at least 7 days at room temperature, refrigeration is recommended as soon as possible. 4. Clean the template before use for the next sample or use a new disposable template. 5. Label each sample clearly with a unique sample identifier. 6. Prepare a minimum of two field blanks with one field blank for every ten samples. **SAMPLE PREPARATION**: 7. Desorption from media: a. Remove cap from shipping container. Sample media should fit loosely in the container. If not, rearrange media carefully with rinsed forceps or transfer to a larger container. If the sample media are transferred to a larger container, do not discard the original container. Samples may consist of more than one wipe. If this is the case, internal standard and desorption solution volumes may be adjusted accordingly. b. Spike exactly 50 µL of internal standard spiking solution onto each wipe sample. c. Add 30 mL desorption solution. If the samples were transferred to a larger container, the original shipping container must be rinsed with the desorption solution first, shaken, and the rinsate decanted into the larger container. d. Cap securely and mix contents by inverting the tubes end over end on a rotary mixer at 10-30 rpm for at least one hour. NOTE 1: The desorption solution must percolate freely through the gauze wipes. e. Filter an aliquot of the sample through a 0.45 µm membrane. 8. Transfer the filtered sample into a vial and cap. 9. Analyse samples, standards, blanks, and Quality Control samples by LC-MS/MS or LC-QToF-MS using in-house method LTM-ORG-2240.	

Analyte	Method Summary	Reference Method
Organics Laboratory		
Methane in Water	The measurement of dissolved gases such as methane, ethane, and ethylene in ground water is important in determining whether intrinsic bioremediation is occurring in a fuel- or solvent contaminated aquifer. A helium headspace is generated above a water-filled bottle. Gases that are dissolved in the water partition between the gas and liquid phases and equilibrate rapidly. An aliquot of this headspace is analysed by gas chromatography to determine the gases' concentration in this phase. The concentration of the gas dissolved in the water can then be calculated based on its partitioning properties, as indicated by its Henry's Law constant using in-house LTM-ORG-2070	**Analysis of Dissolved Methane, Ethane, and Ethylene in Ground Water by a Standard Gas Chromatographic Technique., Don H. Kampbell* and Steve A. Vandegrift., U.S. Environmental Protection Agency, Journal of Chromatographic Science, Vol. 36, May 1998**
Per- and Polyfluoroalkyl Substances (PFAS) in Water - Potable, Groundwater & Surface Water	Environmental samples are prepared and extracted using method-specific procedures. Sample extracts are subjected to clean-up procedures designed to remove interferences. Analyses of the sample extracts are conducted by LC-MS/MS in the multiple reaction monitoring (MRM) mode. Sample concentrations are determined by isotope dilution or extracted internal standard quantification using isotopically labelled compounds added to the samples before extraction. Aqueous samples are spiked with isotopically labelled standards, extracted using solid-phase extraction (SPE) cartridges and undergo clean-up using carbon before analysis. This method measures the analytes as either their anions or neutral forms – see Table 4: PFAS Analytes (n=30) and Table 5: PFAS Analytes – Supplemental. Individual PFAS analytes are identified through peak analysis of the quantification and confirmation ions, where applicable. Quantitative determination of target analyte concentrations is made with respect to an isotopically labelled PFAS standard; the concentrations are then used to convert raw peak areas in sample chromatograms to final concentrations. Results for target analytes are recovery corrected by the method of quantification (i.e., either isotope dilution or extracted internal standard quantification). Isotopically labelled compound recoveries are determined by comparison to the responses of one of seven non-extracted internal standards (a.k.a., the	**US EPA Method 533: Determination of Per- And Polyfluoroalkyl Substances in Drinking Water by Isotope Dilution Anion Exchange Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry., November 2019** **US EPA Method 537.1 Determination of Selected Per- and Polyfluorinated Alkyl Substances in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS)** **Version 1.0, November 2018** **Method 1633** **Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, and Tissue**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	"recovery" standards) and are used as general indicators of overall analytical quality. The quality of the analysis is assured through reproducible calibration and testing of the extraction, clean-up, and LC-MS/MS systems. Branched and linear isomers are used for calibration standards when they are commercially available as certified standards. Table 2 lists standards that are currently commercially available and used. The target analyte response for analytes containing branched and linear isomers results from the summation of peaks from all isomers. The limit of reporting is listed in Table 3: PFAS LORs - Water, Soil/Sediments & Biotic Matrices. The LOR obtainable is dependent on the matrix and method. The reporting limit may be affected by the presence of other contaminants or components in individual samples that cause analytical interferences that raise the achievable LOR. This problem is more likely to occur in complex matrices such as soil, waste, biosolids and biota samples. An initial calibration is prepared for each native compound. Internal standard calibration is applied to determine the native compounds that do not have exact labelled analogues and are not being quantified by isotope dilution. The recoveries of the labelled analogues themselves are determined by internal standard quantitation (ISTD) and used as a quality control check on the overall analytical process. Branched and linear isomers are used for calibration standards when they are commercially available as certified standards. Table 2 lists standards that are currently commercially available and used. The target analyte response for analytes containing branched and linear isomer result from the summation of peaks from all isomers. Limit of reporting is listed in Table 3: PFAS LORs - Water, Soil/Sediments & Biotic Matrices. The LOR obtainable is dependent on the matrix and method. The limit of reporting may be affected by the presence of other contaminants or components in individual	**Samples by LC-MS/MS, January 2024** **Department of Defense (DoD) Department of Energy (DOE) Consolidated Quality Systems Manual (QSM) for Environmental Laboratories** **Manual Version 6.0 2023** **UNITED STATES DEPARTMENT OF DEFENSE** **Table B-24: Per- and Polyfluoroalkyl Substances (PFAS) Analysis by LC/MS/MS (Method 1633) 2023**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	samples that cause analytical interferences that raise the achievable LOR. This problem is more likely to occur in complex matrices such as soil, waste, biosolids and biota samples. Analyse samples, standards, blanks, and Quality Control samples by LC-MS/MS using in-house method LTM-ORG-2100.	
Per- and Polyfluoroalkyl Substances (PFAS) in Soils, Sediments, Biosolids & Tissues	Environmental samples are prepared and extracted using method-specific procedures. Sample extracts are subjected to clean-up procedures designed to remove interferences. Analyses of the sample extracts are conducted by LC-MS/MS in the multiple reaction monitoring (MRM) mode. Sample concentrations are determined by isotope dilution or extracted internal standard quantification using isotopically labelled compounds added to the samples before extraction. Solid samples are spiked with isotopically labelled standards, extracted into basic methanol, and cleaned up by carbon and SPE cartridges before analysis. This method measures the analytes as either their anions or neutral forms. Individual PFAS analytes are identified through peak analysis of the quantification and confirmation ions, where applicable. This method measures the analytes as either their anions or neutral forms – see Table 4: PFAS Analytes (n=30) and Table 5: PFAS Analytes – Supplemental. Individual PFAS analytes are identified through peak analysis of the quantification and confirmation ions, where applicable. Quantitative determination of target analyte concentrations is made with respect to an isotopically labelled PFAS standard; the concentrations are then used to convert raw peak areas in sample chromatograms to final concentrations. Results for target analytes are recovery corrected by the method of quantification (i.e., either isotope dilution or extracted internal standard quantification). Isotopically labelled compound recoveries are determined by comparison to the responses of one of seven non-extracted internal standards (a.k.a., the “recovery” standards) and are used as general indicators of overall analytical quality.	**Method 1633** **Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, and Tissue Samples by LC-MS/MS, January 2024** **Department of Defense (DoD) Department of Energy (DOE) Consolidated Quality Systems Manual (QSM) for Environmental Laboratories** **Manual Version 6.0 2023** **UNITED STATES DEPARTMENT OF DEFENSE** **Table B-24: Per- and Polyfluoroalkyl Substances (PFAS) Analysis by LC/MS/MS (Method 1633) 2023**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	The quality of the analysis is assured through reproducible calibration and testing of the extraction, clean-up, and LC-MS/MS systems. Branched and linear isomers are used for calibration standards when they are commercially available as a certified standard. Table 2 lists standards that are currently commercially available and used. The target analyte response for analytes containing branched and linear isomer result from the summation of peaks from all isomers. The limit of reporting (LOR) is listed in Table 3: PFAS LORs - Water, Soil/Sediments & Biotic Matrices. The LOR obtainable is dependent on the matrix and method. The limit of reporting may be affected by the presence of other contaminants or components in individual samples that cause analytical interferences that raise the achievable LOR. This problem is more likely to occur in complex matrices such as soil, waste, biosolids and biota samples. Analyse samples, standards, blanks, and Quality Control samples by LC-MS/MS using in-house method LTM-ORG-2100.	
Per- and Polyfluoroalkyl Substances (PFAS) in Biotic Matrices	The sample is cryogenically milled with dry ice, a homogenate taken and spiked with isotopically labelled surrogates solution. The sample is sonicated and vortexed and then neutralised with HCl before adding acetonitrile and again sonicated and vortexed before QuEChERS extraction is undertaken. Depending on the particular biotic matrix, ENVI-carb SPE may be utilised prior to mixed-mode reversed phase WAX SPE was used before concentration to a known volume. An injection is made into an LC equipped with a C18 column that is interfaced with a tandem mass spectrometer (MS/MS). The analytes are separated and identified by comparing the acquired mass spectra and retention times to reference spectra and retention times for calibration standards acquired under identical LC-MS/MS conditions. The concentration of each analyte is determined by using the isotope dilution technique. Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available – see	**Method 1633** **Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, and Tissue Samples by LC-MS/MS, January 2024** **Department of Defense (DoD) Department of Energy (DOE) Consolidated Quality Systems Manual (QSM) for Environmental Laboratories** **Manual Version 6.0 2023** **UNITED STATES DEPARTMENT OF DEFENSE**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	Table below. Labelled compounds are enriched with deuterium to produce ^{2}H-labeled analogues, stable isotopes of oxygen-18 to produce ^{18}O-labelled analogues or carbon-13 to produce ^{13}C-labeled analogues. The labelled analogues are spiked into each sample to allow identification and correction of the concentration of the native compounds in the extraction, clean-up and the analytical process. Correction of report results along with a statement of the recovery for labelled analogues are included in the certificate of analysis. Typical recoveries are between 50-150% (± 50%) depending on media and the specific analyte. An initial calibration is prepared for each native compound. Internal standard calibration is applied to the determination of the native compounds that do not have exact labelled analogues and that are not being quantified by isotope dilution. The recoveries of the labelled analogues themselves are determined by internal standard quantitation (ISTD) and used as a quality control check on the overall analytical process. Branched and linear isomers are used for calibration standards when they are commercially available as a certified standard. Table 2 lists standards that are currently commercially available and used. The target analyte response for analytes containing branched and linear isomer result from the summation of peaks from all isomers. Limit of reporting is listed in Table 3: PFAS LORs - Water, Soil/Sediments & Biotic Matrices. The LOR obtainable is dependent on the matrix and method. The limit of reporting may be affected by the presence of other contaminants or components in individual samples that cause analytical interferences that raise the achievable LOR. This problem is more likely to occur in complex matrices such as soil, waste, biosolids and biota samples. Analyse samples, standards, blanks, and Quality Control samples by LC-MS/MS using in-house method LTM-ORG-2320.	**Table B-24: Per- and Polyfluoroalkyl Substances (PFAS) Analysis by LC/MS/MS (Method 1633) 2023**
	Applicability. OTM-45 is a performance-based method applicable to the collection and	

Analyte	Method Summary	Reference Method
Organics Laboratory		
Per- and Polyfluoroalkyl Substances (PFAS) in Air Matrices	quantitative analysis of specific semi-volatile (Boiling point > 100°C) and particulate-bound per- and polyfluorinated alkyl substances (PFAS) in air emissions from stationary sources. This method can also be used for the collection and recovery of ionic and covalent PFAS for nontargeted analysis (NTA) of PFAS compounds. This method describes the sampling and sample recovery procedures used to measure individual semi-volatile PFAS from stationary source air emissions. OTM-45 incorporates by reference some of the specifications (e.g., equipment and supplies) and procedures (e.g., sampling and sample preparation) from other methods that are essential to conducting OTM-45. To obtain reliable samples, source sampling teams should be trained and experienced with the following additional EPA test methods: Method 1; Method 2; Method 3; Method 4; and Method 5 of Appendices A-1, A-2, and A-3 to 40 Code of Federal Regulations (CFR) Part 60. Laboratory analysis teams should be trained and experienced in the use of liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS) multiple reaction monitoring (MRM) as described in EPA Method 537.1 & 533 and Method 1633. This method identifies and determines the concentration in mass per unit gas volume sampled of specific PFAS compounds in source emissions. Gaseous and particulate bound target pollutants are withdrawn from the gas stream isokinetically and collected in the sample probe, on a glass fibre or quartz filter, on a packed column of adsorbent material and in a series of impingers. The target compounds are extracted from the individual sample collection media. The OTM-45 train results in four (4) discreet sample extract fractions for analysis. The extracts are analysed by LCMS/MS in the MRM detection mode. Quantification of each analyte is calculated using the isotope dilution technique. For QC purposes, the percent recoveries of the pre-extraction standards are calculated using the integrated peak areas of pre-analysis standard(s), which are added to the final extract and function as traditional internal standards, exclusively applied to the pre-extraction standards. The use of pre-sampling standards added to XAD-2 collection media prior to sampling and analysed in the same manner as targeted PFAS compounds serves	**Other Test Method 45 (OTM-45) Measurement of Selected Per- and Polyfluorinated Alkyl Substances from Stationary Sources**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	as an indication of the method's quantitative capture efficiency. This method is not intended to differentiate between target compounds in particle or vapor fractions. This method uses isotopically labelled standards to improve method accuracy and precision.	
Per- and Polyfluoroalkyl Substances (PFAS) in Air Matrices	OTM-50 is a performance-based method applicable to the collection and quantitative analysis of specific volatile fluorinated compounds (VFCs). The target list originates from known industrial products and products of incomplete thermal destruction. This method can be used to collect and analyse gas samples into 6-litre passivated silicon ceramic lined stainless-steel canisters, or equivalent, from stationary sources for the purpose of determining the concentration of target VFCs in Table OTM-50-1. Gaseous emissions samples collected by this method are intended to be sampled from industrial source ducts, vents, stacks, etc. Samples may be collected directly from the non-combustion stacks or vents when the moisture is relatively low (<3% volume/volume (v/v)), when sample gas temperatures are <250°F (121°C), and when acid gases are not present. To collect samples outside these criteria, sample conditioning, including moisture, acid gas, and temperature management, as described in this method, is required. This method involves collection of VFCs in evacuated passivated silicon ceramic lined stainless-steel canisters, or equivalent. Field sampling and recovery staff must be trained in the best practices for extracting representative gas samples from an emission vent or stack, conditioning the gas if necessary to manage moisture and acid gases, and collecting the gas sample in passivated stainless-steel canisters. Gas samples are collected from a sampling manifold using evacuated canisters equipped with a critical orifice for sample flow control. Samples are collected directly from the manifold with or without a water and acid gas management system. Water and acid gas management in the sampling system is described in this method. 4 This method provides procedures for the preparation, blanking, evacuation, and shipping of passivated canisters prior to sample collection. Sample transport, preservation and storage are also described. Measurements of stack carbon	**Other Test Method 50 (OTM-50) Sampling and Analysis of Volatile Fluorinated Compounds from Stationary Sources Using Passivated Stainless-Steel Canisters**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	dioxide (CO_2) are also required. The volumetric concentration of CO_2 is important information for the proper preparation of calibration standards and analytical laboratory method optimization. The source moisture concentration is necessary to determine what sampling train configuration is appropriate. The volumetric moisture concentration is also needed to calculate results on both a wet and dry basis. Water and CO2 measurements in conjunction with stack or duct volumetric flows are required as part of this method. VFCs are identified and quantified in gas samples from canisters by GC/MS. Additional volatile compounds present in canister samples that are not on the target list are reported with their best-available matches to mass spectral reference libraries.	
Determination of Organotins in Solid & Aqueous samples by LC-ICP-MS	This in-house method details a rapid method developed for the determination of Monobutyltin (MBT), Dibutyltin (DBT) and Tributyltin (TBT) by liquid chromatography coupled to inductively coupled plasma-mass spectrometry (LC-ICP-MS) in solids (soil, sediment, biosolids), and aqueous (drinking water, surface water, saline water, waste water, leachates) samples by solid phase extraction (SPE). This method provides a viable alternative to the more common analysis by gas chromatography-inductively coupled plasma-mass spectrometry for contaminated sediment without the requirement of sample derivatisation. Only analysts who have proficiency in the following attributes are allowed to utilise this method: trained and gained practical experience in LC-ICP-MS operations, LC-ICP-MS troubleshooting and maintenance skills, understanding of LC-ICP-MS principles and techniques and proficiency in chromatographic and Mass Spectrometry interpretations. Organotin compounds (OTCs) are organometallic derivatives of tin that have the highest number of commercial applications than any other element. Mostly they have been used as stabilisers and catalysts in the production of polyvinyl chloride (PVC), polyurethanes and silicones. Tributyltin (TBT) is the most common of a group of organotin compounds, which have widespread usage in marine antifouling points and for wood preservation. Due to their widespread use, OTCs may be found in different ecosystems. Today, their high toxicity for	**Bishop, D, P. et al. (2015), Speciation and quantification of organotin compounds in sediment and drinking water by isotope dilution liquid chromatography-inductively coupled plasma-mass spectrometry Anal. Methods, 2015, 7, 5012.** **Zuliani, Tea & Lespes, Gaetane & Milacic, Radmila & Ščančar, Janez. (2010). Development of the extraction method for the simultaneous determination of butyl, phenyl and octyltin compounds in sewage sludge. Talanta. 80. 1945-51. 10.1016/j.talanta.2009.10.050.**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	aquatic and terrestrial organisms is well documented. As a result of its physical and chemical properties nearly all TBT found in natural waters is bound to suspended particles in the water or associated to dissolved organic matter. Tributyltin Oxide (TBTO) is usually applied as a slow-release coating to boats. Under various conditions, different authors have estimated that 10 to 90% of Tributyltin Oxide introduced into water is adsorbed onto particles. Some of the factors on which the adsorption depends are temperature; salinity nature, size and amount of suspended particles and presence of dissolved organic matter in the water. As particles in the water column may settle out, TBT is removed from the water and introduced into the sediment. Organotin compounds are common contaminants in Australia in ports and harbours and are frequently present at high levels in berths and inner harbour areas. This method is designed to detect the organotin compounds without the use of either hydrolysis or derivatisation in the extraction procedure. Aqueous samples require extraction and are passed through a Bond Elut C18 SPE cartridge. The collected extract is concentrated down to dryness using a nitrogen blowdown system with a heated water bath. The samples are reconstituted in a set volume of mobile phase. Soil/sediment samples are extracted using an organic solvent. For all samples the final extract is injected into an LC-ICP-MS where the analytes are separated by reverse phase liquid chromatography and analysed via inductively coupled plasma – mass spectrometry (LC-ICP-MS). Identification of the Organotins and surrogate is primarily by retention time and comparison of retention times (and intensity ratios) with those of the calibration standards. External standard approach using a single-quadrupole mass spectrometer is used for quantitative detection analysis. Organotin compounds reported are Tributyltin Tributyltin as Sn Tributyltin Oxide Dibutyltin Dibutyltin as Sn	

