

Chapter 2: Sampling

2.1 Object

The object of sampling is to obtain a sample of a size suitable for tests, in which the probability of a constituent being present is determined only by its level of occurrence in the seed lot.

2.2 Definitions

2.2.1 Seed lot

A seed lot is a specified quantity of seed that is physically and uniquely identifiable.

2.2.2 Sublot

A sublot is a portion of not less than 20 % of the seed lot. Each container of a sublot must be marked with the identification of the seed lot.

2.2.3 Primary sample

A primary sample is a portion taken from the seed lot during one single sampling action.

2.2.4 Composite sample

The composite sample is formed by combining and mixing all the primary samples taken from the seed lot.

2.2.5 Subsample

A subsample is a portion of a sample obtained by reducing a sample.

2.2.6 Submitted sample

A submitted sample is a sample that is to be submitted to the testing laboratory and may comprise either the whole of the composite sample or a subsample thereof. The submitted sample may be divided into subsamples packed in different material meeting conditions for specific tests (e.g. moisture or health).

2.2.7 Duplicate sample

A duplicate sample is another sample obtained for submission from the same composite sample and marked 'Duplicate sample'.

2.2.8 Working sample

The working sample is the whole of the submitted sample or a subsample thereof, on which one of the quality tests described in these ISTA Rules is made and must be at least the weight prescribed by the ISTA Rules for the particular test.

2.2.9 Sealed

Sealed means that a container in which seed is held is closed in such a way, that it cannot be opened to gain access to the seed and closed again, without either destroying the seal or leaving evidence of tampering. This definition refers to the sealing of seed lots, as well as of seed samples.

2.2.10 Self-sealing containers

The 'valve-pack' bag is a specific type of self-sealing container. It is filled through a sleeve-shaped valve which is automatically closed by the completion of filling the bag.

2.2.11 Marked/labelled

A container of a seed lot can be considered as marked or labelled when there is a unique identification mark on the container, which defines the seed lot to which the container belongs. All containers of a seed lot must be marked with the same unique seed lot designation (numbers, characters or combination of both). Should the unique identification mark be indicated on a label attached to the container, it must not be possible to remove the label and replace it with another label without showing signs of tampering. Marking of samples and subsamples must ensure that there is always an unambiguous link between the seed lot and the samples and subsamples.

2.2.12 Treated seed

‘Seed treatment’ is a generic term which indicates that a seed lot has been subjected to:

- a. the application of a compound including film coatings, polymers, pesticides, fungicides, biologicals, identifying colourants, dyes and/or other additives;
- b. the application of a biological product including micro-organisms;
- c. a process including wetting and drying; or
- d. an energy form including heat, radiation, electricity or magnetism;

but does not specify the application method.

Seed treatment does not significantly change the shape and the general size of the untreated seed with only a minimal weight gain. Treated seeds are usually tested without removing the treatment and according to the same rules as untreated seeds.

2.2.13 Coated seeds

Coated seeds are seeds covered with material in such a way that in most cases the seeds cannot be identified without removing the covering material. The material may contain pesticides, fungicides, biologicals, identifying colourants, dyes and/or other additives. The following types of coated seeds are defined:

Seed pellets. More or less spherical units, usually incorporating a single seed with the size and shape of the seed no longer readily evident.

Encrusted seed. Units more or less retaining the shape of the seed with the size and weight changed to a measurable extent.

Seed granules. Units more or less cylindrical, including types with more than one seed per granule.

Seed tapes. Narrow bands of material, such as paper or other degradable material, with seeds spaced randomly, in groups or in a single row.

Seed mats. Broad sheets of material, such as paper or other degradable material, with seeds placed in rows, groups or at random throughout the sheets.

2.2.14 Small seed lots

Small seed lots are seed lots of high-value seed, where obtaining a submitted sample of standard size could have a substantial effect on the quantity of the remaining seed lot. High-value seed includes, but is not limited to, hybrid vegetable seeds that are sold per seed, or seed that is not commercially available and is used for research or for higher generation multiplication.

2.3 General principles

A composite sample is obtained from the seed lot by taking primary samples from different positions in the whole seed lot and combining them. From this composite sample, subsamples are obtained by sample reduction procedures at one or more stages forming the submitted sample and finally the working samples for testing. For issuing ISTA Certificates, specific requirements have to be fulfilled as given under 2.5.4. Further information on seed sampling can be found in the current *ISTA Handbook on Seed Sampling*.

2.4 Apparatus

Sampling and sample reduction must be performed using appropriate techniques and equipment that is clean and in good condition as described in 2.5.1 and 2.5.2.2.

Containers used to collect primary samples, composite samples and during mixing and dividing must be static-free to avoid chaff or small seeds adhering to the inside of the containers.

2.5 Procedures

2.5.1 Procedures for sampling a seed lot

2.5.1.1 Preparation of a seed lot and conditions for sampling

At the time of sampling, the seed lot must be as uniform as practicable. If the seed lot is found to be obviously heterogeneous, sampling must be refused or stopped. In cases of doubt heterogeneity can be determined as described under 2.9.

Seed may be sampled in containers or from the seed stream, either before or when it enters containers. The containers in which seed is held must be fit for purpose, i.e. must not damage the seed, must be clean to avoid cross contamination, and must be sealable. The containers must be labelled or marked before or just after sampling is completed.

The seed lot must be so arranged that each part of the seed lot is conveniently accessible.

2.5.1.2 Minimum sampling intensity

For seed lots in containers holding up to and including 100 kg, the minimum sampling intensity is the following:

- For containers holding between 15 kg and 100 kg (inclusive) of seed, the number of primary samples according to Table 2A.
- For containers holding less than 15 kg of seed, containers must be combined into sampling units not exceeding 100 kg, e.g. 20 containers of 5 kg, 33 containers of 3 kg or 100 containers of 1 kg. The sampling units must be regarded as containers as described in Table 2A.
- For seed pellets, seed granules, seed tapes and seed mats, containers of less than 300 000 seed units must be combined to sampling units not exceeding 2 000 000 seeds. The sampling units must be regarded as containers as described in Table 2A.

Table 2A. Minimum sampling intensity for seed lots in containers holding up to and including 100 kg seed

Number of containers	Minimum number of primary samples to be taken
1–4	3 primary samples from each container
5–8	2 primary samples from each container
9–15	1 primary sample from each container
16–30	15 primary samples, one each from 15 different containers
31–59	20 primary samples, one each from 20 different containers
60 or more	30 primary samples, one each from 30 different containers

When sampling seed in containers holding more than 100 kg of seed, or from streams of seed entering containers, the sampling intensity according to Table 2B must be regarded as the minimum requirement.