Analyte	Method Summary	Reference Method
Organics Laboratory		
	Monobutyltin Monobutyltin as Sn Analyse samples, standards, blanks, and Quality Control samples by LC-ICP/MS using in-house method LTM-ORG-2400.	
Pharmaceuticals and personal care products (PPCPs) in Aqueous Samples by LC-MS/MS	Pharmaceuticals and personal care products (PPCPs) include any product used by individuals for personal health or cosmetic reasons, or used by agribusiness to enhance growth or health of livestock. This definition encompasses thousands of chemicals that make up fragrances, cosmetics, over-the-counter drugs, and veterinary medicines. PPCPs enter the environment in many ways (e.g. wastewater discharge) are increasingly being detected at low levels in ground and surface waters around the world. PPCPs are regarded as chemicals of emerging concern (CEC) because little is known about their impact on the environment, particularly to aquatic organisms. In-house method LTM-ORG-2390 is for the determination of pharmaceuticals and personal care products (PPCPs) in aqueous environmental samples by high performance liquid chromatography combined with tandem mass spectrometry (LC-MS/MS). Refer to Table 6 for complete analyte list. Analyse samples, standards, blanks, and Quality Control samples by LC-MS/MS using in-house method LTM-ORG-2390.	**US EPA Method 1694: Pharmaceuticals and Personal Care Products in Water, Soil, Sediment and Biosolids by HPLC/MS/MS, December 2007** **US EPA-820-R-10-008: Stability of Pharmaceuticals, Personal Care Products, Steroids, and Hormones in Aqueous Samples, POTW Effluents, and Biosolids. Sept 2010**
Total Oxidisable Precursor Analysis (TOPA) - Screen	Samples are treated via hydroxyl radical oxidation using an activated agent with overnight heating which converts the masked fluorinated precursors to their equivalent detectable perfluoroalkyl carboxylic acids (PFCAs) and perfluoroalkyl sulfonates (PFSAs). LC-MS/MS is done before and after TOP Analysis.	**Oxidative Conversion as a Means of Detecting Precursors to Perfluoroalkyl Acids in Urban Runoff, Erika F. Houtz and David L. Sedlak, Environ. Sci. Technol. 2012**
Total Oxidisable Precursor Analysis (TOPA) – Detailed	Detailed TOPA includes multiple mass-labelled analogues added prior to the “cooking” step and the recoveries provided along with dilutions required are reported in the certificate of analysis. Sum of all PFAS are reported pre- and post TOPA along with. Results for total PFAS concentration post-TOPA should be greater or equal to the total PFAS concentration pre-TOPA, (signifies no material losses observed in	**Draft PFAS National Environmental Management Plan: Version 3.0** **National Chemicals Working Group of the Heads of EPAs**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	preparation steps, noting a decrease of up to 10% might be expected due to normal analytical variability). – the sum of PFCA post-TOPA should be equal to or greater than the sum of PFCA pre-TOPA, which signifies any precursors being converted to PFCA products. – the sum of PFSA post-TOPA should approximate the sum of PFSA pre-TOPA, signifying that precursors did not convert to PFSA products. – for a full oxidation, no PFAA precursors (e.g. 6:2 FTS, FOSA) are detectable post oxidation, signifying complete oxidation. – for situations where a near complete oxidation is acceptable, minimal PFAA precursors are detectable post oxidation signified by: • for aqueous samples, sum of [PFAA precursors] divided by sum of [Total PFAS] <5%. • for soil samples, sum of [PFAA precursors] divided by sum of [Total PFAS] <10%. • noting greater leniency may be applied for samples where PFAS were detected ≤ 10 times LOR.	**Australia and New Zealand** **2022**

Analyte	Method Summary	Reference Method

Organics Laboratory

Table 1: Per- and Polyfluoroalkyl Substances (PFAS)

Native PFAS Perfluoroalkyl carboxylic acids (PFCAs)	Extracted Internal Standard Analytes (EIS) Isotope Dilution Quantification Standard
Perfluorobutanoic acid (PFBA)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]butanoic acid
Perfluoropentanoic acid (PFPeA)	Perfluoro-n-[1,2,3,4,5-$^{13}C_5$]pentanoic acid
Perfluorohexanoic acid (PFHxA)	Perfluoro-n-[1,2,3,4,5-$^{13}C_5$]hexanoic acid
Perfluoroheptanoic acid (PFHpA)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]heptanoic acid
Perfluorooctanoic acid (PFOA)	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octanoic acid
Perfluorononanoic acid (PFNA)	Perfluoro-n-[1,2,3,4,5-$^{13}C_5$]nonanoic acid
Perfluorodecanoic acid (PFDA)	Perfluoro-n-[1,2,3,4,5,6-$^{13}C_6$]decanoic acid
Perfluoroundecanoic acid (PFUnA)	Perfluoro-n-[1,2-$^{13}C_2$]undecanoic acid
Perfluorododecanoic acid (PFDoA)	Perfluoro-n-[1,2-$^{13}C_2$]dodecanoic acid
Perfluorotridecanoic acid (PFTrDA)	Perfluoro-n-[1,2-$^{13}C_2$]tetradecanoic acid ISTD
Perfluorotetradecanoic acid (PFTeDA)	Perfluoro-n-[1,2-$^{13}C_2$]tetradecanoic acid
Perfluoroalkane sulfonic acids (PFSAs)	
Perfluoropropanesulfonic acid (PFPrS)	Sodium perfluoro-n-[2,3,4-$^{13}C_3$]butane sulfonate ISTD
Perfluorobutanesulfonic acid (PFBS)	Sodium perfluoro-n-[2,3,4-$^{13}C_3$]butane sulfonate
Perfluoropentane sulfonic acid (PFPeS)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]octane sulfonic acid ISTD
Perfluorohexane sulfonate (PFHxS)	Sodium perfluoro-n-[$^{18}O_2$]hexanesulfonate
Potassium perfluorohexanesulfonate (linear and branched isomers) (br-PFHxSK)	Sodium perfluoro-n-[$^{18}O_2$]hexanesulfonate
Perfluoroheptane sulfonate (PFHpS)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]octane sulfonic acid
Perfluorooctane sulfonate (PFOS)	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octane sulfonate
Potassium perfluorooctanesulfonate (linear and branched isomers) (br-PFOSK)	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octane sulfonate
Perfluorononanesulfonic acid (PFNS)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]octane sulfonic acid ISTD
Perfluorodecanesulfonic acid (PFDS)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]octane sulfonic acid ISTD
Perfluoroalkane sulfonamides (FASAs), Perfluoroalkane sulfonamido ethanols (FASEs) and N-alkyl perfluoroalkane sulfonamido ethanols (MeFA Perfluoroalkane sulfonamido acetic acids (FASAAs) and N-alkyl perfluoroalkane sulfonamido acetic acids (MeFASAAs, EtFASAAs)	
Perfluorooctane sulfonamide (FOSA)	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octane sulfonamide
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	N-methyl-d_3-perfluoro-n-octanesulfonamide
N-ethylperfluoro-1-octanesulfonamide (N-EtFOSA)	N-ethyl-d_5-perfluoro-n-octanesulfonamide
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol (N-MeFOSE)	2-(N-methyl-d_3-perfluoro-1-octane sulfonamido)-ethanol-d_4
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol (N-EtFOSE)	2-(N-ethyl-d_5-perfluoro-1-octane sulfonamido)-ethanol-d_4
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	N-ethyl-d_5-perfluoro-n-octanesulfonamidoacetic acid
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MEFOSAA)	N-methyl-d_3-perfluoro-1-octanesulfonamidoacetic acid
Fluorotelomers n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2 FTSA)	1H,1H,2H,2H-perfluoro-1-[1,2-$^{13}C_2$]hexane sulfonate (4:2 FTSA)
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2 FTSA)	1H,1H,2H,2H-perfluoro-1-[1,2-$^{13}C_2$]octane sulfonate (6:2 FTSA)
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2 FTSA)	1H,1H,2H,2H-perfluoro-1-[1,2-$^{13}C_2$]decane sulfonate (8:2 FTSA)
1H, 1H, 2H, 2H-perfluorododecane sulfonic Acid (10:2 FTSA)	1H,1H,2H,2H-perfluoro-1-[1,2-$^{13}C_2$, D4]dodecane sulfonate (10:2 FTSA)
Additional PFAS Compounds	
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX)	$^{13}C_3$-Hexafluoropropylene oxide dimer acid ($^{13}C_3$-HFPO-DA)
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11Cl-PF3OUdS) [11Cl-F53B]	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octane sulfonate
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9Cl-PF3ONS) [9Cl-F53B]	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octane sulfonate ISTD
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octanoic acid ISTD
Nonafluoro-3,6-dioxaheptanoic acid (NFDHA)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]heptanoic acid ISTD
Perfluoro(2-ethoxyethane)sulfonic acid (PFEESA)	Sodium perfluoro-n-[2,3,4-$^{13}C_3$]butane sulfonate ISTD
Perfluoro-3-methoxypropanoic acid (PFMPA)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]butanoic acid ISTD
Perfluoro-4-methoxybutanoic acid (PFMBA)	Perfluoro-n-[1,2,3,4,5-$^{13}C_5$]pentanoic acid ISTD
6:2 fluorotelomer sulfonamide alkylbetaine (6:2 FTAB)	N-ethyl-d_5-perfluoro-n-octanesulfonamide ISTD
3:3 Fluorotelomercarboxylic acid (3:3 FTCA)	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octanoic acid ISTD
5:3 Fluorotelomer carboxylic acid (5:3 FTCA)	Perfluoro-n-[1,2,3,4,5,6,7,8-$^{13}C_8$]octanoic acid ISTD
Perfluoroethylcyclohexane sulfonate (PFECHS)	Perfluoro-n-[1,2,3,4-$^{13}C_4$]octane sulfonic acid ISTD

Analyte	Method Summary	Reference Method
Organics Laboratory		
	Table 2: Currently Available Certified PFAS Standards Containing Branched and Linear Isomers • Perfluorohexanesulfonic acid (PFHxS) • Perfluorooctanesulfonic acid (PFOS) • 2-(N-methylperfluorooctanesulfonamido) acetic acid (NMeFOSAA) • 2-(N-ethylperfluorooctanesulfonamido) acetic acid (NEtFOSAA) The instrument calibration summary should identify which analytes were calibrated using standards that contained branched and linear isomers of the analyte. Branched and linear isomers should be used for calibration standards when they are commercially available as a certified standard. Table 2 lists standards that are currently commercially available and used. The target analyte response for analytes containing branched and linear isomer should be result of the summation of peaks from all isomers. If a certified standard is not available, a technical standard may be used to identify retention time and ion transition ratios, but may not be used for calibration. In these instances, a certified linear standard should be used to build the calibration curve, and the samples must be quantified for all isomers that meet the technical grade standard identification for retention time and ion transitions.	
Total Organofluorine (TOF)	The determination of total organic fluorine (TOF) approaches quantitation of the unknown mass of PFAS from the angle of determining the fluorine content of a sample. With the use of combustion ion chromatography (CIC) a wide range of matrices are analysed for Total Fluorine (TF), Total Organic Fluorine (TOF), Total Adsorbable Organofluorine (AOF) in water, or Extractable Organic Fluorine (EOF) in solids. NOTE: TOF which is mainly reserved for products such as AFFF, textiles, food packaging, highly contaminated aqueous samples, etc. and is a direct combustion of these samples with inorganic fluoride also being measured. For organic fluorine without the contribution from inorganic fluoride please see below. NOTE: TF = IF + OF OF = E(A)OF + NE(NE(A)OF) Where TF – Total F	**Modified ASTM D7359-18 Standard Test Method for Total Fluorine, Chlorine and Sulfur in Aromatic Hydrocarbons and Their Mixtures by Oxidative Pyrohydrolytic Combustion followed by Ion Chromatography Detection (Combustion Ion Chromatography-CIC) for Total F**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	IF = Inorganic F OF – Organically bound fluorine E(A)OF – Extractable (absorbable organically bound F) NE(A)OF = non-extractable (non-absorbable organically bound fluorine. If **Total PFAS** shall be measured the non-absorbable/non-extractable fraction must be made absorbable/extractable!!!	
Adsorbable Organofluorine (AOF)	For the trace level determination of adsorbable organic fluorine (AOF) in water, the sample must first be passed through a mixed-mode weak anion exchange solid-phase extraction (SPE) cartridge thereby adsorbing the PFAS compounds. AOF is then determined by eluting the contents of the SPE cartridge with NaOH in methanol, evaporating and reconstituting the extract, and finally determining the fluoride content of the extract by CIC. The LOR is dependent on the volume passed through the SPE, so the presence of suspended solids does impose limits on the procedure, but for clean waters the LOR is 0.01 mg F/L. Where significant levels of suspended solids are encountered the LOR may be limited to 0.1 mg F/L and the suspended solids may be determined separately by direct combustion.	**Modified ASTM D7359-18 Standard Test Method for Total Fluorine, Chlorine and Sulfur in Aromatic Hydrocarbons and Their Mixtures by Oxidative Pyrohydrolytic Combustion followed by Ion Chromatography Detection (Combustion Ion Chromatography-CIC) for Total F** **Method 1621** **Determination of Adsorbable Organic Fluorine (AOF) in Aqueous Matrices by Combustion Ion Chromatography (CIC)** **January 2024**
Extractable Organofluorine (EOF)	For solid samples, where LORs lower than the direct combustion method of 0.5 mg F/kg are required, extraction can be performed using the same solvent systems used for conventional targeted LC-MS/MS methods. The resulting concentrate is then combusted, giving an extractable organofluorine result. A LOR of 0.2 mg F/kg is achievable. In-house method LTM-ORG-2380 Determination of Total Organofluorine by Combustion Ion Chromatography (CIC).	Extraction follows **Method 1633 - Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, and Tissue Samples by LC-MS/MS, January 2024** Analysis by modified **ASTM D7359-18 Standard Test Method for Total Fluorine, Chlorine and Sulfur in**