Table 2B. Minimum number of primary samples to be taken from seed lots in containers holding more than 100 kg of seed, or from seed streams

Seed lot size	Number of primary samples to be taken
Up to 500 kg	At least five primary samples
501–3 000 kg	One primary sample for each 300 kg, but not less than five
3 001–20 000 kg	One primary sample for each 500 kg, but not less than 10
20 001 kg and above	One primary sample for each 700 kg, but not less than 40

When sampling a seed lot of up to 15 containers, regardless of their size, the same number of primary samples must be taken from each container.

Sampling intensity for coated seeds is as described in Tables 2A and 2B.

2.5.1.3 Taking primary samples

When defining the number and/or the size of primary samples, the seed sampler needs to ensure (besides meeting the minimum sampling intensity) that the minimum amount of seed required for the requested test(s) is sent to the testing laboratory and enough seed remains available for obtaining duplicate samples if requested.

Primary samples of approximately equal size must be taken from a seed lot, irrespective of where in the lot or container the primary sample is taken.

When the seed lot is in containers, the containers to be sampled must be selected at random or according to a systematic plan throughout the seed lot. Primary samples must be drawn from the top, middle and bottom of containers, but not necessarily from more than one position in any container, unless so specified in Tables 2A and 2B.

When the seed is in bulk or in large containers, the primary samples must be drawn from random positions.

Containers must be opened or pierced for abstraction of primary samples. The sampled containers must then be closed or the contents transferred to new containers.

When seed is to be packed in special types of containers (e.g. small, not penetrable, or moisture-proof containers), it should be sampled, if possible, either before or during the filling of the containers.

Sampling seed lots of seed tapes and seed mats should be done by taking packets or pieces of tape or mat.

The instruments being used must neither damage the seed nor select according to seed size, shape, density, chaffiness or any other quality trait. All sampling apparatus must be clean before use to prevent cross contaminations. Triers must be long enough so that the opening at the tip reaches at least half of the diameter of the container. When the container is not accessible from opposite sides, the trier must be long enough to reach the opposite side.

Sampling seed lots may be done by one of the methods listed below.

a. Automatic sampling from a seed stream. Seed may be sampled by automatic sampling devices, provided that the instrument uniformly samples the cross section of the seed stream and the material entering the instrument does not bounce out again. It may be operated either under manual or automatic control. The intervals between taking primary samples should be constant.

b. Manual sampling from a seed stream. Seed streams may also be sampled by using manual instruments when fulfilling the requirements listed under 'a'.

c. Sampling stick. The sampling stick (e.g. stick trier, sleeve type trier, spiral trier) consists of two parts, one of which fits loosely inside the other, but tightly enough so that seed or impurities do not slip between them. The outer part has a solid pointed end. Both parts have slots in their walls so that the cavity of the inner part can be opened and closed by moving the two parts against each other by either a twisting or a push-pull motion.

The sampling stick may be used horizontally, diagonally or vertically. The spiral trier has slots in a spiral arrangement for their subsequent opening from the tip to the handle and may only be used for seeds of a size smaller than *Triticum aestivum* L. subsp. *aestivum*.

However, when used vertically or diagonally downwards, the sampling stick must either have partitions dividing the instrument into a number of compartments or have slots in a spiral arrangement. The minimum inside diameter should be wide enough to allow the smooth and free flow of seed and contaminants into the sampling stick.

When using the sampling stick, insert it in the closed position into the container, gently push it so that the point reaches the required position, open the sampling stick, agitate it slightly to allow it to fill completely, gently close and withdraw it and empty the primary sample into a container. Care should be exercised in closing the sampling stick so that seeds are not damaged.

d. Nobbe trier. The Nobbe trier (dynamic spear) is a pointed tube with an opening near the pointed end. Seed passes through the tube and is collected in a container. The minimum internal diameter of the Nobbe trier should be wide enough to allow the smooth and free flow of seed and contaminants through the trier. When using the Nobbe trier, insert it at an angle of about 30° to the horizontal plane with the opening facing down, push the trier until it reaches the required position and revolve it through 180°. Withdraw it with decreasing speed from the container, gently agitating the trier to help maintain an even flow of seed, and

collect the seed sample coming from the trier in a suitable container.

e. Cargo sampler. The cargo sampler (bulk sampler) consists of a special type of chamber that is fixed to a shaft. The lower part of the chamber is cone-shaped with a pointed end. To reach a greater depth, the shaft may be lengthened by screwing on successive extensions. There is a closing system in the chamber that may be a collar on the outside of the instrument, a wing connected to a door or a valve with a spring. Some cargo samplers can be closed before they are drawn back from the sampling position; others cannot be closed, so that the filled chamber is open during withdrawal. For all species, the minimum inside diameter can be about 35 mm and the depth 75 mm. When using the cargo sampler, insert it in the closed position into the container, gently push it vertically into the seed so that the point reaches the required position, pull the cargo sampler back about 10 cm or turn it (depending on the closing system), agitate it slightly to allow it to fill completely, gently close if possible and withdraw it and empty the primary sample into a container. Care should be exercised in closing the cargo sampler, so that the seeds are not damaged.

f. Sampling by hand. This method can be used for all species and may be the most suitable method for seed that may be damaged by the use of triers, seeds with wings, seeds with low moisture content, seed tapes and seed mats.

For hand sampling seed in containers, all positions inside the containers must be accessible. Containers with layers which are not accessible from the regular opening may have to be cut open, sampled and repackaged. Containers may also be partially or completely emptied during the sampling process to gain access to all positions in the containers. For sampling by hand, clean the hand and roll the sleeve up if necessary, insert the open hand into the container to the required position, close and withdraw the hand, taking great care that the fingers remain tightly closed about the seeds so none may escape, and empty the hand into a receiving pan.

2.5.1.4 Obtaining the composite sample

Where possible, the primary samples are compared with each other during sampling. The primary samples can only be combined to form the composite sample if they appear to be uniform. If not, the sampling procedure must be stopped. When primary samples are collected directly into one container, the content of this container may be regarded as the composite sample only if it appears uniform. If not, it must not be used for obtaining a submitted sample.

2.5.1.5 Obtaining the submitted sample

The composite sample can be submitted to the seed testing laboratory if it is of appropriate size for the tests to be conducted, or if it is difficult to mix and reduce the composite sample properly under warehouse conditions.

2.5.1.5.1 Obtaining the submitted sample for all tests

If the composite sample is too big, the submitted sample must be obtained by reducing the composite sample to an appropriate size by one of the methods referred to in 2.5.2.2. In the case of very large composite samples, a method according to 2.5.1.3 may also be used.

2.5.1.5.2 Obtaining the submitted sample for determination of moisture content

Obtaining submitted samples of the required size for moisture testing must be carried out in such a way that changes in moisture content are minimal.

Samples must be taken in the following way from the composite sample: first, mix the composite sample by either stirring it or by passing it through a mechanical divider and combining preferably once but not more than three times. Then, take a minimum of three subsamples from different positions and combine them to create the submitted sample for moisture testing.

2.5.1.5.3 Obtaining duplicate samples

Duplicate samples, which were requested no later than at the time of sampling, must be prepared in the same way as the submitted sample.

2.5.1.6 Packing and dispatch of the submitted sample

The submitted sample must be marked with the same identification as the seed lot. For an Orange International Seed Lot Certificate, the sample must be sealed, if it is not delivered personally by the sampler to the laboratory on the same premises (see 2.5.4.3). The additional information required according to 1.4.2 as well as the name of any chemical treatment applied must be provided.

Submitted samples must be packed so as to prevent damage during transit. Submitted samples should be packed in breathable containers.

Submitted samples for moisture testing, and samples from seed lots which have been dried to low moisture content, must be packed in moisture-proof containers which contain as little air as possible. Submitted samples for germination tests, viability tests and health tests may only be packed in moisture-proof containers if suitable storage conditions can be assured.

Submitted samples must be dispatched to the seed testing laboratory without delay.

2.5.1.7 Storage of submitted samples before testing

Every effort must be made to start testing a submitted sample on the day of receipt. Storage of orthodox seeds, when necessary, should be in a cool, well-ventilated room.

Non-orthodox (i.e. recalcitrant or intermediate) seeds should be tested as soon as possible after obtaining the submitted sample from the composite sample without any storage. Handling of the submitted sample and, if necessary, storage should be done under species specific optimum conditions.

2.5.2 Procedures for obtaining the submitted and working sample

2.5.2.1 Minimum size of working sample

Minimum sizes of working samples are prescribed in the appropriate chapter for each test. The working sample weights for purity analyses given in Table 2C are calculated to contain at least 2500 seeds. These weights are recommended for normal use in purity tests, see 3.5.1.

The sample weights in column 5 of Table 2C, Part 1, for counts of other species are 10 times the weights in column 4, subject to a maximum of 1000 g.

Working samples of all coated seeds except those defined as treated seed in 2.2.12 must contain at least the number of pellets, seeds or granules indicated in column 3 of Table 2D, Part 1 and Part 2. If a smaller sample is used, the actual number of pellets, seeds or granules in the sample must be reported.

2.5.2.2 Sample reduction methods

If the seed sample needs to be reduced to a size equal to or greater than the size prescribed, the seed sample must first be thoroughly mixed for all dividers and methods excluding the Variable sample divider and Rotary divider, where mixing takes place during the dividing process. The submitted/working sample must then be obtained either by repeated halving or by abstracting and subsequently combining small random portions. The apparatus and methods for sample reduction are described in 2.5.2.2.1 to 2.5.2.2.4. One, two or more of these methods may be used in one sample reduction procedure. When using one of the dividers described for seed pellets the distance of fall must not exceed 250 mm.

After obtaining a working sample the remainder must be re-mixed before a second working sample is obtained.

Except in the case of seed health, the method of hand halving must be restricted to certain genera listed in 2.5.2.2.4. Only the spoon method and the hand halving method may be used in the laboratory to obtain working samples for seed health testing where other samples or equipment may be contaminated by spores or other propagating material.

For seed tapes and mats take pieces of tape or mat at random, to provide sufficient seeds for the test.

2.5.2.2.1 Mechanical divider method

This method is suitable for all kinds of seeds except some very chaffy seeds. The apparatus divides a sample passed through it into two or more approximately equal parts. The submitted sample can be mixed by passing it through the divider, recombining the parts and passing the whole sample through a second time, and similarly, a third time if necessary. The sample is reduced by passing the seed through repeatedly and removing parts on each occasion. This process of reduction is continued until a working sample of approximately, but not less than, the required size is obtained.

- a. Conical divider. The conical divider (Boerner type) consists of a hopper, cone and a series of baffles directing the seed into two spouts. The baffles form alternate channels and spaces of equal width. They are arranged in a circle and are directed inward and downward, the channels leading to one spout and the spaces to an opposite spout. A valve or gate at the base of the hopper retains the seed. When the valve is opened the seed falls by gravity over the cone where it is evenly distributed to the channels and spaces, then passes through the spouts into the seed pans.

Channels and spaces must be wide enough to allow the smooth free flow of seed and contaminants. The more channels and spaces, the better the accuracy. Typical commercial conical dividers have about 15 channels and 15 spaces.

- b. Soil divider. The soil divider (riffle divider) consists of a hopper with attached channels or ducts alternately leading to opposite sides. Channels must be wide enough to allow the smooth free flow of seed and contaminants. The more channels, the better the accuracy. A minimum of 10 channels is required.

In using the divider the seed is placed evenly into a pouring pan and then poured in the hopper at approximately equal rates along the entire length. The seed passes through the channels and is collected in two receiving pans.

- c. Centrifugal divider. In the centrifugal divider (Gamet type) the seed flows downward through a hopper onto a shallow cup or spinner. Upon rotation of the spinner by an electric motor the seeds are thrown out by centrifugal force and fall downward. The circle or area where the seeds fall is equally divided into two parts by a stationary baffle so that approximately half the seeds fall in one spout and half in the other spout.

The centrifugal divider tends to give variable results unless the spinner is operated after having poured the seed centrally into the hopper.

- d. **Rotary divider.** The rotary divider comprises a rotating crown or base unit usually with 6 to 32 attached subsample containers, a vibration chute and a hopper. In using the divider the seed is poured into the hopper and the rotary divider is switched on so that the crown or base unit with the containers rotates with high speed and the vibration chute starts to feed the seed into the inlet cylinder of the rotating crown/base unit. The longer the duration of the dividing operation, the better the accuracy.

The feeding rate and therefore the duration of the dividing operation can be adjusted by the distance between the funnel of the hopper and chute, and the vibration intensity of the chute.

There are two principles: (i) the inlet cylinder feeds the seed centrally onto a distributor within the rotating crown or base unit, distributing the seed to all containers simultaneously; and (ii) the inlet cylinder feeds the seed de-centrally into the inlets of the containers rotating underneath the inlet cylinder so that the seed stream is subdivided into a lot of subsamples.

For this type of divider, mixing and dividing takes place in one operation.

- e. **Variable sample divider.** The variable sample divider consists of a pouring hopper and a rotating tube underneath. The tube distributes the seed stream from the pouring hopper onto the inner surface of a further hopper, which is well fitted into a third hopper, all being concentric. In the second and the third hopper there are slots that can be twisted against each other resulting in wider or narrower slots. The effect is that variable proportions will pass through the slots. The position of the hoppers in relation to each other can be adjusted accurately, resulting in predetermined sample sizes.

Depending on the design, the sample poured into the hopper can be divided into one or up to eight subsamples. The operation of these types of dividers can be controlled with computer software, which enables it to provide two or more subsamples with different predetermined sizes, in one operation.

For this type of divider, mixing and dividing takes place in one operation.