Analyte	Method Summary	Reference Method
Organics Laboratory		
		Aromatic Hydrocarbons and Their Mixtures by Oxidative Pyrohydrolytic Combustion followed by Ion Chromatography Detection (Combustion Ion Chromatography-CIC) for Total F
Tyre Wear Compounds **6-PPD quinone, HMMM & 1,3-DPG**	Aqueous samples are prepared and extracted using solid-phase extraction (SPE) procedures. Analyses are conducted by LC/MS/MS to identify 6PPD-Q (2-anilo-5-[(4-methylpentan-2-yl)amino]cyclohexa-2,5-diene-1,4-dione, or N-(1,3-Dimethylbutyl)-N′-phenyl-p-phenylene-diamine-quinone), hexamethoxymethylmelamine (HMMM) & 1,3-diphenylguanidine (DPG) based on retention time and multiple reaction monitoring (MRM) transitions and quantify it by isotope dilution techniques in environmental samples.	**USEPA Method 1634** **Determination of 6PPD-Quinone in Aqueous Matrices Using Liquid Chromatography with Tandem Mass Spectrometry (LC/MS/MS)** **January 2024**
TRH (Volatile)/BTEX **C6-C10 – 2013 NEPM Fractions** **C6-C9 – 1999 NEPM Fractions**	A 10 g soil sample is extracted with 20 mL methanol, tumbled for 1 hour, and analysed with solvent and instrument check surrogates. Clay samples must be completely disintegrated before an aliquot is taken for analysis. Water direct injection of supplied sample (unopened) and analysis with solvent and instrument check surrogates. Analysis by capillary column Purge and Trap GCMS (Eurofins in-house method numbers: Total Recoverable Hydrocarbons (TPH), Method: LTM-ORG-2010, Method: LTM-GEN-7080 Moisture). Owing to the differential responses of mass spectrometric detectors towards aliphatic and aromatic compounds, it is essential that the standard contain representatives of both groups. This standard should therefore consist of about 40% aromatic and 60% aliphatic target analytes, to be representative of a typical Australian fuel. The aromatic compounds shall comprise the components of BTEX. The aliphatics shall comprise equal proportions of all n-alkanes in the nC_6 to nC_{10} range. Analyse samples, standards, blanks, and Quality Control samples by GC-MS using in-house method LTM-ORG-2010.	**USEPA Method 8260D** **NEPM 2013 Schedule B3 Appendix 1: Determination of total recoverable hydrocarbons (TRH) in soil**
Total Recoverable Hydrocarbons C10- C36 – 1999 NEPM Fractions	Soil – 10 g soil and anhydrous sodium sulfate extracted with 20 mL dichloromethane/acetone (1:1), and tumbled for a minimum of 1 hour. Clay samples must be completely	**USEPA Method 8015C** **NEPM 2013 Schedule B3 Appendix 1:**

Analyte	Method Summary	Reference Method
Organics Laboratory		
>C10-C40 – 2013 NEPM Fractions	disintegrated before an aliquot is taken for analysis. Water - One 250 mL of water sequentially extracted in a separatory funnel three times with 20mL dichloromethane. Analysis by capillary column GC/FID (Eurofins in-house method numbers: Total Recoverable Hydrocarbons (TRH), Method: LTM-ORG-2010, Method: LTM-GEN-7080 Moisture)	**Determination of total recoverable hydrocarbons (TRH) in soil**
TRH (Silica Gel)	Sample extracts obtained from the appropriate TRH method are exchanged to a non-polar solvent and are passed through a column containing 1 gram of 100% activated silica gel. Elution is achieved with a small volume of 1:1 DCM:pentane or 1:1 DCM:hexane. The eluted solvent is then concentrated and analysed by the appropriate TRH analysis procedure. A decanoic acid reverse surrogate is used to provide assurance of the effectiveness of the silica-gel clean-up. Analyse samples, standards, blanks, and Quality Control samples by GC-FID using in-house method LTM-ORG-2010.	**USEPA Method 3630C** **NEPM Appendix 1: Determination of total recoverable hydrocarbons (TRH) in soil**
VOCs	10 g soil extracted with 20 mL methanol, tumbled for 1 hour, and analysed with solvent and instrument check surrogates. Clay samples must be completely disintegrated before an aliquot is taken for analysis. Water direct injection of supplied sample (unopened) and analysis with solvent and instrument check surrogates. Analysis by capillary column Purge and Trap GC-MS (Eurofins in-house method numbers Method: LTM-ORG-2150, LTM-ORG-2160, and Method: LTM-GEN-7080 Moisture).	**US EPA Method 8260D**
Semi-volatile Organic Compounds (SVOCs)	The samples are prepared for analysis by gas chromatography/mass spectrometry (GC/MS) using the appropriate sample preparation (refer to Method 3500) and, if necessary, sample clean-up procedures (refer to Method 3600). The semi-volatile compounds are introduced into the GC/MS by injecting the sample extract into a gas chromatograph (GC) with a narrow-bore fused-silica capillary column. The GC column is temperature-programmed to separate the analytes, which are then detected with a mass spectrometer (MS) connected to the gas chromatograph. Analytes eluted from the capillary column are introduced into the mass spectrometer via a jet separator or a direct	**USEPA Method 8270E**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	connection. Identification of target analytes is accomplished by comparing their mass spectra with the electron impact (or electron impact-like) spectra of authentic standards. Quantitation is accomplished by comparing the response of a major (quantitation) ion relative to an internal standard using a five-point calibration curve. NOTE: This method can be used in conjunction with the following sample preparation procedures: Water (including TCLP leachates) - US EPA Methods 3510, 3520, 3535 Soil/sediment – US EPA Methods 3540, 3541, 3545, 3546 3550, 3560, 3561. Analyse samples, standards, blanks, and Quality Control samples by GC-MS/MS using in-house method LTM-ORG-2190.	
Phenols/PAHs/ PCBs/OCs & OPPs	Soil – 10 g soil, surrogates, mixed with anhydrous sodium sulfate and extracted with 20mL dichloromethane/acetone (1:1), and tumbled for a minimum of 1 hour. Clay samples must be completely disintegrated before an aliquot is taken for analysis. Water – 250 mL water sample plus surrogates triple extracted with dichloromethane (base and neutrals). Leachate – 250 mL water sample plus surrogates triple extracted with dichloromethane (base and neutrals). Analysis by capillary column GC/MS (Eurofins in-house Methods LTM-ORG-2130, LTM-ORG-2140 Method: LTM-GEN-7080 Moisture).	**USEPA Method 8270E**
Analysis of Phenoxy Acid Herbicides in Aqueous and Soil Samples by HPLC	A 100-mL water sample is adjusted to a basic pH with sodium hydroxide, shaken, and allowed to set for 1 hour to hydrolyse chlorinated esters. The sample is acidified with H_3PO_4, filtered, and the chlorinated acids are extracted from a 20-mL aliquot. The aliquot is pumped through a high-performance liquid chromatography (HPLC) cartridge (containing C-18-silica), trapping the chlorinated acids. The concentrator cartridge is valved in-line with the C-18 analytical column following extraction. The acids are separated by HPLC and detected using an ultraviolet (UV) absorption spectrometer. LABORATORY TEST METHOD NUMBER: LTM-ORG-2180	**US EPA -NERL: Method 555: Chlorinated Acids in Water Using HPLC/UV**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	Soil – 10 g soil, surrogates, mixed with anhydrous sodium sulfate are extracted using acetonitrile in an ultrasonic bath, or shaker filtered, diluted with water as appropriate, adjusted to a basic pH with sodium hydroxide, shaken, and allowed to set for 1 hour to hydrolyse chlorinated esters. The sample is acidified with H_3PO_4, filtered, and the chlorinated acids are extracted from a 20-mL aliquot. The aliquot is pumped through a high-performance liquid chromatography (HPLC) cartridge (containing C-18-silica), trapping the chlorinated acids. The concentrator cartridge is valved in-line with the C-18 analytical column following extraction. The acids are separated by HPLC and detected using an ultraviolet (UV) absorption spectrometer. LABORATORY TEST METHOD NUMBER: LTM-ORG-2180	
EXPLOSIVES **Nitroaromatics, nitramines, and nitrate esters by high performance liquid chromatography (HPLC)**	Soil – 10 g soil, surrogates, mixed with anhydrous sodium sulfate are extracted using acetonitrile in an ultrasonic bath, or shaker filtered, diluted with water as appropriate, and analysed by HPLC with UV/DAD detection. Clay samples must be completely disintegrated before an aliquot is taken for analysis. Water – 250 mL water sample plus surrogates are pre-concentrated using solid-phase extraction, as described in USEPA Method 3535 and then diluted with water as appropriate for the selected separations. Leachate – 250 mL water sample plus surrogates extracted with SPE. Analyse samples, standards, blanks, and Quality Control samples by LC-UV using in-house method LTM-ORG-2230.	**USEPA Method 8330B**
DIOXINS **Tetra- through Octa-Chlorinated Dioxins and Furans by Isotope Dilution GC-MS/MS**	This method is for determination of tetra- through octa-chlorinated dibenzo-p-dioxins (CDDs) and dibenzofurans (CDFs) in water, soil, sediment, sludge, tissue, and other sample matrices by high resolution gas chromatography/tandem mass spectrometry (HRGC-MS/MS). The seventeen 2,3,7,8-substituted CDDs/CDFs may be determined by this method. Specifications are also provided for separate determination of 2,3,7,8-tetrachloro-dibenzo-p-dioxin (2,3,7,8-TCDD) and 2,3,7,8-tetrachloro-dibenzofuran (2,3,7,8-TCDF). The detection limits and quantitation levels in this method are usually dependent on the level of interferences rather than instrumental	**USEPA Method 1613B**

<table>
<tr><th>Analyte</th><th>Method Summary</th><th>Reference Method</th></tr>
<tr><td colspan="3">Organics Laboratory</td></tr>
<tr><td></td><td>limitations. Laboratory Test Method Number: LTM-ORG-2330</td><td></td></tr>
<tr><td>PBDEs
Polybrominated diphenyl ethers by HRGC/HRMS†</td><td>This method is for determination of polybrominated diphenyl ethers in water, soil, sediment, sludge, tissue, and other sample matrices by high resolution gas chromatography/high resolution mass spectrometry (HRGC/HRMS).
<table>
<tr><th>TriBDE</th><th>TetraBDE</th><th>PentaBDE</th></tr>
<tr><td>BDE-17
BDE-28</td><td>BDE-47
BDE-49
BDE-66
BDE-71
BDE-77</td><td>BDE-85
BDE-99
BDE-100
BDE-119
BDE-126</td></tr>
<tr><th>HexaBDE</th><th>HeptaBDE</th><th>OctaBDE</th></tr>
<tr><td>BDE-138
BDE-153
BDE-154
BDE-156</td><td>BDE-183
BDE-184
BDE-191</td><td>BDE-196
BDE-197</td></tr>
<tr><th>NonaBDE</th><th>DecaBDE</th><th></th></tr>
<tr><td>BDE-206
BDE-207</td><td>BDE-209</td><td></td></tr>
</table>
The detection limits and quantitation levels in this method are usually dependent on the level of interferences rather than instrumental limitations.
†Analysis subcontracted to Eurofins GfA Lab Service GmbH – Hamburg, Germany</td><td>Method 1614A
Brominated Diphenyl Ethers in Water, Soil, Sediment, and Tissue by HRGC/HRMS
May 2010</td></tr>
<tr><td>QToF Discovery Suite
GC-QToF-MS & LC-QToF-MS</td><td>Since the launch of the first commercial instrument nearly 25 years ago, QToF-MS has largely remained in the domain of research laboratories. Eurofins is proud to bring this technology to the commercial market in an affordable and accessible format. The QToF-MS Discovery Suite consists of five new test suites tailored to the contaminated land sector:
1. PFAS Characterisation (Compact): Screen for legacy per- and polyfluoroalkyl substances (PFAS), precursors and dead end products in water, soil, sediment and aqueous film-forming foams (AFFF). Suspect screening of high-resolution data against a library containing ~100 PFAS compounds, created from authentic reference materials. Use this test for site investigations, simple source tracking, AFFF characterisation, or precursor identification.
2. PFAS Characterisation (Comprehensive): Need to dig a little deeper? Have a complex site investigation? Our comprehensive PFAS</td><td>Schymanski, E.L et al. (2014) Environ Sci Technol 48: 2097-98. DOI: 10.1021/es5002105</td></tr>
</table>

Analyte	Method Summary	Reference Method
Organics Laboratory		
	characterisation uses multiple sample extraction strategies to capture anionic, cationic and zwitterionic compounds.[5] Data is screened against a large database containing >5000 PFAS and related compounds, and your report contains an interpretative analysis of your PFAS profile from our expert team. **3. Contaminant Screening:** Suspect screening of sediment, soil, water and air samples for thousands of contaminants including pharmaceuticals and personal care products; forensic compounds, toxins, drugs and their metabolites; pesticides, industrial chemicals and surfactants; petrochemicals; food flavours and fragrances; human and plant metabolites **4. Targeted Screening & Product Verification:** Identify suspect contaminants, confirm active constituents, or investigate potential product failures in consumer products or impacted environmental samples. Unequivocal compound identification (confidence level 1)[6] and quantitative data can be provided on request. **5. Environmental Forensics:** Work with our team of highly experienced analytical chemists to design and execute a bespoke workflow for your most complex samples. We can provide technical guidance on sampling design and analysis and summarise the results in a custom report tailored to you.	
Coal Tar	This test method sets out the procedure for indicating the presence of tar or pitch in asphalt. (a) The method identifies the presence of phenol which is contained in tar or pitch *NOTE 1: Coal tar contains approximately 1% of phenol whereas bitumen does not contain any phenol.* (b) If the sample contains a mixture of tar asphalt and other non-tar material (e.g. bitumen asphalt), the method approaches its detection limit if there is less than 10% tar asphalt or less than 0.5% tar present (c) The test includes the following steps: (i) The method extracts any weakly acidic phenols from the asphalt with dilute alkali (ii) An aromatic amine reacts with nitrous acid to produce a diazo compound (iii)When a diazo compound reacts with the extracted phenol it forms a strongly coloured	**RMS Test method T542** **Identification of tar or pitch in asphalt** **NOVEMBER 2012**

Analyte	Method Summary	Reference Method
Organics Laboratory		
	diazo dye (usually red) which indicates the presence of tar in the original asphalt. *NOTE 2: Reporting is as "Tar present" or "Tar absent".* *NOTE 3: this method is non-specific and measures the presence of "phenol" so it experiences potential interferences from other phenolic compounds other than those present in coal tar and where highly weathered the existence of "phenol" may be tenuous.* *NOTE 4: Alternative procedures include the analysis of PAHs and speciated phenolics concentrating on presence of specific PAH biomarkers and cresols/xylenols.* Analyse samples, standards, blanks, and Quality Control samples by GC-QToF/MS.	