2.5.2.2.2 Modified halving method

The apparatus comprises a tray into which fits a grid of equal-sized cubical cells, open at the top and every alternate one having no bottom. After preliminary mixing,

the seed is poured evenly over the grid. When the grid is lifted, approximately half the sample remains on the tray. The submitted sample is successively halved in this way until a working sample, of approximately but not less than the required size, is obtained.

2.5.2.2.3 Spoon method

The spoon method is restricted to species with seeds smaller than *Triticum aestivum* L. subsp. *aestivum*, to the genera *Arachis*, *Glycine* and *Phaseolus*, and to tree genera *Abies*, *Cedrus* and *Pseudotsuga*. For all other species it can only be used to obtain working samples in the laboratory for seed health tests (7.4.1).

A tray, a spatula and a spoon with a straight edge are required. After preliminary mixing, pour the seed evenly over the tray; do not shake the tray thereafter. With the spoon in one hand, the spatula in the other, and using both, remove small portions of seed from not less than five random places. Sufficient portions of seed are taken to constitute a subsample of the required size.

2.5.2.2.4 The hand halving method

This method is restricted to the following genera of chaffy seeds:

Agrimonia, *Andropogon*, *Anthoxanthum*, *Arrhenatherum*, *Astrebula*, *Beckmannia*, *Bouteloua*, *Briza*, *Cenchrus*, *Chloris*, *Dichanthium*, *Digitaria*, *Echinochloa*, *Ehrharta*, *Elymus*, *Eragrostis*, *Gomphrena*, *Gossypium* (linted seed only), *Melinis*, *Oryza*, *Pennisetum* (non *glaucum*), *Psathyrostachys*, *Scabiosa*, *Sorghastrum*, *Stylosanthes* (non *guianensis*), *Trisetum*, *Urochloa*;

to the following genera of easily damaged fragile seeds:

Arachis, *Glycine* and *Phaseolus*;

and to the following genera and species of tree and shrub seeds:

Acer, *Aesculus*, *Ailanthus*, *Castanea*, *Cedrela*, *Corylus*, *Fagus*, *Fraxinus*, *Juglans*, *Liriodendron*, *Pinus cembra*, *Pinus pinea*, *Platanus*, *Populus*, *Quercus*, *Salix*, *Tectona*, *Ulmus*.

The hand halving method can also be used with the species where all other dividing methods are extremely difficult or impossible to use.

For all other species it can be used only to obtain working samples in the laboratory for seed health tests (7.4.1).

For applying the hand halving method, pour the sample evenly onto a smooth clean surface, thoroughly mix the seed into a mound with a flat-edged spatula, divide the mound into half and halve each half again – giving four portions – and halve each portion again – giving eight portions, arrange the portions in two rows of four, combine and retain alternate portions: e.g. combine the first and third portions in the first row with the second and fourth in the second row, remove the remaining four portions. Repeat the procedure using the retained portions until obtaining the required sample size.

2.5.3 Storage of samples after testing

The primary aim of storage of samples after testing is to be able to repeat the original tests carried out on the submitted sample. Therefore, storage conditions should be such that changes in the seed quality traits tested are minimal. For example, in the case of the purity test or other seed count, the sample should be stored in such a way that the physical identity is kept (see 3.5.2 and 4.5.2). In the case of germination, viability or health test of orthodox seeds the sample should be stored under cool and dry conditions. For such tests in recalcitrant and intermediate seeds of tropical and subtropical species, long term storage is not possible. For such seed of temperate species storability depends on the fungal status and to some extent whether the seed is dormant or not. All factors pertaining to storage need to be determined on a species basis. Protection against insects and rodents may be necessary.

To provide for re-testing by the original or by another seed testing laboratory, samples on which ISTA Certificates have been issued must be stored at least for one year from the receipt of the sample. Submitted samples in moisture proof containers, and samples of recalcitrant or intermediate species, must be stored under appropriate conditions for as long as it can be expected that the results of a re-test are not affected by the storage.

When a re-test in a different testing laboratory is required, a portion must be drawn from the stored sample in accordance with 2.5.2.2, and submitted to the designated testing laboratory. The remainder must be retained in store.

2.5.4 Conditions for issuing Orange International Seed Lot Certificates

The sampling methods laid down in the ISTA Rules must be followed when seed samples are drawn for the issue of Orange International Seed Lot Certificates. Further conditions have to be fulfilled as listed below.

2.5.4.1 Seed lot size

The seed lot must not exceed the quantity indicated in column 2 of Table 2C, subject to a tolerance of 5 % with the exception of:

- seed being transported loose in bulk containers. The conditions under which this exception may be permitted are laid down in Chapter 17.
- seed pellets, seed granules, seed tapes or seed mats. The maximum number of seeds that a seed lot of seed pellets, seed granules, seed tapes or seed mats may contain is 1 000 000 000 (10 000 units of 100 000) except that the weight of the seed lot, including the coating material may not exceed 40 000 kg subject to a tolerance of 5 % (42 000 kg).
- seed lots of species of Poaceae produced in a seed company that has been approved to make larger seed lots. The conditions under which this may be permitted are laid down in 2.5.4.2.
- seed lots of species of Poaceae produced in a seed company that has applied for approval to make larger seed lots according to 2.5.4.2. The heterogeneity of the seed lot must be tested according to 2.9 and the seed lot must not show significant heterogeneity.

Maximum lot size for treated and encrusted seeds is defined by applying the quantities indicated in Table 2C to the seeds without coating material.

A seed lot in excess of the prescribed quantity must be subdivided into seed lots not larger than the prescribed quantity, each of which must be labelled or marked with a separate seed lot identification.

2.5.4.2 Large seed lots of Poaceae

2.5.4.2.1 Definitions

Large seed lots of Poaceae species may have a maximum size of 25 000 kg (with a 5 % tolerance), if produced by an approved production plant.

For the purposes of large seed lots of Poaceae species, the following species with similar characteristics are regarded as two species groups:

Species group 1:

Lolium perenne, *Lolium multiflorum*, *Lolium ×hybridum* (previously *Lolium ×boucheanum*), *×Festulolium*, *Festuca pratensis*, *Festuca arundinacea* and *Phleum pratense*.

Species group 2:

Festuca rubra, *Festuca ovina*, *Festuca filiformis*, *Festuca heterophylla*, *Festuca trachyphylla*, *Dactylis glomerata*, *Poa pratensis* and *Poa trivialis*.

Approval which was granted following heterogeneity testing of any species of a group is also valid for all other species of the same group.

For all other species of Poaceae, approval must be requested and granted separately for each individual species.

2.5.4.2.2 Approval

Approval is granted after heterogeneity testing of six large seed lots of the species group or individual species for which the approval is requested. Heterogeneity testing must be carried out according to 2.9, and must as a minimum be based on purity and other seed count. At least five of the six tested seed lots must have a non-significant level of heterogeneity.