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
Total Metals (As, Cd, Cr, Cu, Ni, Pb, Zn)	A portion of soil or water undergoes acidic digestion using either microwave or automated hot block. Analysis by ICP-MS. (Eurofins in-house method LTM-MET-3040 LTM-GEN-7080 Moisture).	**USEPA Method 6020B** **USEPA Method 3010A** **USEPA Method 3015A**
Total Mercury (Hg)	A portion of soil or water undergoes acidic digestion using either microwave or automated hot block. Analysis by ICP/MS.(Eurofins in-house method LTM-MET-3040, LTM-GEN-7080 Moisture).	**USEPA Method 6020B** **USEPA Method 3010A** **USEPA Method 3015A**
Filtered Metals (As, Cd, Cr, Cu, Ni, Pb, Zn)	Filtered (0.45µm) and acidified in the field prior to analysis. Analysis by ICP-MS. (Eurofins in-house method LTM-MET-3040).	**USEPA Method 6020B** **USEPA Method 3010A** **USEPA Method 3015A**
Filtered Mercury (Hg)	Filtered, oxidation and final reduction. Analysis by FIMS. (Eurofins in-house method LTM-MET-3040).	**USEPA Method 7471B** **USEPA Method 3010A** **USEPA Method 3015A**
Alkalinity	Alkalinity is a measure of the acid neutralising capacity of waters. It is a measure of how much acid (H^+) is required to lower the pH to a specific level. In most waters, alkalinity is a function of the concentrations of carbonate [CO_3^{2-}], bicarbonate [HCO_3^-] and hydroxyl [OH^-] ions present. For this method it is assumed that other weak inorganic or organic acids, such as silicic, phosphoric and boric acids are absent. Measuring alkalinity is important in determining a stream's ability to neutralise acidic pollutants from rainfall or wastewater. Total alkalinity is affected by environmental factors; rain, acidic sanitisers, addition of fill water and other product applications can all change the alkalinity over time. Most alkalinity in surface water comes from calcium carbonate ($CaCO_3$), being leached from rocks and soil. This process is enhanced if the rocks and soil have been broken up for any reason, such as mining or urban development. Alkalinity is significant in the treatment of wastewater and drinking water because it will influence treatment process such as anaerobic digestion. Water may be unsuitable for use in irrigation if the alkalinity level in the water is higher than the natural level of alkalinity in the soil. This method covers the determination of alkalinity of all types of water. Alkaline ions present in the sample are neutralised by titration with a standard acid solution. Titration to different pH endpoints allows ion speciation to be determined. This method determines alkalinity relative to pre-designated endpoints measured by a pH meter. The end-points designated are pH 8.3 (Phenolphthalein Alkalinity) and pH 4.5 (Total Alkalinity). Titration by colour can also be used	**APHA 2320 B.**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	to analyse alkalinity, refer to APHA Method 2320 B (2.1) for details. Alkalinity is expressed in terms of the amount of calcium carbonate that would need to be dissolved in fresh water to give the same alkalinity. Alkalinity is reported as mg $CaCO_3$/L. The typical range of applicability is 20 – 4000 mg $CaCO_3$/L. Range can be extended with smaller sample volume and/or alternate titrant concentration(s). Analyse samples, standards, blanks, and Quality Control samples by titrimetric assay using in-house method LTM-INO-4250.	
Ammonia in Water	Alkaline phenol and hypochlorite react with ammonia to form indophenol blue that is proportional to the ammonia concentration. The blue colour is intensified with sodium nitroprusside. This method determines ammonia in drinking, surface, and saline waters; domestic and industrial wastes. Analyse samples, standards, blanks, and Quality Control samples by spectrometric assay using in-house method LTM-INO-4200	**APHA 4500-NH3 B, C, D, F, H**
Anions in Water	Bromide; bromate; chloride; chlorite; chlorate; fluoride; iodide; nitrate; nitrite; phosphate; sulfate by ion chromatography (IC). Analyse samples, standards, blanks, and Quality Control samples by IC using in-house method LTM-INO-4270.	**APHA 4110 B**
Anions in Soils	Tests for water-soluble anions on milled air-dry sample are suitable for use on all soils in clarified/filtered1:5 soil/water extracts. Bromide; bromate; chloride; chlorite; chlorate; fluoride; iodide; nitrate; nitrite; phosphate; sulfate by IC. Analyse samples, standards, blanks, and Quality Control samples by IC using in-house method LTM-INO-4270.	**APHA 4110 B**
Biochemical Oxygen Demand (5 days, 20ºC)	The BOD test is an empirical bioassay-type test which measures the dissolved oxygen consumed by microbial life while assimilating and oxidising organic matter in a sample. A waste sample (or dilution) is incubated for five days 20 ºC ± 1 ºC in the dark. Dissolved oxygen is measured before and after incubation using a modified Winkler or oxygen probe method. The reduction in dissolved oxygen during the incubation period yields a measure of BOD. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4010.	**APHA 5210**.

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
Chemical Oxygen Demand (COD)	Most types of organic matter are oxidized by a boiling mixture of chromic and sulfuric acids. A sample is refluxed in strongly acid solution with a known excess of potassium dichromate ($K_2Cr_2O_7$). After digestion, the remaining unreduced $K_2Cr_2O_7$ is titrated with ferrous ammonium sulfate to determine the amount of $K_2Cr_2O_7$ consumed and the oxidisable matter is calculated in terms of oxygen equivalent. Keep ratios of reagent weights, volumes, and strengths constant when sample volumes other than 50 mL are used. The standard 2-h reflux time may be reduced if it has been shown that a shorter period yields the same results. Some samples with very low COD or with highly heterogeneous solids content may need to be analysed in replicate to yield the most reliable data. Results are further enhanced by reacting a maximum quantity of dichromate, provided that some residual dichromate remains. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4220.	**APHA 5220 C**.
Water-soluble chloride **Chloride - 1:5 soil/water extract**	Chloride in soil is extracted in deionised water and the chloride concentration determined by ion chromatography or spectrometric assay. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4170 (IC) or LTM-INO-4090.	**APHA Method 4500-Cl Rayment & Higginson 1992, "Australian Laboratory Handbook of Soil and Water Chemical. Methods". NEPM 2013 - Schedule B3 - Guideline on Laboratory Analysis of Potentially Contaminated Soil**
Chromium - hexavalent	This procedure measures only hexavalent chromium, (Cr6+). The hexavalent chromium is determined colorimetrically by reaction with diphenylcarbazide in acid solution. A red-violet coloured complex of unknown composition is produced. The colorimetric method is useful for the determination of hexavalent chromium in a natural or treated water in the range from 0.005 to 1 mg/L. This range can be extended by appropriate sample dilution or concentration and/or use of longer cell paths. Normal level analyses in waters uses in-house LTM- INO-4100 Analysis of hexavalent chromium in water by discrete analyser.	**APHA Standard Methods for the Examination of Water & Wastewater. 24th Edition 2023. 3500-Cr-B**
Colour - Visual Comparison Method	Colour is determined by visual comparison of the sample with known concentrations of coloured solutions. Comparison also may be made with special, properly calibrated glass colour disks. The platinum-cobalt method of measuring colour is the standard	**APHA 2120 B**.

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	method, the unit of colour being that produced by 1 mg platinum/L in the form of the chloroplatinate ion. The ratio of cobalt to platinum given (2120B.4) matches the colour of natural waters.	
Cyanide	**Free Cyanide (CN_F)** Only hydrogen cyanide and the cyanide ion in solution can be classed as "free" cyanide. The proportions of HCN and CN- in solution are according to their equilibrium equation; this is influenced by the solution pH. Methods used to detect free cyanide should not alter the stability of weaker cyanide complexes, as they may otherwise be included in the free cyanide result. Methods used to detect free cyanide should be clear of interferences due to the presence of high concentrations of more stable cyanide complexes or other cyanide forms. If not, the interference must be quantified and allowed for in the result. **Weak Acid Dissociable Cyanide (CN_{WAD})** Unlike the definition of "free cyanide" which identifies the specific cyanide species being measured, WAD cyanide refers to those cyanide species measured by specific analytical techniques. WAD cyanide includes those cyanide species liberated at moderate pH of 4.5 such as HCN (aq) and CN-, the majority of Cu, Cd, Ni, Zn, Ag complexes and others with similar low dissociation constants. Methods used to measure WAD should be free from interferences due to the presence of high concentrations of more stable cyanide complexes or other cyanide forms. If not, the interference must be quantified and allowed for in the result. **Total Cyanide (CN_T)** This measurement of cyanide includes all free cyanide, all dissociable cyanide complexes and all strong metal cyanide including ferro-cyanide $Fe(CN)_6^{4-}$, ferri-cyanide $Fe(CN)_6^{3-}$, and portions of hexacyano cobaltate $Co(CN)_6^{3-}$, and those of gold and platinum. Only the related or derived compounds cyanate (CNO^-) and thiocyanate (SCN^-) are excluded from the definition of total cyanide. Methods used to determine total cyanide must be shown to be capable of quantitatively determining all stable complexes of cyanide, including the cobalt cyanide complex. If methods determine other analytes as well (e.g. include SCN^-), those analytes need to be determined separately and allowed for in the total result.	**APHA 4500-CN B, C, D, E, I, N, O and USEPASW 846 9010, 9013, 9014, 9213**.

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4020.	
Electrical Conductivity	This in-house method will determine the concentration of ions in a soil-water suspension, expressed in µS/cm units. The conductivity is measured electrometrically at constant temperature (e.g., 25 ^{o}C). Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4030.	**NEPM Schedule B3**
Ferrous (Fe^{2+})	Iron is brought into solution, reduced to the ferrous state by boiling with acid and hydroxylamine, and treated with 1,10-phenanthroline at pH 3.2 to 3.3. Three molecules of phenanthroline chelate each atom of ferrous iron to form an orange-red complex. The coloured solution obeys Beer's law; its intensity is independent of pH from 3 to 9. A pH between 2.9 and 3.5 insures rapid colour development in the presence of an excess of phenanthroline. Colour standards are stable for at least 6 months. Ferrous iron by DA using in-house LTM-INO-4190.	**APHA 3500-Fe B Phenanthroline Method**
Fluoride in Water	A small volume of sample, typically 50-100 uL, is introduced into an ion chromatograph. The anions of interest are separated and measured, using a system comprised of a guard column, separator column, suppressor device, and conductivity detector. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4270.	**APHA 4500-F^-**
Fluoride in Soils	To determine the total amounts of fluorine in samples, combustion ion chromatography (CIC) can be used, which is a sensitive and versatile method. CIC involves oxidative pyrolysis of samples at high temperatures, followed by absorption of combustion by-products and injection into an IC instrument for analysis. This method is applied to various solid matrices, including soils, sediments, semisolids & biosolids, liquids such as AFFF. For soil samples they are directly combusted and the fluoride absorbed determined by IC. Total fluoride by combustion ion chromatography (CIC) using in-house LTM-INO-4150 (Part A)	**ASTM D7359 Standard Test Method for Total Fluorine, Chlorine and Sulfur in Aromatic Hydrocarbons and Their Mixtures by Oxidative Pyrohydrolytic Combustion followed by Ion Chromatography Detection (Combustion Ion Chromatography-CIC).**
Methylene blue active substances (MBAS)	Methylene blue active substances (MBAS) bring about the transfer of methylene blue, a cationic dye, from an aqueous solution into an immiscible organic liquid upon equilibration. This occurs through ion pair formation by the MBAS anion and the methylene blue cation. The intensity of the resulting blue colour in the organic phase is a measure of MBAS. Anionic	**APHA 5540 C**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	surfactants are among the most prominent of many substances, natural and synthetic, showing methylene blue activity. The MBAS method is useful for estimating the anionic surfactant content of waters and wastewaters, but the possible presence of other types of MBAS always must be kept in mind. This method is relatively simple and precise. It comprises three successive extractions from acid aqueous medium containing excess methylene blue into chloroform (CHCl3), followed by an aqueous backwash and measurement of the blue colour in the CHCl3 by spectrophotometry at 652 nm using in-house LTM-INO-4030 MBAS as MW: 288 (filtered).	
Nitrite, Total Oxidised Nitrogen (NOx) and Nitrate with photometric detection using Discrete Analyser	A discrete analysis is a system of quantitative spectrophotometric determinations utilising automated analytical techniques to perform chemical reactions with high precision and reliability. The sample and the reagents are pipetted by the instrument into a cell and mixed before incubation. After incubation, the absorbance of the solution is measured at the wavelength applicable to the determination. Nitrite (NO_2^-) is determined through formation of a reddish purple azo dye produced at pH 2.0 to 2.5 by coupling diazotised sulfanilamide with N-(1-naphthyl)-ethylenediamine dihydrochloride (NED dihydrochloride). The colour system is measured at 540 nm. The applicable detection range of spectrophotometric measurements, as dictated by Beer's Law, is 10 to 1000 µg NO_2^- N/L, with Method Detection Limits (MDL) and Limits of Reporting (LOR) determined within this range. See method validation for detail. Higher concentrations can be determined by diluting the sample. Total Oxidised Nitrogen (NO_X) Nitrate (NO_3^-) is reduced to nitrite (NO_2^-) by vanadium chloride. The total nitrite ions (Total Oxidised Nitrogen, TON, NO_X) are then reacted with sulphanilamide and NED dihydrochloride under acidic conditions to form a pink azo-dye. The absorbance is measured at 540 nm and is related to the NO_X concentration by means of a calibration curve. Higher concentrations can be determined by diluting the sample. Nitrate concentration is a result of the calculation of NOX concentration minus nitrite concentration noting the need for standardised units (i.e. mg N/L, or "as N"). Nitrogen-nitrate, nitrite, oxides of nitrogen, total by DA using in-house LTM-INO-4350	**APHA 4500-NO_3^-**
Oil and Grease	This method is for determination of n-hexane extractable material (HEM; oil and grease) and n-	**USEPA Method 1664, Revision A**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	hexane extractable material that is not adsorbed by silica gel (SGT-HEM; non-polar material) in surface and saline waters and industrial and domestic aqueous wastes. Extractable materials that may be determined are relatively non-volatile hydrocarbons, vegetable oils, animal fats, waxes, soaps, greases, and related materials. The method is based on prior United States Environmental Protection Agency (US EPA) methods for determination of "oil and grease" and "total petroleum hydrocarbons". The term "n-hexane extractable material" reflects that this method can used to determine materials other than oils and greases. Similarly, the term "silica gel treated n-hexane extractable material" reflects that this method can be used to determine material that is not adsorbed by silica gel (non-polar material). This method is not applicable to measurement of materials that volatilise at temperatures below approximately 85°C. Petroleum fuels from gasoline through #2 fuel oil may be partially lost in the solvent removal operation. Some crude oils and heavy fuel oils contain a significant percentage of materials that are not soluble in n-hexane. Accordingly, recoveries of these materials may be low. This method is capable of measuring HEM and SGT-HEM in the range of 10 to 1000 mg/L, and may be extended to higher levels by analysis of a smaller sample volume collected separately. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4180.	**n-Hexane Extractable Material (HEM; Oil and Grease) and Silica Gel Treated n-Hexane Extractable Material (SGT-HEM; Non-polar Material) by Extraction and Gravimetry**
% Organic Matter	Gravimetric determination based on ashing at >600 °C	**NEPM Schedule B3**
pH in Waters	This in-house method will determine the concentration of hydrogen ions (H+) in a water, expressed in pH units. The pH is measured electrometrically at constant temperature (e.g. 25°C). LTM-GEN-7090 pH electrometric measurement in water matrices by ISE.	**APHA 4500-H^+**
pH in Soils (1:5 aqueous extract) **pH in Soils (1:5 $CaCl_2$ extract)**	This in-house method will determine the concentration of hydrogen ions (H+) in a soil-water or soil-calcium chloride suspension, expressed in pH units. The pH is measured electrometrically at constant temperature (e.g. 25°C). LTM-GEN-7090_R0 pH electrometric measurement in water & soil-type matrices by ISE.	**NEPM Schedule B3**
Phosphorus	Phosphorus analyses embody two general procedural steps: (a) conversion of the phosphorus form of interest to dissolved orthophosphate, and (b) colorimetric determination of dissolved orthophosphate. The separation of phosphorus into its various forms is	**APHA 4500 P**.

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	defined analytically but the analytical differentiations have been selected so that they may be used for interpretive purposes. Filtration through a 0.45-µm-pore-diam membrane filter separates dissolved from suspended forms of phosphorus. No claim is made that filtration through 0.45-µm filters is a true separation of suspended and dissolved forms of phosphorus; it is merely a convenient and replicable analytical technique designed to make a gross separation. Pre-filtration through a glass fibre filter may be used to increase the filtration rate. Phosphates that respond to colorimetric tests without preliminary hydrolysis or oxidative digestion of the sample are termed “reactive phosphorus.” While reactive phosphorus is largely a measure of orthophosphate, a small fraction of any condensed phosphate present usually is hydrolysed unavoidably in the procedure. Reactive phosphorus occurs in both dissolved and suspended forms. Acid hydrolysis at boiling-water temperature converts dissolved and particulate condensed phosphates to dissolved orthophosphate. The hydrolysis unavoidably releases some phosphate from organic compounds, but this may be reduced to a minimum by judicious selection of acid strength and hydrolysis time and temperature. The term “acid-hydrolysable phosphorus” is preferred over “condensed phosphate” for this fraction. The phosphate fractions that are converted to orthophosphate only by oxidation destruction of the organic matter present are considered “organic” or “organically bound” phosphorus. The severity of the oxidation required for this conversion depends on the form—and to some extent on the amount—of the organic phosphorus present. Like reactive phosphorus and acid-hydrolysable phosphorus, organic phosphorus occurs both in the dissolved and suspended fractions. The total phosphorus as well as the dissolved and suspended phosphorus fractions each may be divided analytically into the three chemical types that have been described: reactive, acid hydrolysable, and organic phosphorus. As indicated, determinations usually are conducted only on the unfiltered and filtered samples. Suspended fractions generally are determined by difference; however, they may be determined directly by digestion of the material retained on a glass-fibre filter. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4290.	

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
Silica by Molybdosilicate Metho	Ammonium molybdate at pH approximately 1.2 reacts with silica and any phosphate present to produce heteropoly acids. Oxalic acid is added to destroy the molybdophosphoric acid but not the molybdosilicic acid. Even if phosphate is known to be absent, the addition of oxalic acid is highly desirable and is a mandatory step in both this method and 4500-SiO2 D. The intensity of the yellow colour is proportional to the concentration of "molybdate-reactive" silica. This method is recommended for relatively pure waters containing from 0.4 to 25 mg SiO_2/L. As with most colorimetric methods, the range can be extended by diluting, by concentrating, or by varying the light path. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4300.	**APHA 4500-SiO2 C**
Sulfate (as SO_4^{2-})	Sulfate ion (SO_4^{2-}) is precipitated in an acetic acid medium with barium chloride ($BaCl_2$) so as to form barium sulfate ($BaSO_4$) crystals of uniform size. Light absorbance of the $BaSO_4$ suspension is measured by a photometer and the SO_4^{2-} concentration is determined by comparison of the reading with a standard curve using in-house LTM-INO-4110 Sulfate by Discrete Analyser	**APHA 4500-SO_4^{2-} E. Turbidimetric Method**
Sulfide (as S^{2-})	The methylene blue method is based on the reaction of sulfide, ferric chloride, and dimethyl-p-phenylenediamine to produce methylene blue. Ammonium phosphate is added after colour development to remove ferric chloride colour. The procedure is applicable at sulfide concentrations between 0.1 mg/L and 20.0 mg/L. For samples which are turbid, highly coloured or which contain significant amounts of solids, an impinging procedure is used to separate the sulfide from the sample matrix. This procedure involves acidifying the sample to liberate H2S followed by purging the sample with air into a zinc acetate solution, which captures sulfide as ZnS. This solution is then analysed as the sample. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4011 Determination of sulfide in waters and waste waters	**APHA 4500-S^{2-} D**
Sulfur (Total) - Total Sulfur by Combustion using the LECO Analyser	The LECO 832 Elemental Determinators are specifically designed to determine the sulfur and carbon content in a wide variety of organic materials such as coal, coke and fuel oils, as well as some inorganic materials such as soils, cements and limestone by high temperature combustion with non-dispersive infrared detection (NDIR).	**Total Sulfur by Combustion using the Leco Analyzer, California Department of Food and Agriculture, Revision: 1, Date: 12 May 22**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	Analysis begins as a sample is weighed into a combustion boat and placed into the furnace typically regulated at 1350°C with a pure oxygen environment. The sample combusts, releasing carbon as CO_2 gas with the sulfur forms oxidized and released as SO2 gas. After a pre-set time, additional oxygen is introduced via a ceramic lance directly above the sample to accelerate the combustion of refractory materials. The combustion gases are swept to the back of the furnace and then forward through the inner and outer furnace tubes, allowing the combustion gases to remain in the high temperature zone of the furnace, ensuring efficient oxidation. Upon exiting the furnace, the combustion gases flow through anhydrone tubes removing moisture and on to the flow controller, setting the flow of the combustion gases through the NDIR sulfur and/or carbon detection cells. Non-dispersive infrared cells are based on the principle that CO_2 and SO_2 absorb infrared (IR) energy at unique wavelengths within the IR spectrum. Incident IR energy at these wavelengths is absorbed as the gases pass through IR absorption cells with the absorption being dependent upon the path length of the cell. The Dual Range (DR) sulfur model has a wider sulfur range due to a short and long path length IR cells provided for measurement of high and low range sulfur signals. The software automatically selects which cell to use for optimum measurement. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4440 Determination of Total Sulfur in Soils by LECO Combustion IR.	
Total Organic Carbon in Water	Total Carbon (TC) is measured by injecting a portion of the water sample into a heated combustion tube packed with an oxidation catalyst. The water is vaporised and TC, the organic carbon and the inorganic carbon, is converted to carbon dioxide (CO2). The carbon dioxide is carried with the carrier gas stream from the combustion tube to a NDIR (non-dispersive infrared gas analyser) and concentration of carbon dioxide is measured. The TC concentration of the sample is obtained by using the calibration curve prepared with standard solutions. Inorganic Carbon (IC) is measured by injecting a portion of the sample into an IC reaction chamber filled with phosphoric acid solution. All IC is converted to carbon dioxide and concentration of carbon dioxide is measured with a NDIR.	**APHA 5310 B**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	TOC may be obtained as the difference of TC and IC. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-INO-4060.	
Total Organic Carbon in Soil	Total Carbon (TC) is measured by placing a quantity of the soil sample into a heated combustion tube packed with an oxidation catalyst. The samples are weighed into reusable quartz crucibles and loaded onto the integrated autosampler. The unique vertical "bottom-to-top" sample introduction system lifts the samples into the high-temperature combustion oven. At 1050°C the samples are oxidised and the carbon in the sample is converted into CO_2. The CO_2 is measured by Near Infra Red Detection (NDIR). After complete combustion, the sample crucible is automatically removed from the oven. Any remains of the sample are removed from the combustion zone with the crucible, so no ash build-up will occur, which reduces maintenance on the instrument. Combining a high-temperature combustion oven with a sensitive dual-range Infrared detector allows the instrument to analyse samples containing carbon concentrations from low ppm levels to % levels. The TC concentration of the sample is obtained by using the calibration curve prepared with standard solutions. Inorganic Carbon (IC) is measured by treating the sample into an IC reaction chamber filled with an acid solution. All IC is converted to carbon dioxide and concentration of carbon dioxide is measured with a NDIR. TOC may be obtained as the difference between TC and IC. Analyse samples, standards, blanks, and Quality Control samples using the in-house method LTM-INO-4060.	**ISO 10694:1996(MAIN)** **Soil quality - Determination of organic and total carbon after dry combustion (elementary analysis)** **EN 13639:2017(MAIN)** **Determination of total organic carbon in limestone**
Total Dissolved Solids (TDS) Dried at 180°C	A well-mixed sample is filtered through a standard glass fibre filter. The filtrate is evaporated and dried to constant weight at 180ºC. This method determines filterable residue in drinking, surface, and saline waters; domestic and industrial wastes. (A) Mineral Waters: Highly mineralized waters containing significant concentrations of calcium, magnesium, chloride and/or sulfate may be hygroscopic and will require prolonged drying, desiccation and rapid weighing. (B) Bicarbonate: Samples containing high concentrations of bicarbonate will require careful and possibly prolonged drying at 180oC to insure that all the bicarbonate is converted to carbonate. (C) High Residue Levels: Too much residue in the evaporating dish will crust over and entrap water that will not be driven off during drying. Total residue	**APHA 2540 C.**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	should be limited to about 200 mg. in-house method LTM-INO-4170.	
Total Suspended Solids (TSS) Dried at 103°C – 105°C	Suspended solids are those that are retained on a glass-fibre filter. The unfiltered sample is mixed thoroughly and an appropriate volume is rapidly poured into a graduated cylinder. The suspended solids are collected on a glass fibre filter, and the insoluble residue is dried at 104 ± 1°C and weighed. This method may be used to determine the suspended-solids concentration of any natural or treated water or industrial waste. In-house method LTM-INO-4070	**APHA 2540 D.**
Turbidity	This method is based on a comparison of the intensity of light scattered by the sample under defined conditions with the intensity of light scattered by a standard reference suspension under the same conditions. The higher the intensity of scattered light, the higher the turbidity. Formazin polymer is used as the primary standard reference suspension. In-house method LTM-INO- 4140.	**APHA 2130**
Fixed and Volatile Solids Ignited at 550°C	The residue from LTM-INO-4070 or LTM-INO-4170 is ignited to constant weight at 550°C. The remaining solids represent the fixed total, dissolved, or suspended solids while the weight lost on ignition is the volatile solids. The determination is useful in control of wastewater treatment plant operation because it offers a rough approximation of the amount of organic matter present in the solid fraction of wastewater, activated sludge, and industrial wastes.	**APHA 2540 E.**
Residue, Volatile (Gravimetric, Ignition at 550°C)	The residue obtained from the determination of total, filterable or non-filterable residue is ignited at 550°C in a muffle furnace. The loss of weight on ignition is reported as mg/L volatile residue. This method determines the weight of solid material combustible at 550°C. The test is useful in obtaining a rough approximation of the amount of organic matter present in the solid fraction of sewage, activated sludge, industrial wastes, or bottom sediments.	**APHA 2540 E.**
General		
Cation exchange capacity (CEC)	Cation exchange capacity (CEC) is a measure of the soil's ability to hold positively charged ions. It is a very important soil property influencing soil structure stability, nutrient availability, soil pH and the soil's reaction to fertilisers and other ameliorants (Hazleton and Murphy 2007). The clay mineral and organic matter components of soil have negatively charged sites on their surfaces which adsorb and hold positively charged ions (cations) by electrostatic force. This electrical charge is	**NEPM** **Schedule B3**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	critical to the supply of nutrients to plants because many nutrients exist as cations (e.g. magnesium, potassium and calcium). In general terms, soils with large quantities of negative charge are more fertile because they retain more cations (McKenzie et al. 2004) however, productive crops and pastures can be grown on low CEC soils. The main ions associated with CEC in soils are the exchangeable cations calcium (Ca^{2+}), magnesium (Mg^{2+}), sodium (Na^{+}) and potassium (K^{+}) (Rayment and Higginson 1992), and are generally referred to as the base cations. In most cases, summing the analysed base cations gives an adequate measure of CEC ("CEC by bases"). However, as soils become more acidic these cations are replaced by H^{+}, Al^{3+} and Mn^{2+}, and common methods will produce CEC values much higher than what occurs in the field (McKenzie et al. 2004). NOTE: Only CEC & ESP are calculated by this method. Conducted by in-house Method LTM-MET-3060 – Cation Exchange Capacity (CEC) by bases & Exchangeable Sodium Percentage (ESP).	
Clay Content	This method is based on the Soil Classification assessment by Hydrometer outlined in the Australian Standard 1289.3.6.3 (Determination of the particle size distribution of a soil – Standard method of fine analysis using a hydrometer). This method quantitatively determines the physical proportions of three sizes of primary soil particles, by determining their settling rates in an aqueous solution using a hydrometer. The three categories of particles measured are defined as follows:- 1. Sand Ranges from 2000 µm to 50 µm 2. Silt Ranges from 50 µm – 2 µm 3. Clay Less than 2 µm Settling rates of primary soil particles are measured using a hydrometer.	**AS1289.3.6.3**
Moisture	Gravimetric determination based on drying at 105-110 °C. MOISTURE CONTENT IN SOIL OR OTHER SOLID MATRICES BY GRAVIMETRY LTM-GEN-7080 Moisture.	**NEPM** **Schedule B3**
Leaching Procedures	This in-house method is for the preparation of leachates collected from soil, sediments, sludges, and other solid matrices using a rotary vessel extraction procedure. The method allows for the substitution of laboratory grade de-ionised water, EP or SPLP fluids, or site water supplied by the client as the extraction fluid. The solid portion of the sample is reduced in particle size, if necessary, and leached by rotary vessel agitation with a selected leaching fluid. The sample leachate is then extracted/ analysed by an	**Toxicity Characteristic Leaching Procedure (TCLP) USEPA Method 1311** **Australian Standard Leaching Procedure (ASLP) AS 4439.2: 2019; AS4439.3: 2019**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	additional test method, as per client request. (Eurofins in-house method LEACHING PROCEDURE FOR VOLATILE AND NON-VOLATILE ANALYTES FROM SOILS AND SOLID WASTES LTM-GEN-7010.	
LEAF 1313	Liquid –Solid Partitioning as a Function of Extract pH for Constituents in Solid Materials using a Parallel Batch Extraction Procedure. Nine (9) Parallel extractions of a particle sized reduced solid material in dilute acid or base and reagent water. Series of eluates having pH values ranging from 2-13. Liquid solid ratio of 10:1. Eluate is centrifuged and filtered for COPCs. Designed to provide aqueous extracts representing the liquid-solid portioning [LSP] curve as a function of pH for inorganics and non-volatile organics in solid materials	**EPA SW-846 Method 1313**
LEAF 1314	Liquid –Solid Partitioning as a Function of Liquid-Solid Ration for Constituents in Solid Materials using an Up-Flow Percolation Column Procedure Eluent is introduced into a column with packed particle sized reduced solid material in an up-flow pumping mode. Flow rate is maintained between 0.5-1.0 LS/Day. Eluent is collected at predetermined times, filtered and analysed for COPCs. Total time of test is approximately 14 days. Designed to provide the liquid – solid portioning [LSP] of inorganic constituents and non-volatile organics in granular solid material as a function of liquid to solid [LS] ratio under percolation conditions.	**EPA SW-846 Method 1314**
LEAF 1315	Mass Transfer Rates of Constituents in Monolithic or Compacted Granular Materials using a Semi-dynamic Tank Leaching Procedure. Leaching of continuously water saturated monolithic or compacted granular material in an eluent-filled tank with periodic renewal of the leaching solution. LS ratio of 9 mL eluent per cm2 of surface area. Eluent is collected at predetermined times and analysed for COPCs. Eluate is centrifuged and filtered for COPCs. Total time of test is 63 days. Designed to provide the mass transfer [release rates] of inorganic analytes contained in a monolith or compacted granular material. Under diffusion controlled release conditions, as a function of leaching time.	**EPA SW-846 Method 1315**
LEAF 1316	Liquid- Solid Partitioning as a Function of Liquid-Solid Ratio for Constituents in Solid Materials using a Parallel Batch Extraction Procedure. Five (5) Parallel extractions of a particle-size reduced solid material in reagent water over a range of L/S	**EPA SW-846 Method 1316**

Analyte	Method Summary	Reference Method
Inorganics Laboratory		
	values from 0.5 to 10 mL eluant/g dry material. Depending on particle size, sample is tumbled between 24 and 72 hours. Eluate is centrifuged and filtered for COPCs. Designed to provide the liquid-solid portioning{LSP] of inorganic and non-volatile organics at the natural pH of the solid material as a function of liquid to solid ratio [L/S] under conditions that approach liquid-solid chemical equilibrium.	