2.5.4.2.3 Check sampling and testing

After approval, the large seed lots of a production plant must be monitored by check sampling and further heterogeneity testing, according to 2.9, and as a minimum based on purity and other seed count.

Of the first 100 large seed lots per species group, 4 are randomly selected (4 % check sampling) and tested for heterogeneity. If none of these are heterogeneous, the check-sampling rate is reduced to 3 % for the following 100 lots, and to 2 % for subsequent lots.

However, if a check sample is found to show significant heterogeneity, the check-sampling rate must remain at 4 %, or again be increased from 3 to 4 % or from 2 to 3 %, as applicable (Fig. 2.1).

In six consecutive check samples tested, a maximum of one sample may show significant heterogeneity.

Hence, a heterogeneous sample must be followed by at least five non-heterogeneous samples in order for approval to be retained (Fig. 2.1).

2.5.4.2.4 Withdrawal of approval

If more than one of the last six consecutive check samples tested shows significant heterogeneity, approval must be withdrawn for the species or species group and production plant concerned, and the company must re-apply for approval (Fig. 2.1).

2.5.4.2.5 Responsibility

The Certifying or Designated Authority in a country is responsible for:

- the decision of approval of the seed company (production plant);
- ensuring that each production plant is approved separately, if a seed company has more than one production plant;
- ensuring that the testing is done by an ISTA-accredited laboratory;
- the check-sampling programme.

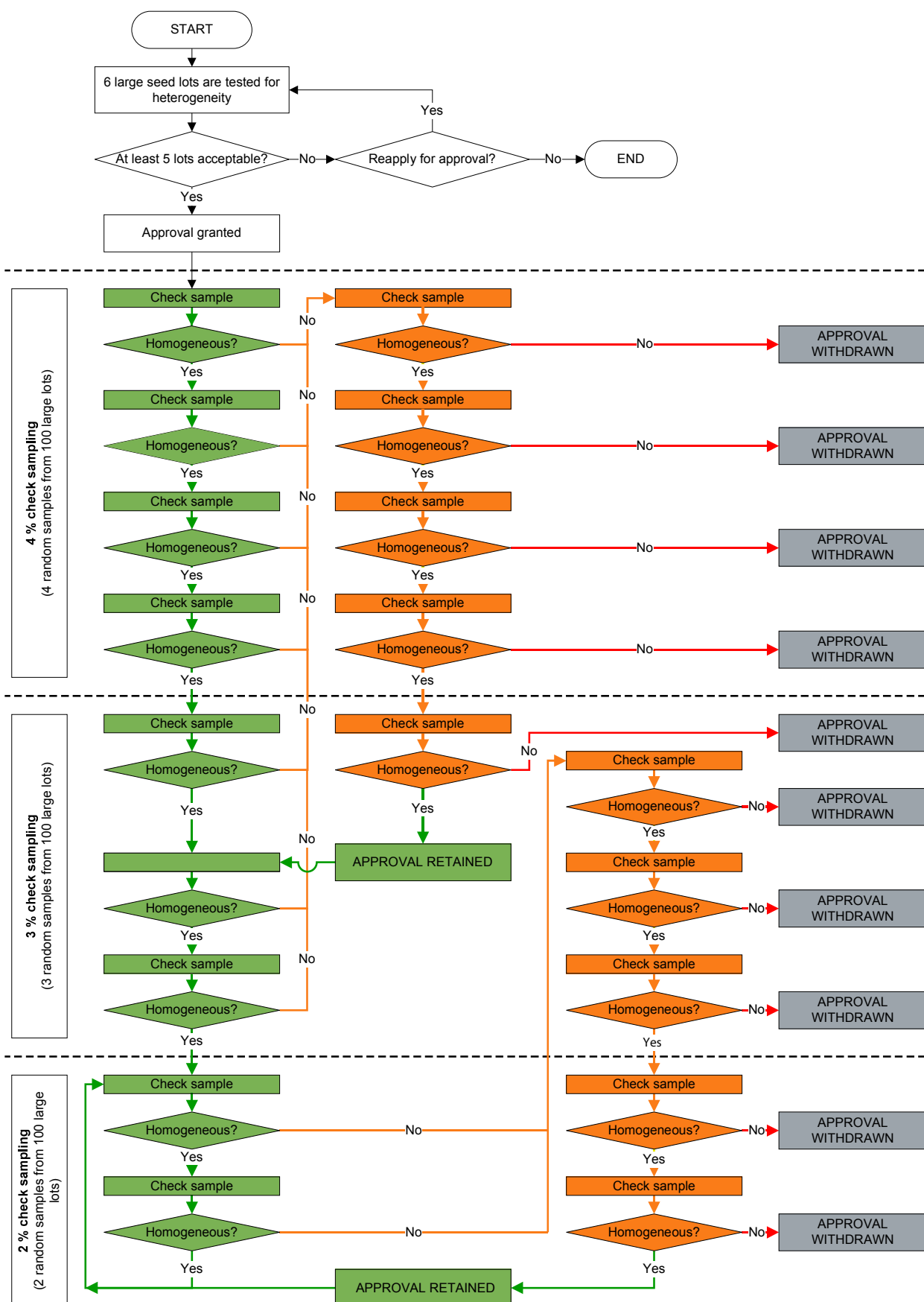


Figure 2.1. Flow chart describing the approval procedure and check-sampling programme with regard to large seed lots of Poaceae species (2.5.4.2.2–4).

2.5.4.3 Marking/labelling and sealing of containers

The seed lot must be in marked/labelled containers which are self-sealing, sealed (or capable of being sealed) or under the control of the seed sampler.

Where the seed lot is already marked/labelled and sealed before sampling, the seed sampler must verify the marking/labelling and sealing on the containers. Otherwise the sampler has to mark/label the containers and must seal every container before the seed lot leaves their control.

The samplers are personally responsible for the seals, labels and bags supplied to them and it is their duty to ensure that primary, composite or submitted samples must never be left in the hands of persons not authorised by the seed testing laboratory unless they are sealed in such a way that they cannot be tampered with.

2.5.4.4 Sampling from the seed lot

For sampling from the seed lot methods listed under 2.5.1 must be used. Automatic seed samplers must be approved by the ISTA seed testing laboratory according to the 'Protocol for the approval of automatic seed samplers' as approved by the ISTA membership and published on the ISTA website.

An Orange International Seed Lot Certificate issued on a seed lot (see 2.2.1) is still valid after re-packaging the seed lot in new containers provided that:

- The identity of the seed in the initial seed lot is preserved.
- The seed lot designation (see 2.2.11) is not changed.
- The moving of the seed into the new containers is done under the control of an ISTA seed sampler.
- There is no processing of the seed during filling of the new containers.

2.5.4.5 Submitted sample

The minimum sizes of submitted samples are as follows:

- If a determination of other seeds by number is required: the weight prescribed in Table 2C, column 3;
- or**
- If a determination of other seeds by number is not required: the weight prescribed for the working sample for purity analysis in Table 2C, column 4, or in 3.5.1.

If the sample is smaller than prescribed above, the sampler must be notified accordingly and analysis withheld until sufficient seed is received in a single submitted sample. This also applies to the weights of the exceptions listed below. For certain tests or under certain conditions, the following exceptions apply:

- For coated seeds, if a determination of other seeds by number or size grading is required: the number of seeds indicated in Table 2D, Parts 1 and 2, column 2.
- For coated seeds, if a determination of other seeds by number or size grading is not required: the number of seeds indicated for the working sample for purity analysis in Table 2D, Parts 1 and 2, column 3.
- For moisture determination of species that must be ground (see Table 9A): 100 g. For all other species: 50 g.
When moisture meters are to be used for testing, a larger sample size may be necessary. Contact the accredited ISTA laboratory for specific instructions.
- For verification of species and variety: as prescribed in Chapter 8.
- For germination or viability tests of small seed lots (2.2.14): the number of seeds required to complete one of these tests plus 25 seeds for identity assurance.
- For determination of other seeds of small seed lots (2.2.14): the amount necessary to complete this test according to Chapter 4.

The submitted sample must be sealed and labelled or marked.

2.5.4.6 Sample reduction

For sample reduction, methods listed under 2.5.2.2 must be used.

2.5.4.7 Storage of submitted samples after testing

Submitted samples on which ISTA Certificates have been issued must be stored. In the case of small seed lots (see 2.2.14), the remainder of the submitted sample, minus 25 seeds for assurance of identity, may be sent back to the applicant. The seed testing laboratory cannot be held responsible for any deterioration of the sample during storage.

2.6 Calculation and expression of results

No specific calculation or expression of results required except under 2.9 for heterogeneity tests.

2.7 Reporting of results

No specific calculation or expression of results required except under 2.9 for heterogeneity tests.

2.8 Tables for lot size and sample sizes

Table 2C is referred to in various chapters of the ISTA Rules and indicates weights of lots and samples for different species, and the specific names to be used in reporting test results. Each sample size is derived from a nominal thousand-seed weight (TSW) for each species which, on the available evidence, is expected to be adequate for the majority of samples tested.

Where a weight is not given in the table and a count of other species is requested, the submitted sample must contain a minimum of 25 000 seeds.

Note 1: Names with an asterisk are not included in the *ISTA List of Stabilised Plant Names*. Names without an asterisk are included in the *ISTA List of Stabilised Plant Names* (but not the synonym which follows some of these names), or, in the case of generic names (e.g. *Pyrus* spp.) conserved by the International Botanical Congress and listed in the *International Code of Nomenclature*. Changes in the Stabilised List agreed at the 2019 ISTA Congress are included in this version of Table 2C. Where plant names have been changed, the old name is included with a cross reference to the new name. This applies only to 2019 Congress changes; previous cross references have been removed.

Note 2: For all species the maximum seed lot size stated can be exceeded by no more than 5 %, except for:

- seed being transported loose in bulk containers. The conditions under which this exception may be permitted are stated in Chapter 17;
- seed pellets, seed granules, seed tapes or seed mats (see 2.5.4.1);
- species of Poaceae listed in Table 2C Part 1 (see 2.5.4.2).

For production plants approved under 2.5.4.2, the maximum seed lot weight for Poaceae species listed in Table 2C Part 1 is 25 000 kg (with a 5 % tolerance).

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>Abelmoschus esculentus</i> (L.) Moench	20 000	1 000	140	1 000
<i>Achillea millefolium</i> L.	10 000	5	0.5	5
<i>Aeschynomene americana</i> L.	10 000	120	12	120
<i>Agropyron cristatum</i> (L.) Gaertn.	10 000	40	4	40
<i>Agropyron desertorum</i> (Fisch. ex Link) Schult.	10 000	60	6	60
<i>Agrostis canina</i> L.	10 000	5	0.25	2.5
<i>Agrostis capillaris</i> L.	10 000	5	0.25	2.5
<i>Agrostis gigantea</i> Roth	10 000	5	0.25	2.5
<i>Agrostis stolonifera</i> L. (includes <i>A. palustris</i> Hudson)	10 000	5	0.25	2.5
<i>Allium cepa</i> L.	10 000	80	8	80
<i>Allium fistulosum</i> L.	10 000	50	5	50
<i>Allium porrum</i> L.	10 000	70	7	70
<i>Allium schoenoprasum</i> L.	10 000	30	3	30
<i>Allium tuberosum</i> Rottler ex Spreng.	10 000	100	10	100
<i>Alopecurus pratensis</i> L.	10 000	30	3	30
<i>Alysicarpus vaginalis</i> (L.) DC.	10 000	40	4	40
<i>Andropogon gayanus</i> Kunth	10 000	80	8	80
<i>Andropogon gerardi</i> Vitman	10 000	70	7	70
<i>Andropogon hallii</i> Hack.	10 000	100	10	100
<i>Anethum graveolens</i> L.	10 000	40	4	40
<i>Anthoxanthum odoratum</i> L.	10 000	20	2	20
<i>Anthriscus cerefolium</i> (L.) Hoffm.	10 000	60	6	60
<i>Anthyllis vulneraria</i> L.	10 000	60	6	60
<i>Apium graveolens</i> L.	10 000	10	1	10
<i>Arachis hypogaea</i> L.	30 000	1 000	1 000	1 000
<i>Arctium lappa</i> L.	10 000	50	5	50
<i>Arrhenatherum elatius</i> (L.) P.Beauv. ex J.Presl & C.Presl	10 000	80	8	80
<i>Asparagus officinalis</i> L.	20 000	1 000	100	1 000
<i>Astragalus cicer</i> L.	10 000	90	9	90
<i>Astrebula lappacea</i> (Lindl.) Domin	10 000	200	20	200
<i>Atriplex hortensis</i> L.	5 000	10	2.5	–
<i>Atropa belladonna</i> L.	10 000	30	3	30
<i>Avena nuda</i> L.	30 000	1 000	120	1 000
<i>Avena sativa</i> L.	30 000	1 000	120	1 000
<i>Avena strigosa</i> Schreb.	30 000	500	50	500
<i>Avenella flexuosa</i> (L.) Parl. (previously <i>Deschampsia flexuosa</i> (L.) Trin.)	10 000	10	1	10
<i>Axonopus compressus</i> (Sw.) P.Beauv.	10 000	10	1	10
<i>Axonopus fissifolius</i> (Raddi) Kuhlman.	10 000	10	1	10
<i>Beckmannia eruciformis</i> (L.) Host	10 000	20	2	20
<i>Beta vulgaris</i> L. (multi-germ varieties)	20 000	500	50	500
<i>Beta vulgaris</i> L. (mono-germ varieties)	20 000	500	30	300
<i>Borago officinalis</i> L.	10 000	450	45	450
<i>Bothriochloa insculpta</i> (Hochst. ex A.Rich.) A.Camus	10 000	20	2	20
<i>Bothriochloa pertusa</i> (L.) A.Camus	10 000	10	1	10
<i>Bouteloua gracilis</i> (Kunth) Lag. ex Griffiths	10 000	60	6	60
(<i>Brachiaria brizantha</i> (Hochst. ex A.Rich.) Stapf see <i>Urochloa brizantha</i> (Hochst. ex A.Rich.) R.D.Webster)				
(<i>Brachiaria decumbens</i> Stapf see <i>Urochloa decumbens</i> (Stapf) R.D.Webster)				
(<i>Brachiaria humidicola</i> (Rendle) Schweick. see <i>Urochloa humidicola</i> (Rendle) Morrone & Zuloaga)				