Analyte	Method Summary	Reference Method
Asbestos Laboratory		
Asbestos in Soils	The whole sample submitted is first dried and then sieved through a 10 mm sieve followed by a 2 mm sieve. All fibrous matter viz greater than 10mm, greater than 2 mm as well as the material passing through the 2 mm sieve are retained and analysed for the presence of asbestos. If the sub 2 mm fraction is greater than approximately 30 g to 60 g then a sub-sampling routine based on ISO 3082:2017(E) *Iron ores - Sampling and Sample Preparation Procedures* is employed. Depending on the nature and size of the soil sample, the sub-2 mm residue material may need to be sub-sampled for trace analysis in accordance with AS 5370:2024. Conducted in accordance with the Australian Standard AS 5370:2024: Method for the Qualitative Identification of Asbestos in Bulk Samples and in-house Method LTM-ASB-8020 by polarised light microscopy (PLM) and dispersion staining (DS) techniques. Bulk samples include building materials, soils and ores	***Australian Standard AS 5370:2024 Sampling and qualitative identification of asbestos in bulk materials (ISO 22262-1:2012, MOD),*** *formerly* ***AS 4964-2004***
Bonded asbestos-containing material (ACM)	The material is first examined and any fibres isolated and where required interfering organic fibres or matter may be removed by treating the sample for several hours at a temperature not exceeding 400°C ± 30°C. The resultant material is then ground and examined in accordance with AS 5370:2024and ores	***Australian Standard AS 5370:2024 Sampling and qualitative identification of asbestos in bulk materials (ISO 22262-1:2012, MOD),*** *formerly* ***AS 4964-2004***
Asbestos fibres in Air	Conducted in accordance with the National Occupational Health & Safety Commission - Guidance Note on The Membrane Filter Method for Estimating Airborne Asbestos Fibres 2nd Edition [NOHSC:3003(2005)] and in-house Method LTM-ASB-8010.	**National Occupational Health and Safety Commission Guidance Note on the Membrane Filter Method for Estimating Airborne Asbestos Fibres NOHSC:3003**

Air		
Filters - Total Metals (As, Cd, Cr, Cu, Ni, Pb, Zn)	The filter is digested in a hot block set to 90°C - 95°C for 2.5 hours using an extraction fluid containing hydrochloric acid (HCl) and nitric acid (HNO_3). Two aliquots of hydrogen peroxide (H_2O_2) are added after 1.5 hours and 2.0 hours of extraction and are allowed to effervesce. After extraction, the samples are filtered and diluted to a final volume of 50 mL. The extract is analysed by ICP-MS, and the data are collected using the manufacturer's software. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-MET-3040.	**EQL-0512-201 - US Environmental Protection Agency**

Analyte	Method Summary	Reference Method
Miscellaneous		
Foreign Materials	This test method sets out the procedure for the determination of the foreign materials content in a sample of recycled crushed concrete. The sample submitted for testing shall be reduced, as required, by quartering or riffling to provide a subsample of at least 6 kg. **Procedure** (a) The test portion shall be dried to constant mass in an oven at a temperature within the range of 50°C to 60°C. Record the mass of the portion. (b) The dried test portion shall be let to cool down to ambient temperature. (c) The test portion shall be divided into lots in order to avoid overloading of the test sieve. The size of the lot shall be such that after sieving, the mass retained on the 4.75 mm AS sieve shall not exceed the permissible mass specified in the following table for either a 200 mm, 300 mm or 450 mm diameter test sieve. (d) Each lot shall be sieved for not less than two minutes. Continue agitation until no more than 1 per cent of the residue passes the sieve. (e) On completion of sieving, weigh the material retained and record the mass. The mass of material from each lot retained shall be added together and considered as a single size increment. (f) Sort out by hand all foreign material retained and classify it in accordance with the following classifications: **Type I:** Metal, Glass, Asphalt, Stone, Ceramics and Slag (other than blast furnace slag) **Type II:** Plaster, Clay lumps and other Friable Material **Type III:** Rubber, Plastic, Bitumen, Paper, Cloth, Paint, Wood and other Vegetable Matter	**RMS Test method T276** **Foreign materials content of recycled** **crushed concrete** **OCTOBER 2012**

Analyte	Method Summary	Reference Method
Mycology Laboratory		
Analysis of Zefon Bio-Tapes™ **LTM-MLD-5010**	This test method uses optical microscopy for the detection, semi-quantification, and identification of fungal structures in tape lift preparations. This test method describes the preparation techniques for tape-lift matrices, the procedure for confirming the presence of fungal structures, and the reporting of observed fungal structures The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.	**ASTM D7658 – 17 Standard Test Method for Direct Microscopy of Fungal Structures from Tape**
Analysis of Impaction Air Sampling Cassettes **LTM-MLD-5020**	This test method is a procedure that uses direct microscopy to analyse the deposit on an inertial impaction sample. This test method describes procedures for categorising and enumerating fungal structures by morphological type. Typically, categories may be as small as genus (for example, *Cladosporium*) or as large as phylum (for example, basidiospores). This method contains two procedures for enumerating fungal structures: one for slit impaction samples and one for circular impaction samples. This test method is applicable for impaction air samples, for which a known volume of air (at a rate as recommended by the manufacturer) has been drawn, and is also applicable for blank impaction samples. Enumeration results are presented in fungal structures/sample (fs/sample) and fungal structures/m^3 (fs/m^3). The range of enumeration results that can be determined with this method depends on the size of the spores on the sample trace, the amount of particulate matter on the sample trace, the percentage of the sample trace counted, and the volume of air sampled. This method addresses only the analysis of samples. The sampling process and interpretation of results is outside the scope of this method. The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.	**D7391 – 09** **Standard Test Method for Categorization and Quantification of Airborne Fungal Structures in an Inertial Impaction Sample by Optical Microscopy**

Analyte	Method Summary	Reference Method
Air Toxics Laboratory		
TRH by Modified US EPA TO-15*	The laboratory performed analysis following modified EPA TO-15 for Total Recoverable Hydrocarbon (TRH) fractions using electron ionisation GC/MS in full scan mode. The method involves concentrating up to 0.2 litres of air. The concentrated aliquot is then flash vaporised and swept through a water management system to remove water vapour. Following dehumidification, the sample passes directly into the GC-MS for analysis. All sample-related peaks including BTEX and naphthalene eluting within their respective carbon range are included in the TRH result. The >C6-C10 TRH range is defined as the total ion area of peaks eluting after n-Hexane and including n-Decane referenced to the response factor of Toluene. The >C10-C12 TRH range is defined as the total area of peaks eluting after n-Decane and including n-Dodecane and reference to the response factor of n-Decane. Hydrocarbons heavier than C12 do not reliably recover from summa canisters due to their low vapour pressure. As a result, the reported range was limited to C12 rather than C16 as defined in Table C1[1]. If requested, the fraction >C6-C10 minus BTEX (F1) and >C10-C12 minus naphthalene (modified F2) were reported following the definition listed in the previous paragraph except BTEX and naphthalene peaks were removed from the total ion peak area. Naphthalene elutes outside the >C10-C12 range on the system used for sample analysis. As a result, >C10-C12 TRH value is equivalent to the modified F2 value.	**USEPA Compendium Method TO-15 Determination Of Volatile Organic Compounds (VOCs) in Air Collected in Specially-Prepared Canisters and Analyzed by Gas Chromatography/Mass Spectrometry (GC/MS)**
Modified US EPA TO-15 & VPH Fractions*	The laboratory performed analysis via EPA Method TO-15 and Eurofins Air Toxics VPH (Volatile Petroleum Hydrocarbon) methods for the Determination of VPH Fractions using GC/MS in the full scan mode. The method involves concentrating up to 0.5 litres of air. The concentrated aliquot is then flash vaporised and swept through a water management system to remove water vapour. Following dehumidification, the sample passes directly into the GC/MS for analysis. This method is designed to measure gaseous	**USEPA Compendium Method TO-15 Determination Of Volatile Organic Compounds (VOCs) in Air Collected in Specially-Prepared Canisters and Analyzed by Gas**

[1] CRC CARE 2013, Petroleum hydrocarbon vapour intrusion assessment: Australian guidance, CRC CARE Technical Report no. 23, CRC for Contamination Assessment and Remediation of the Environment, Adelaide, Australia.

Analyte	Method Summary	Reference Method
Air Toxics Laboratory		
	phase aliphatic and aromatic compounds in ambient air and soil gas collected in stainless steel Summa canisters. Eurofins Air Toxics VPH method is a hybrid of EPA TO-15 method viz chromatographic peaks were identified via mass spectrum as either aliphatic or aromatic petroleum hydrocarbons and included in the appropriate range as defined by the method. The volatile Aliphatic hydrocarbons are collectively quantified within the C5 to C6 range, C6 to C8 range, C8 to C10 range and the C10 to C12 range. Additionally, the volatile Aromatic hydrocarbons are collectively quantified within the C8 to C10 range and the C10 to C12 range. The Aromatic ranges refer to the equivalent carbon (EC) ranges. (Please note that benzene constitutes the >C5-C7 aromatic range and toluene constitutes the >C7-C8 aromatic range. Benzene and toluene concentrations are reported on the TO-15 work order fraction.) Aliphatic data is calculated from the Total Ion Chromatogram (TIC) which has been reprocessed in a duplicate file differentiated from the original by the addition of an alphanumeric extension. The Aromatic calculation also uses the information contained in the associated extracted ion file.	**Chromatography/Mass Spectrometry (GC/MS)**
Modified Natural Gas Analysis by ASTM D-1946*	The laboratory performed analysis via Modified ASTM Method D-1946 for Methane and fixed gases in air using GC/FID or GC/TCD. The method involves direct injection of 1.0 mL of sample. On the analytical column employed for this analysis, Oxygen co-elutes with Argon. The corresponding peak is quantitated as Oxygen.	**ASTM D1946-77** **Standard Method for Analysis of Reformed Gas by Gas Chromatography**
Analysis of volatile and semi-volatile organic compounds in vapor by thermal desorption GC/MS full scan using modified EPA method TO-17, **SOP#109**	The laboratory performed analysis via The laboratory performed the analysis via modified EPA Method TO-17 using GC/MS in the full scan mode. TO-17 'VI' sorbent tubes are thermally desorbed onto a secondary trap. The trap is thermally desorbed to elute the components into the GC/MS system for compound separation and detection. A modification that may be applied to EPA Method TO-17 at the client's discretion is the requirement to transport sorbent tubes at 4 degrees C. Laboratory studies demonstrate a high level of stability for VOCs on the TO-17 'VI' tube at room temperature for periods of up to 14 days. Tubes can be shipped to and from	**Modified EPA Method TO-17 (VI Tubes)***

<table>
<tr><th>Analyte</th><th>Method Summary</th><th>Reference Method</th></tr>
<tr><td colspan="3">Air Toxics Laboratory</td></tr>
<tr><td></td><td>
the field site at ambient conditions as long as the 14-day sample hold time is upheld. Trip blanks and field surrogate spikes are used as additional control measures to monitor recovery and background contribution during tube transport. Since the TO-17 VI application significantly extends the scope of target compounds addressed in EPA Method TO-15 and TO-17, the laboratory has implemented several method modifications outlined in the table below.

Specific project requirements may override the Eurofins Air Toxics modifications.
<table>
<tr><td>Requirement</td><td>TO-17</td><td>Eurofins Air Toxics Modifications</td></tr>
<tr><td>Initial Calibration</td><td>%RSD
≤ 30% with 2 allowed out up to 40%</td><td>VOC list:
%RSD ≤ 30% with 2 allowed out up to 40%</td></tr>
<tr><td>SVOC list: %RSD</=30% with 2 allowed out up to 40%</td><td></td><td></td></tr>
<tr><td>Daily Calibration</td><td>%D for each target compound within ± 30%.</td><td>Fluorene, Phenanthrene, Anthracene, Fluoranthene, and Pyrene within ± 40%D</td></tr>
<tr><td>Audit Accuracy</td><td>70-130%</td><td>Second source recovery limits for Fluorene, Phenanthrene, Anthracene, Fluoranthene, and Pyrene = 60-140%.</td></tr>
<tr><td>Distributed Volume Pairs</td><td>Collection of distributed volume pairs required for monitoring ambient air to insure high quality.</td><td>If site is well-characterised or performance previously verified, single tube sampling may be appropriate. Distributed pairs may be impractical for soil gas collection due to configuration and volume constraints.</td></tr>
<tr><td>Analytical Precision</td><td>≤ 20% RPD</td><td>≤ 30% RPD for Fluorene,</td></tr>
</table>
</td><td></td></tr>
</table>

Analyte	Method Summary	Reference Method
Air Toxics Laboratory		
	Phenanthrene, Anthracene, Fluoranthene, and Pyrene.	
Volatile organic compounds (VOCs) - chemically desorbed with CS_2	Code RAD130 cartridge is a stainless steel net cylinder, with 100 mesh grid opening and 5.8 mm diameter, packed with 530 ± 30 mg of activated charcoal, particle size is 35-50 mesh. Volatile organic compounds are trapped by adsorption and recovered by carbon disulfide desorption, analysis is performed by GC-MS. white diffusive body code RAD120  supporting plate code RAD121  vertical adapter code RAD122 (optional)  Chemi-adsorbing cartridge code RAD130  Extraction: A volume of 2.0 mL of CS_2 and 100 µL of internal standard solution is added directly in the radiello glass tube. The tube is shaken gently for 30 minutes. Sampling rates varies from the value at 298 K on the effect of temperature (in Kelvin) as expressed by the following equation: $$Q_K = Q_{298}(\frac{K}{298})$$ where Q_K is the sampling rate at the temperature K and Q_{298} is the reference value at 298 K. This produces a variation of ±5% for 10 °C variation (upwards or downwards) from 25 °C. Sampling rate is invariant with humidity in the range 15 - 90% and with wind speed between 0.1 and 10 $m.s^{-1}$. NOTE: where uptake rates (Q_K) are unpublished then they have been estimated from like compounds. Results for these compounds are semi-quantitative.	**Passive Sampler – radiello® User Manual 2019**

Analyte	Method Summary	Reference Method
Air Toxics Laboratory		
	Average concentration (in µg.m^{-3}) over the whole exposure time is calculated according to the following expression: $$C\ (\mu g.m^{-3}) = \frac{m\ (\mu g)}{Q_K\ (mL.min^{-1}).t(min)}.10^6$$ m = mass of analyte in µg determined by GC-MS t = exposure time in minutes	
Volatile organic compounds (VOCs) - thermally desorbed	Code RAD145 is a stainless steel net cylinder, with 3 x 8 µm mesh opening and 4.8 mm diameter, packed with 350 ± 10 mg of graphitised charcoal (Carbograph 4), particle size is 35-50 mesh. Volatile organic compounds are trapped by adsorption and recovered by thermal desorption, analysis is performed by GC-MS. yellow diffusive body code RAD1202  supporting plate code RAD121  vertical adapter code RAD122 (optional)  Chemi-adsorbing cartridge code RAD145  Code RAD145 cartridge has been dimensioned to fit the diameter of the Markes Unity thermal desorption system that is used in conjunction with an Agilent GC-MS. Sampling rates varies from the value at 298 K on the effect of temperature (in Kelvin) as expressed by the following equation: $$Q_K = Q_{298}(\frac{K}{298})$$ where Q_K is the sampling rate at the temperature K and Q_{298} is the reference value at 298 K. This produces a variation of ±5% for	**Passive Sampler – radiello® User Manual 2019**

Analyte	Method Summary	Reference Method
Air Toxics Laboratory		
	10 °C variation (upwards or downwards) from 25 °C. Sampling rate is invariant with humidity in the range 15 - 90% and with wind speed between 0.1 and 10 $m.s^{-1}$. Do not expose directly radiello to rain: even if small amounts of water are adsorbed by Carbograph 4, they can nevertheless interfere with analysis. NOTE: where uptake rates (Q_K) are unpublished then they have been estimated from like compounds. Results for these compounds are semi-quantitative. Average concentration (in $\mu g.m^{-3}$) over the whole exposure time is calculated according to the following expression: $$C\ (\mu g.m^{-3}) = \frac{m\ (\mu g)}{Q_K\ (mL.min^{-1}).t(min)}.10^6$$ m = mass of analyte in µg determined by GC-MS t = exposure time in minutes	
Volatile organic compounds (VOCs) – passive samplers	Companion EPA Methods 325A (Sampler Deployment and VOC Sample Collection) and 325B (Sampler Preparation and Laboratory Analysis) select benzene as the representative compound to evaluate the overall emissions from refineries. Passive sampling onto sorbent tubes followed by Thermal Desorption-Gas-Chromatography/Mass Spectrometry (TD-GC/MS) analysis has been established as the standard air monitoring technology for the EPA's new rule. Passive sampling tube shelter assemblies will be hung at various locations along the fence line/property boundary surrounding refineries. After two weeks (14 days) passive sampling tubes can be detached from their shelters, re-sealed and sent to a laboratory equipped with TD-GC/MS for analysis. Per EPA Method 325, all tubes must be replaced with freshly conditioned and qualified sampling tubes every 14 days to ensure continuous monitoring. The methods provide a low cost alternative to screen fugitive or area emissions as compared to active sampling methods that involve pumped sorbent tubes or time weighted average canister sampling.	**US EPAMethod 325B—Volatile Organic Compounds from Fugitive and Area Sources:** **Sampler Preparation and Analysis**

Analyte	Method Summary	Reference Method
Air Toxics Laboratory		
	While the rule is currently limited to the monitoring of benzene, Method 325 can also be extended to include other compounds of concern at ambient monitoring sites. Additional target VOCs include 1,3-Butadiene, Toluene, Ethyl Benzene, and Xylenes as well as other chemicals for which diffusive sampling rates have been determined. Reporting limits less than 1 µg/m^3 can be easily achieved over a 7-day period. Extending the sampling period to 14 days translates to reporting limits less than 0.5 µg/m^3.	