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>(Brachiaria mutica</i> (Forssk.) Stapf see <i>Urochloa mutica</i> (Forssk.) T.Q.Nguyen)				
<i>(Brachiaria ramosa</i> (L.) Stapf see <i>Urochloa ramosa</i> (L.) T.Q.Nguyen)				
<i>(Brachiaria ruziziensis</i> R.Germ. & C.M.Evrard see <i>Urochloa ruziziensis</i> (R.Germ. & C.M.Evrard) Crins)				
<i>Brassica carinata</i> A.Braun	10 000	100	10	100
<i>Brassica juncea</i> (L.) Czern.	10 000	40	4	40
<i>Brassica napus</i> L.	10 000	100	10	100
<i>Brassica napus</i> L. var. <i>napobrassica</i> (L.) Rchb.*	10 000	100	10	100
<i>Brassica nigra</i> (L.) W.D.J.Koch	10 000	40	4	40
<i>Brassica oleracea</i> L. (all varieties)	10 000	100	10	100
<i>Brassica rapa</i> L. (includes <i>B. campestris</i> L.)	10 000	70	7	70
<i>Bromus arvensis</i> L.	10 000	60	6	60
<i>(Bromus carinatus</i> Hook. & Arn. see <i>Bromus carinatus</i> Hook. & Arn. var. <i>carinatus</i>)				
<i>Bromus carinatus</i> Hook. & Arn. var. <i>carinatus</i> (previously <i>Bromus carinatus</i> Hook. & Arn.)	10 000	200	20	200
<i>Bromus carinatus</i> Hook. & Arn. var. <i>marginatus</i> (Steud.) Barkworth & Anderton (previously <i>Bromus marginatus</i> Steud.)	10 000	200	20	200
<i>Bromus catharticus</i> Vahl	10 000	200	20	200
<i>Bromus erectus</i> Huds.	10 000	100	10	100
<i>Bromus hordeaceus</i> L.	10 000	50	5	50
<i>Bromus inermis</i> Leyss.	10 000	90	9	90
<i>(Bromus marginatus</i> Steud. see <i>Bromus carinatus</i> Hook. & Arn. var. <i>marginatus</i> (Steud.) Barkworth & Anderton)				
<i>Bromus riparius</i> Rehmman	10 000	90	9	90
<i>Bromus sitchensis</i> Trin.	10 000	200	20	200
<i>Cajanus cajan</i> (L.) Huth	20 000	1 000	300	1 000
<i>Calopogonium mucunoides</i> Desv.	20 000	400	40	400
<i>Camelina sativa</i> (L.) Crantz	10 000	40	4	40
<i>Cannabis sativa</i> L.	10 000	600	60	600
<i>Capsicum</i> spp.	10 000	150	15	150
<i>Carthamus tinctorius</i> L.	25 000	900	90	900
<i>Carum carvi</i> L.	10 000	80	8	80
<i>Cenchrus ciliaris</i> L. (fascicles)	10 000	60	6	60
<i>(Cenchrus setiger</i> Vahl see <i>Cenchrus setigerus</i> Vahl)				
<i>Cenchrus setigerus</i> Vahl (previously <i>Cenchrus setiger</i> Vahl)	20 000	150	15	150
<i>Centrosema molle</i> Mart. ex Benth.	20 000	600	60	600
<i>Centrosema pascuorum</i> Mart. ex Benth.	20 000	550	55	550
<i>Chamaecrista rotundifolia</i> (Pers.) Greene	10 000	100	10	100
<i>Chenopodium quinoa</i> Willd.	10 000	100	10	100
<i>Chloris gayana</i> Kunth	10 000	10	1	10
<i>Cicer arietinum</i> L.	30 000	1 000	1 000	1 000
<i>Cichorium endivia</i> L.	10 000	40	4	40
<i>Cichorium intybus</i> L.	10 000	50	5	50
<i>Citrullus lanatus</i> (Thunb.) Matsum. & Nakai	20 000	1 000	250	1 000
<i>Claytonia perfoliata</i> Donn ex Willd.	10 000	20	2	20
<i>Corchorus capsularis</i> L.	10 000	150	15	150
<i>Corchorus olitorius</i> L.	10 000	150	15	150
<i>Coriandrum sativum</i> L.	10 000	400	40	400