Analyte	Method Summary	Reference Method
Hygiene Laboratory		
Respirable Crystalline Silica	This in-house method will determine the mass of respirable crystalline silica (RCS) present on 25 mm or 37 mm filter papers, using X-Ray diffraction, when using an analytical approach where the dust on the air sample filter is directly analysed by the instrument. If the dust deposit of sample on the filter is too deep the X-ray radiation might not penetrate into the whole sample and the radiation may also be absorbed by the sample's matrix, then digestion using tetrahydrofuran is employed. The suspended particulate is deposited on a silver membrane filter and then analysed by the instrument. The expression RCS includes the most common polymorphs quartz and cristobalite. The less common polymorphs of crystalline silica, such as tridymite, are not included within the scope of this method because a standard reference material is not available. Both alpha-quartz and cristobalite are reported and are expressed in µg. The limit of reporting is 5 µg. When the volume of air sampled is known the concentration of RCS in the air is readily calculated and expressed as mg/m^3. This method is restricted to use by, or under the supervision of, analysts trained in the use of X-Ray diffractometers. Analyse samples, standards, blanks, and Quality Control samples using in-house method LTM-HYG-0001.	**NIOSH 7500 Manual of Analytical Methods (NMAM), Fourth Edition. SILICA, CRYSTALLINE, by XRD (filter redeposition).**
Diesel Particulate Matter as Elemental Carbon	Respirable particulates are collected onto a quartz filter. A standard sized punch is taken from the exposed filter and placed into a quartz oven. The oven is purged with helium and a stepped temperature ramp increases the oven temperature thermally desorbing organic compounds into an oxidising oven. The thermally desorbed compounds flow through a MnO_2 oxidising oven and quantitatively converted to CO_2. This is mixed with hydrogen gas and flows through a heated nickel catalyst to be quantitatively converted to methane where it is subsequently measured using a flame ionisation detector FID detector. The oven is then cooled and the flow stream is switched to an oxidising helium /oxygen carrier gas. A second temperature ramp is used and any elemental carbon is then	**NIOSH 5040 Manual of Analytical Methods (NMAM), Fourth Edition. DIESEL PARTICULATE MATTER. 5040. (as Elemental Carbon).**

Analyte	Method Summary	Reference Method
Hygiene Laboratory		
	detected in the same manner. The final stage of the analysis is the fixed volume loop which injects a gas standard at the end of the analysis. All peak areas are compared to the standard for each analysis to normalise variations in instrument performance. Analyse samples, standards, blanks, and Quality Control samples using in-house method ARL Method No. 161	
Respirable Dust Analysis	Airborne concentrations of respirable dust in the workplace are determined by passing a measured volume of air through a 25 or 37 mm, 5 µm pore size filter of predetermined mass by reweighing the filter at the end of the sampling period, the quantity of material collected is determined by difference. To separate the respirable fraction, a miniature cyclone separator is used prior to the filter. Analyse samples, standards, blanks, and Quality Control samples using in-house method ARL Method No. 098	**AS 2985 Workplace atmospheres - Method for sampling and gravimetric determination of respirable dust**
Inhalable Dust Analysis	Airborne concentrations of dust in the workplace are determined by passing a measured volume of air through a 25 mm filter of predetermined mass by re-weighing the filter at the end of the sampling period, the quantity of material collected is determined by difference. Analyse samples, standards, blanks, and Quality Control samples using in-house method ARL Method No. 097	**AS 3640 Workplace atmospheres - Method for sampling and gravimetric determination of inhalable dust**
Dust Deposition in Ambient Air	This LTM-INO-4160 method sets out a procedure for the sampling of particulate matter that is deposited from the atmosphere, and procedures for the gravimetric determination of the mass deposition rate of insoluble solids, ash, combustible matter, soluble solids and total solids from ambient air. A method to determine deposition rates for metals present in deposited matter is also provided. NOTES: 1 The sample obtained by the sampling procedure specified may be subjected to physical or chemical analysis. 2 The method provides an estimate of the mean surface concentration of deposited matter settling from the air over a sampling period, typically one month. Particulate deposition rates of 0.1 g/m^2 month and above may be determined using a monthly sampling period.	**AS/NZS 3580.10.1:2016 Methods for sampling and analysis of ambient air Determination of particulate matter - Deposited matter - Gravimetric method**

Analyte	Method Summary	Reference Method
Hygiene Laboratory		
	3. A copper sulfate biocide is routinely added but alternatives may be included if metals are to be determined.	

Analyte	Method Summary	Reference Method
Microplastics Laboratory		
Microplastics in water (Environmental/Potable)	Definition: Microplastics in Drinking Water by the California State Water Board 'Microplastics in Drinking Water' are defined as solid polymeric materials to which chemical additives or other substances may have been added, which are particles that have at least three dimensions that are greater than 1 nanometre (nm) and less than 5,000 micrometres (µm). Polymers derived in nature that have not been chemically modified (other than by hydrolysis) are excluded. Vacuum membrane filtration is usually the first step in the process, and the amount of suspended solids present may limit the volume filtered. The volume filtered is reported for each sample. Depending on their complexity, environmental water samples will require several purification steps to remove the natural sample matrix, combining a density separation to remove the bulk mineral fraction and chemical digestion steps to eliminate the remaining organic matter from the sample whilst leaving the microplastics intact. Note: these pre-treatments do not degrade the polymers tested here. The final extract is then passed through a 5 µm membrane filter. The retained particulates are transferred onto an infrared-reflective slide and analysed using the Laser Direct Infrared (LDIR) Chemical Imaging system. The system detects sizes and enumerates the particles present in the sample. Then, an infrared (IR) spectrum is collected (in the IR fingerprint region, 1800 cm^{-1} - 975 cm^{-1}) for each particle detected. The collected spectra are compared to a verified reference library for chemical identification. A minimum match quality of ≥80% is applied for identification. The automated LDIR workflow can analyse particles in the range of 20 µm to 500 µm. For particles between >500 µm and ≤5000 µm, the identification using Attenuated Total Reflection (ATR) is available upon request and is not part of this workflow unless requested. The library used is verified with two independent reference materials for the following polymers: polyethylene (PE), polypropylene (PP), polystyrene (PS), polyvinylchloride (PVC), polyethylene terephthalate (PET), polycarbonate	**SWB-MP1-rev1 Standard Operating Procedures for Extraction and Measurement by Infrared Spectroscopy of Microplastic Particles in Drinking Water by the California State Water Board.**

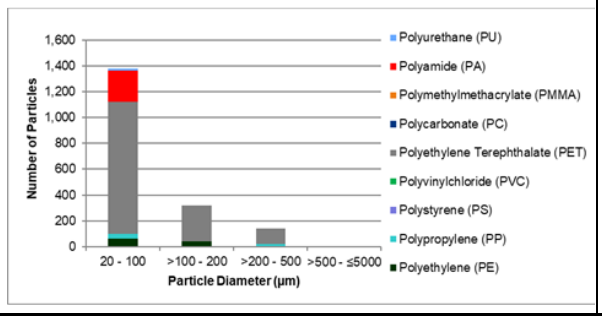
Analyte	Method Summary	Reference Method
Microplastics Laboratory		
	(PC), polymethylmethacrylate (PMMA), polyamide (PA) and polyurethane (PU). Analyse samples, standards, blanks, and Quality Control samples by GC-MS using in-house method LTM-MPS-9020 Microplastics in Environmental Water. 	

Table 3: PFAS LORs - Water, Soil/Sediments & Biotic Matrices

Per- and Polyfluoroalkyl Substances (PFAS)	CAS No.[a]	MW	WATER (Potable, surface, groundwater, saline		SOLIDS (Soil, sediment, biosolids)		BIOTA*			
			LOR (µg/L)	LOR Trace (µg/L)	LOR (µg/kg)	LOR Trace (µg/kg)	Type 1 LOR (ng/mL)	Type 2 LOR (µg/kg)	Type 3 LOR (µg/kg)	Type 2 Trace LOR (µg/kg)
Perfluoroalkyl carboxylic acids (PFCAs)										
Perfluorobutanoic acid (PFBA)	375-22-4	214.04	0.05	0.005	5	0.1	0.5	0.5	1	0.1
Perfluoropentanoic acid (PFPeA)	2706-90-3	264.05	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorohexanoic acid (PFHxA)	307-24-4	314.05	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluoroheptanoic acid (PFHpA)	375-85-9	364.06	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorooctanoic acid (PFOA)	335-67-1	414.07	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorononanoic acid (PFNA)	375-95-1	464.08	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorodecanoic acid (PFDA)	335-76-2	514.08	0.01	0.001	5	0.1	0.5	0.5	1	0.5
Perfluoroundecanoic acid (PFUnA)	2058-94-8	564.09	0.01	0.001	5	0.1	0.5	0.5	1	0.5
Perfluorododecanoic acid (PFDoA)	307-55-1	614.10	0.01	0.001	5	0.1	0.5	0.5	1	0.5
Perfluorotridecanoic acid (PFTrDA)	72629-94-8	664.11	0.01	0.001	5	0.1	0.5	0.5	1	0.5
Perfluorotetradecanoic acid (PFTeDA)	376-06-7	714.11	0.01	0.001	5	0.1	0.5	0.5	1	0.5
Perfluoroalkyl sulfonic acids (PFSAs)										
Perfluoropropanesulfonic acid (PFPrS)	423-41-6	250.09	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorobutanesulfonic acid (PFBS)	375-73-5	300.10	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluoropentane sulfonic acid (PFPeS)	2706-91-4	350.11	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorohexane sulfonate (PFHxS)	355-46-4	400.11	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Potassium perfluorohexanesulfonate (linear and branched isomers) (br-PFHxS)			0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluoroheptane sulfonate (PFHpS)	375-92-8	450.12	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorooctane sulfonic acid (PFOS)[g ,h]	1763-23-1	500.13	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Potassium perfluorooctanesulfonate (linear and branched isomers) (br-PFOS)			0.01	0.001	5	0.1	0.5	0.5	1	0.1

Per- and Polyfluoroalkyl Substances (PFAS)	CAS No.[a]	MW	WATER (Potable, surface, groundwater, saline		SOLIDS (Soil, sediment, biosolids)		BIOTA*			
			LOR (µg/L)	LOR Trace (µg/L)	LOR (µg/kg)	LOR Trace (µg/kg)	Type 1 LOR (ng/mL)	Type 2 LOR (µg/kg)	Type 3 LOR (µg/kg)	Type 2 Trace LOR (µg/kg)
Perfluorononanesulfonic acid (PFNS)	68259-12-1	550.13	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluorodecanesulfonic acid (PFDS)	67906-42-7	617.18	0.01	0.001	5	0.1	0.5	0.5	1	0.1
Perfluoroalkane sulfonamides (FASAs), Perfluoroalkane sulfonamido ethanols (FASEs) and N-alkyl perfluoroalkane sulfonamido ethanols (MeFASEs, EtFASEs) Perfluoroalkane sulfonamido acetic acids (FASAAs) and N-alkyl perfluoroalkane sulfonamido acetic acids (MeFASAAs, EtFASAAs)										
Perfluorooctane sulfonamide (FOSA)	754-91-6	499.14	0.05	0.005	10	1	5	0.5	5	0.5
N-Methylperfluorooctane sulfonamide (MeFOSA)	31506-32-8	513.17	0.05	0.005	10	1	5	0.5	5	0.5
N-Ethylperfluorooctane sulfonamide (EtFOSA)	4151-50-2	527.19	0.05	0.005	10	1	5	2	5	0.5
N-Methylperfluorooctane sulfonamidoethanol (MeFOSE)	24448-09-7	557.22	0.05	0.005	10	1	5	1	5	0.5
N-Ethylperfluorooctane sulfonamidoethanol (EtFOSE)	1691-99-2	571.25	0.05	0.005	10	1	5	1	5	0.5
N-Ethylperfluorooctanesulfonamido acetic acid (EtFOSAA)	2991-50-6	585.23	0.05	0.005	10	1	5	0.5	5	0.5
N-Methylperfluorooctanesulfonamido acetic acid (N-MeFOSAA)	2355-31-9	571.21	0.05	0.005	10	1	5	0.5	5	0.5
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)										
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2 FTSA)	757124-72-4	328.15	0.01	0.001	5	0.5	5	0.5	5	0.1
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2 FTSA)	27619-97-2	428.16	0.01	0.001	5	0.5	5	0.5	5	0.1
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2 FTSA)	39108-34-4	528.18	0.01	0.001	5	0.5	5	1	5	0.1
1H, 1H, 2H, 2H-perfluorododecane sulfonate (10:2 FTSA)	120226-60-0	628.20	0.01	0.001	5	0.5	5	1	5	0.5
Additional PFAS Compounds										
Hexafluoropropylene oxide dimer acid (HFPO-DA) [GenX]	13252-13-6[b]	285[f]	0.01		5					
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11Cl-PF_3OUdS) [11Cl-F53B]	763051-92-9[c]	631	0.01		5					
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9Cl-PF_3ONS) [9Cl-F53B]	756426-58-1[d]	531	0.01		5					
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4[e]	377	0.01		5					

Per- and Polyfluoroalkyl Substances (PFAS)	CAS No.[a]	MW	WATER (Potable, surface, groundwater, saline		SOLIDS (Soil, sediment, biosolids)		BIOTA*			
			LOR (µg/L)	LOR Trace (µg/L)	LOR (µg/kg)	LOR Trace (µg/kg)	Type 1 LOR (ng/mL)	Type 2 LOR (µg/kg)	Type 3 LOR (µg/kg)	Type 2 Trace LOR (µg/kg)
Nonafluoro-3,6-dioxaheptanoic acid (NFDHA)	151772-58-6	296.045	0.01		5					
Perfluoro(2-ethoxyethane)sulfonic acid (PFEESA)	113507-82-7	316.09	0.01		5					
Perfluoro-3-methoxypropanoic acid (PFMPA)	377-73-1	230.038	0.01		5					
Perfluoro-4-methoxybutanoic acid (PFMBA)	863090-89-5	280.046	0.01		5					
6:2 fluorotelomer sulfonamide alkylbetaine (6:2 FTAB)[i]	34455-29-3	570.37	0.01		5					
3:3 Fluorotelomercarboxylic acid (3:3 FTCA)	356-02-5	242.093	0.01		5					
5:3 Fluorotelomer carboxylic acid (5:3 FTCA)	914637-49-3	342.108	0.01		5					
Perfluoropropane sulfonic acid (PFPrS)	423-41-6	248.90	0.01		5					
Perfluoroethylcyclohexane sulfonate (PFECHS)	67584-42-3	500.22	0.01		5					

[a] Some PFAS are commercially available as ammonium, sodium and potassium salts. This method measures all forms of the analytes as anions while the counterion is inconsequential. Analytes may be purchased as acids or as any of the corresponding salts (see Section 7.2.3 regarding correcting the analyte concentration for the salt content).
[b] HFPO-DA is one component of the GenX processing aid technology.
[c] 11Cl-PF_3OUdS is available in salt form (e.g., CASRN of potassium salt is 83329-89-9).
[d] 9Cl-PF_3ONS analyte is available in salt form (e.g., CASRN of potassium salt is 73606-19-6)
[e] ADONA is available as the sodium salt (no CASRN) and the ammonium salt (CASRN is 958445-448).
[f] HFPO-DA is not stable in the ESI source and the [M-H]- is not observed under typical ESI conditions. The precursor ion used during method development was [M-CO2]-.
[g] Analyte has multiple resolved chromatographic peaks due to linear and branched isomers. All peaks summed for quantitation purposes.
[h] To reduce bias regarding detection of branched and linear isomers, the m/z 80 product ion must be used for this analyte.
[i] NICNAS 6:2 Fluorotelomer sulfonamide surfactants: Environment tier II assessment 12 December 2019

BIOTA Key
Type 1 - Human and Animal Blood (whole blood & plasma)
Type 2 - Citrus, tomato, zucchini, grasses, squash; muscle tissue of fish, crustaceans, cheese, cow, sheep; kidney tissue of sheep and cow; milk and chicken egg
Type 3 - Sheep and cow liver; olives and avocado

Table 4: PFAS Analytes (n=30)

PFAS Compounds (n=30)	Acronym	CASRN	EPA Method 533	EPA Method 537.1	EPA Method 1633
Perfluoroalkyl carboxylic acids (C4-C14)	**PFCAs**				
Perfluorobutanoic acid	PFBA	375-22-4	X	X	X
Perfluoropentanoic acid	PFPeA	2706-90-3			
Perfluorohexanoic acid	PFHxA	307-24-4	X	X	X
Perfluoroheptanoic acid	PFHpA	375-85-9	X	X	X
Perfluorooctanoic acid	PFOA	335-67-1	X	X	X
Perfluorononanoic acid	PFNA	375-95-1	X	X	X
Perfluorodecanoic acid	PFDA	335-76-2			X
Perfluoroundecanoic acid	PFUnA	2058-94-8	X	X	X
Perfluorododecanoic acid	PFDoA	307-55-1	X	X	X
Perfluorotridecanoic acid	PFTrDA	72629-94-8			X
Perfluorotetradecanoic acid	PFTeDA	376-06-7	X	X	X
Perfluoroalkyl sulfonic acids (C3-C10)	**PFSAs**				
Perfluoropropanesulfonic acid	PFPrS	423-41-6			
Perfluorobutanesulfonic acid	PFBS	375-73-5			X
Perfluoropentane sulfonic acid	PFPeS	2706-91-4			X
Perfluorohexane sulfonate	PFHxS	355-46-4	X	X	X
Perfluoroheptane sulfonate	PFHpS	375-92-8			X
Perfluorooctane sulfonic acid[g,h]	PFOS	1763-23-1	X	X	X
Perfluorononanesulfonic acid	PFNS	68259-12-1			X
Perfluorodecanesulfonic acid	PFDS	67906-42-7			X
Perfluorododecanesulfonic acid	PFDoS	79780-39-5			X
Perfluoroalkane sulfonamides	**FASAs**				
Perfluorooctane sulfonamide	PFOSA	754-91-6			X
N-Methylperfluorooctane sulfonamide	NMeFOSA	31506-32-8			X
N-Ethylperfluorooctane sulfonamide	NEtFOSA	4151-50-2			X
Perfluoroalkane sulfonamido ethanols, N-alkyl perfluoroalkane sulfonamido ethanols	**FASEs, MeFASEs, EtFASEs**				
N-Methylperfluorooctane sulfonamidoethanol	NMeFOSE	24448-09-7			X
N-Ethylperfluorooctane sulfonamidoethanol	NEtFOSE	1691-99-2			X

PFAS Compounds (n=30)	Acronym	CASRN	EPA Method 533	EPA Method 537.1	EPA Method 1633
Perfluoroalkane sulfonamido acetic acids (FASAAs) and N-alkyl perfluoroalkane sulfonamido acetic acids (MeFASAAs, EtFASAAs)					
N-Ethylperfluorooctanesulfonamido acetic acid	NEtFOSAA	2991-50-6			X
N-Methylperfluorooctanesulfonamido acetic acid	NMeFOSAA	2355-31-9			X
n:2 Fluorotelomer sulfonic acids	**n:2 FTSAs**				
1H,1H,2H,2H-Perfluorohexanesulfonic Acid	4:2 FTSA	757124-72-4	X		X
1H,1H,2H,2H-Perfluorooctanesulfonic Acid	6:2 FTSA	27619-97-2	X		X
1H,1H,2H,2H-Perfluorodecanesulfonic Acid	8:2 FTSA	39108-34-4	X		X
1H, 1H, 2H, 2H-perfluorododecane sulfonate	10:2 FTSA	120226-60-0			

Table 5: PFAS Analytes – Supplemental

Compound	Acronym	CASRN	EPA Method 533	EPA Method 537.1	EPA Method 1633
Perfluoroalkyl mono-ether carboxylic acids	mono-ether PFECAs				
Hexafluoropropylene oxide dimer acid	HFPO-DA [GenX)	13252-13-6[b]	X	X	X
Perfluoroalkyl multi-ether carboxylic acids	multi-ether PFECAs				
Nonafluoro-3,6-dioxaheptanoic acid	NFDHA	151772-58-6	X		X
Perfluoro(2-ethoxyethane)sulfonic acid	PFEESA	113507-82-7	X		X
Perfluoro-3-methoxypropanoic acid	PFMPA	377-73-1	X		X
Perfluoro-4-methoxybutanoic acid	PFMBA	863090-89-5	X		X
Perfluoro(3,5-dioxahexanoic) acid	PFO_2HxA	39492-88-1			
Polyfluoroalkyl ether acids					
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid	11Cl-PF_3OUdS [11Cl-F53B]	763051-92-9[c]	X	X	X
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid	9Cl-PF_3ONS [9Cl-F53B]	756426-58-1[d]	X		X
4,8-dioxa-3H-perfluorononanoic acid	ADONA	919005-14-4[e]	X		X
6:2 Fluorotelomer sulfonamide surfactants					
6:2 fluorotelomer sulfonamide alkylbetaine[i]	6:2 FTAB	34455-29-3			

Compound	Acronym	CASRN	EPA Method 533	EPA Method 537.1	EPA Method 1633
3:3 Fluorotelomercarboxylic acid (3-Perfluoropropyl propanoic acid)	3:3 FTCA	356-02-5			X
5:3 Fluorotelomer carboxylic acid (2H,2H,3H,3H-Perfluorooctanoic acid)	5:3 FTCA	914637-49-3			X
7:3 Fluorotelomer carboxylic acid (3-Perfluoroheptyl propanoic acid)	7:3 FTCA	812-70-4			X
Perfluorinated alkane sulfonic acids					
Perfluoroethylcyclohexane sulfonate	PFECHS	67584-42-3			
Cyclohexanesulfonic acid, nonafluorobis(trifluoromethyl)-, potassium salt (1:1)		68156-01-4			
Cyclohexanesulfonic acid, decafluoro(trifluoromethyl)-, potassium salt (1:1)		68156-07-0			
Potassium perfluorocyclohexyl sulfonate		3107-18-4			

[a] Some PFAS are commercially available as ammonium, sodium and potassium salts. This method measures all forms of the analytes as anions while the counterion is inconsequential. Analytes may be purchased as acids or as any of the corresponding salts (see Section 7.2.3 regarding correcting the analyte concentration for the salt content).
[b] HFPO-DA is one component of the GenX processing aid technology.
[c] 11Cl-PF_3OUdS is available in salt form (e.g. CASRN of potassium salt is 83329-89-9).
[d] 9Cl-PF_3ONS analyte is available in salt form (e.g. CASRN of potassium salt is 73606-19-6)
[e] ADONA is available as the sodium salt (no CASRN) and the ammonium salt (CASRN is 958445-448).
[f] HFPO-DA is not stable in the ESI source and the [M-H]- is not observed under typical ESI conditions. The precursor ion used during method development was [M-CO2]-.
[g] Analyte has multiple resolved chromatographic peaks due to linear and branched isomers. All peaks summed for quantitation purposes.
[h] To reduce bias regarding detection of branched and linear isomers, the m/z 80 product ion must be used for this analyte.
[i] NICNAS 6:2 Fluorotelomer sulfonamide surfactants: Environment tier II assessment 12 December 2019

Table 6: Target Analytes for PPCP analysis

Common Name[a]	CAS[b] Registry Number	LOR (µg/L)
Acetaminophen[1,3]	103-90-2	0.025
Atenolol[3]	29122-68-7	0.025
Bezafibrate[4]	41859-67-0	0.025
Caffeine[1,3]	58-08-2	0.025
Carbamazepine[1,3]	298-46-4	0.025
Chlorpheniramine[3]	132-22-9	0.025
Ciprofloxacin[1,3]	85721-33-1	0.025
Clofibric acid[3]	882-09-7	0.025
Fluoxetine[1,3]	54910-89-3	0.025
Metoprolol[3]	51384-51-1	0.025
Norfloxacin[1,3]	70458-96-7	0.025
Propranolol[3]	525-66-6	0.025
Sertraline[3]	79617-96-2	0.025
Sotalol[4]	3930-20-9	0.025
Sulfamethoxazole[3]	723-46-6	0.025
Triclocarban[2,4]	101-20-2	0.025
Trimethoprim[1,3]	738-70-5	0.025
Warfarin[2,4]	81-81-2	0.025

[a] Some PPCP standards are commercially available as ammonium, sodium, and potassium salts. This method measures all forms of the analytes as anions while the identity of the counter ion is inconsequential. Analytes may be purchased as acids or as any of the corresponding salts
[b] Chemical Abstract Service.
[1] Compound targeted in US EPA Method 1694, Group 1 compound
[2] Compound targeted in US EPA Method 1694, Group 3 compound
[3] ISO17034:2016 and ISO17025 accredited
[4] ISO17025 accredited

GLOSSARY

These definitions and purposes are specific to DM1633 method, but have been conformed to common usage to the extent possible.

Units of weight and measure and their abbreviations

Symbols

ºC degrees	Celsius
Da	Dalton (equivalent to “amu” below)
µg	microgram
µL	microliter
µm	micrometre
<	less than
≤	less than or equal
>	greater than
≥	greater than or equal
%	percent
±	plus or minus

Alphabetical abbreviations

amu	atomic mass unit (equivalent to Dalton)
cm	centimetre
g	gram
h	hour
L	litre
M	molar
mg	milligram
min	minute
mL	millilitre
mm	millimetre
m/z	mass-to-charge ratio
ng	nanogram
Q1	quantitation ion
Q2	confirmation ion
rpm	revolutions per minute
v/v	percent volume per volume

Definitions and acronyms (in alphabetical order)

Analyte – A PFAS compound included in this method.

Calibration standard (CS) – A solution prepared from a secondary standard and/or stock solutions and used to calibrate the response of the LC-MS/MS instrument.

Calibration verification standard (CV) – The mid-point calibration standard (CS-4) that is used to verify calibration.

Compound - One of many variants or configurations of a common chemical structure. Individual compounds are identified by the number of carbon atoms and functional group attached at the end of the chain.

Class A glassware – Volumetric glassware that provides the highest accuracy. Class A volumetric glassware complies with the Class A tolerances defined in ASTM E694, must be permanently labelled as Class A, and is supplied with a serialized certificate of precision.

CWA – Clean Water Act

Extracted internal standard (EIS) quantification – The response of the target compound is compared to the response of the labelled analogue of another compound in the same LOC.

LC – Liquid chromatograph or liquid chromatography

Internal standard – A labelled compound used as a reference for quantitation of other labelled compounds and for quantitation of native PFAS compounds other than the compound of which it is a labelled analogue. See Internal standard quantitation.

Instrument sensitivity check – solution used to check the sensitivity of the instrument. The solution contains the native compounds at the concentration of the LOQ.
Internal standard quantitation – A means of determining the concentration of (1) a naturally occurring (native) compound by reference to a compound other than its labelled analogue and (2) a labelled compound by reference to another labelled compound
IPR – Initial precision and recovery; four aliquots of a reference matrix spiked with the analytes of interest and labelled compounds and analysed to establish the ability of the laboratory to generate acceptable precision and recovery. An IPR is performed prior to the first time this method is used and any time the method or instrumentation is modified.
Isotope dilution (ID) quantitation – A means of determining a naturally occurring (native) compound by reference to the same compound in which one or more atoms has been isotopically enriched. The labelled PFAS are spiked into each sample and allow identification and correction of the concentration of the native compounds in the analytical process.
Isotopically labelled compound – An analogue of a target analyte in the method which has been synthesized with one or more atoms in the structure replaced by a stable (non-radioactive) isotope of that atom. Common stable isotopes used are 13C (Carbon-13) or Deuterium (D or 2H). These labelled compounds do not occur in nature, so they can be used for isotope dilution quantitation or other method-specific purposes.
Limit of Quantitation (LOQ) – The smallest concentration that produces a quantitative result with known and recorded precision and bias. The LOQ shall be set at or above the concentration of the lowest initial calibration standard (the lowest calibration standard must fall within the linear range).
Method blank – An aliquot of reagent water that is treated exactly as a sample including exposure to all glassware, equipment, solvents, reagents, internal standards, and labelled compounds that are used with samples. The method blank is used to determine if analytes or interferences are present in the laboratory environment, the reagents, or the apparatus.
Method Detection Limit (MDL) – The minimum measured concentration of a substance that can be reported with 99% confidence that the measured analyte concentration is distinguishable from method blank results (40 CFR 136, Appendix B).
MESA – Mining Enforcement and Safety Administration
Minimum level of quantitation (ML) – The lowest level at which the entire analytical system must give a recognizable signal and acceptable calibration point for the analyte. The ML represents the lowest concentration at which an analyte can be measured with a known level of confidence. It may be equivalent to the concentration of the lowest calibration standard, assuming that all method-specified sample weights, volumes, and clean-up procedures have been employed. Alternatively, the ML may be established by multiplying the MDL (pooled or un-pooled, as appropriate) by 3.18 and rounding the result to the number nearest to 1, 2, or 5 x 10n, where n is zero or an integer (see 68 FR 11770).
MS – Mass spectrometer or mass spectrometry
Matrix Spike/Matrix Spike Duplicate (MS/MSD) – Aliquots of field samples that have been fortified with a known concentration of target compounds, prior to sample preparation and extraction, and analysed to measure the effect of matrix interferences. The use of MS/MSD samples is generally not required in isotope dilution methods because the labelled compounds added to every sample provide more performance data than spiking a single sample in each preparation batch.
Multiple reaction monitoring (MRM) – Also known as selected reaction monitoring (SRM). A type of mass spectrometry where a parent mass of the compound is fragmented through MS/MS and then specifically monitored for a single fragment ion.
Must – This action, activity, or procedural step is required.
NIOSH – The National Institute of Occupational Safety and Health
Non-extracted internal standard (NIS) –Labelled PFAS compounds spiked into the concentrated extract immediately prior to injection of an aliquot of the extract into the LC-MS/MS.
OPR – Ongoing precision and recovery standard (OPR); a method blank spiked with known quantities of analytes. The OPR is analysed exactly like a sample. Its purpose is to assure that the results produced by the laboratory remain within the limits specified in this method for precision and recovery.

Precursor Ion – For the purpose of this method, the precursor ion is the deprotonated molecule ([M-H]- of the method analyte. In MS/MS, the precursor ion is mass selected and fragmented by collisionally activated dissociation to produce distinctive product ions of smaller *m/z*.
PFAS – Per- and Polyfluoroalkyl substances –A group of man-made fluorinated compounds that are hydrophobic and lipophobic, manufactured and used in a variety of industries globally. These compounds are persistent in the environment as well as in the human body.
Reagent water – Water demonstrated to be free from the analytes of interest and potentially interfering substances at the method detection limit for the analyte.
Relative standard deviation (RSD) – The standard deviation multiplied by 100 and divided by the mean. Also termed "coefficient of variation."
Relative Standard Error (RSE) – The standard error of the mean divided by the mean and multiplied by 100.
RF – Response factor.
RR – Relative response.
RT – Retention time**;** the time it takes for an analyte or labelled compound to elute off the HPLC/UPLC column
Should – This action, activity, or procedural step is suggested but not required.
Signal-to-noise ratio (S/N) – The height of the signal as measured from the mean (average) of the noise to the peak maximum divided by the mean height of the noise.
SPE – Solid-phase extraction; a technique in which an analyte is extracted from an aqueous solution or a solid/tissue extract by passage over or through a material capable of reversibly adsorbing the analyte. Also termed liquid-solid extraction.
Stock solution – A solution containing an analyte that is prepared using a reference material traceable to EPA, NIST, or a source that will attest to the purity and authenticity of the reference material.

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