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>(Crambe abyssinica</i> Hochst. ex R.E.Fr. see <i>Crambe hispanica</i> L. subsp. <i>abyssinica</i> (Hochst. ex R.E.Fr.) Prina)				
<i>Crambe hispanica</i> L. subsp. <i>abyssinica</i> (Hochst. ex R.E.Fr.) Prina (previously <i>Crambe abyssinica</i> Hochst. ex R.E.Fr.)	10 000	200	20	200
<i>Crotalaria brevidens</i> Benth. (includes <i>Crotalaria intermedia</i> Kotschy)	10 000	150	15	150
<i>Crotalaria juncea</i> L.	10 000	700	70	700
<i>Crotalaria lanceolata</i> E.Mey.	10 000	70	7	70
<i>Crotalaria pallida</i> Aiton	10 000	150	15	150
<i>Crotalaria spectabilis</i> Roth	10 000	350	35	350
<i>Cucumis melo</i> L.	10 000	150	70	–
<i>Cucumis sativus</i> L.	10 000	150	70	–
<i>Cucumis</i> spp.	10 000	150	70	–
<i>Cucurbita maxima</i> Duchesne	20 000	1 000	700	1 000
<i>Cucurbita moschata</i> Duchesne	10 000	350	180	–
<i>Cucurbita pepo</i> L.	20 000	1 000	700	1 000
<i>Cucurbita</i> spp.	10 000	350	180	–
<i>Cucurbita</i> hybrids	10 000	350	180	–
<i>Cuminum cyminum</i> L.	10 000	60	6	60
<i>Cyamopsis tetragonoloba</i> (L.) Taub.	20 000	1 000	100	1 000
<i>Cynara cardunculus</i> L.	10 000	900	90	900
<i>Cynodon dactylon</i> (L.) Pers.	10 000	10	1	10
<i>Cynosurus cristatus</i> L.	10 000	20	2	20
<i>Dactylis glomerata</i> L.	10 000	30	3	30
<i>Daucus carota</i> L.	10 000	30	3	30
<i>Deschampsia cespitosa</i> (L.) P.Beauv.	10 000	10	1	10
<i>(Deschampsia flexuosa</i> (L.) Trin. see <i>Avenella flexuosa</i> (L.) Parl.)				
<i>Desmodium intortum</i> (Mill.) Urb.	10 000	40	4	40
<i>Desmodium uncinatum</i> (Jacq.) DC.	20 000	120	12	120
<i>Dichanthium aristatum</i> (Poir.) C.E.Hubb.	10 000	30	3	30
<i>Dichondra micrantha</i> Urb.	10 000	50	5	50
<i>Digitaria eriantha</i> Steud. (includes <i>Digitaria decumbens</i> Stent)	10 000	12	1.2	12
<i>Echinochloa crus-galli</i> (L.) P.Beauv.	10 000	80	8	80
<i>Ehrharta calycina</i> Sm.	10 000	40	4	40
<i>Eleusine coracana</i> (L.) Gaertn.	10 000	60	6	60
<i>Elymus lanceolatus</i> (Scribn. & J.G.Sm.) Gould	10 000	80	8	80
<i>Elymus repens</i> (L.) Gould (previously <i>Elytrigia repens</i> (L.) Desv. ex Nevski)	10 000	100	10	100
<i>Elymus trachycaulus</i> (Link) Gould ex Shinnars	10 000	80	8	80
<i>(Elytrigia elongata</i> (Host) Nevski see <i>Thinopyrum elongatum</i> (Host) D.R.Dewey)				
<i>(Elytrigia intermedia</i> (Host) Nevski see <i>Thinopyrum intermedium</i> (Host) Barkworth & D.R.Dewey)				
<i>(Elytrigia repens</i> (L.) Desv. ex Nevski see <i>Elymus repens</i> (L.) Gould)				
<i>Eragrostis curvula</i> (Schrad.) Nees	10 000	10	1	10
<i>Eragrostis tef</i> (Zuccagni) Trotter	10 000	10	1	10
<i>(Eruca sativa</i> Mill. see <i>Eruca vesicaria</i> (L.) Cav. subsp. <i>sativa</i> (Mill.) Thell.)				

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>Eruca vesicaria</i> (L.) Cav. subsp. <i>sativa</i> (Mill.) Thell. (previously <i>Eruca sativa</i> Mill.)	10 000	40	4	40
<i>Fagopyrum esculentum</i> Moench	10 000	600	60	600
<i>Festuca arundinacea</i> Schreb.	10 000	50	5	50
<i>Festuca filiformis</i> Pourr.	10 000	25	2.5	25
<i>Festuca heterophylla</i> Lam.	10 000	60	6	60
<i>Festuca ovina</i> L. (all varieties)	10 000	25	2.5	25
<i>Festuca pratensis</i> Huds.	10 000	50	5	50
<i>Festuca rubra</i> L. s.l. (all varieties)	10 000	30	3	30
<i>Festuca trachyphylla</i> (Hack.) Hack. (synonym <i>Festuca brevipila</i> R.Tracey)	10 000	25	2.5	25
✱ <i>Festulolium</i> Asch. & Graebn.	10 000	60	6	60
<i>Foeniculum vulgare</i> Mill.	10 000	180	18	180
<i>Fragaria</i> spp.	10 000	10	1	10
<i>Galega orientalis</i> Lam.	10 000	200	20	200
<i>Glycine max</i> (L.) Merr.	30 000	1 000	500	1 000
<i>Gossypium</i> spp.	25 000	1 000	350	1 000
<i>Hedysarum coronarium</i> L. (fruit)	10 000	300	30	300
<i>Hedysarum coronarium</i> L. (seed)	10 000	120	12	120
<i>Helianthus annuus</i> L.	25 000	1 000	200	1 000
<i>Hibiscus cannabinus</i> L.	10 000	700	70	700
<i>Holcus lanatus</i> L.	10 000	10	1	10
<i>Hordeum vulgare</i> L. subsp. <i>vulgare</i> (previously <i>Hordeum vulgare</i> L.)	30 000	1 000	120	1 000
<i>Ipomoea aquatica</i> Forssk.	20 000	1 000	100	1 000
<i>Koeleria macrantha</i> (Ledeb.) Schult.	10 000	10	1	10
<i>Kummerowia stipulacea</i> (Maxim.) Makino	10 000	50	5	50
<i>Kummerowia striata</i> (Thunb.) Schindl.	10 000	40	4	40
<i>Lablab purpureus</i> (L.) Sweet	20 000	1 000	600	1 000
<i>Lactuca sativa</i> L.	10 000	30	3	30
<i>Lagenaria siceraria</i> (Molina) Standl.	20 000	1 000	500	1 000
<i>Lathyrus cicera</i> L.	20 000	1 000	140	1 000
<i>Lathyrus hirsutus</i> L.	10 000	700	70	700
<i>Lathyrus sativus</i> L.	20 000	1 000	450	1 000
<i>Lens culinaris</i> Medik.	30 000	600	60	600
<i>Lepidium sativum</i> L.	10 000	60	6	60
<i>Lespedeza juncea</i> (L. f.) Pers.	10 000	30	3	30
<i>Leucaena leucocephala</i> (Lam.) de Wit	20 000	1 000	100	1 000
<i>Linum usitatissimum</i> L.	10 000	150	15	150
<i>Listia bainesii</i> (Baker) B.-E.vanWyk & Boatwr.	10 000	10	1	10
<i>Lolium</i> ✱ <i>hybridum</i> Hausskn.	10 000	60	6	60
<i>Lolium multiflorum</i> Lam.	10 000	60	6	60
<i>Lolium perenne</i> L.	10 000	60	6	60
<i>Lolium rigidum</i> Gaudin	10 000	60	6	60
<i>Lotus corniculatus</i> L.	10 000	30	3	30
<i>Lotus tenuis</i> Waldst. & Kit. ex Willd.	10 000	30	3	30
<i>Lotus uliginosus</i> Schkuhr	10 000	20	2	20
<i>Luffa acutangula</i> (L.) Roxb.	20 000	1 000	400	1 000
<i>Luffa aegyptiaca</i> Mill.	20 000	1 000	250	1 000
<i>Lupinus albus</i> L.	30 000	1 000	450	1 000
<i>Lupinus angustifolius</i> L.	30 000	1 000	450	1 000

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>Lupinus luteus</i> L.	30 000	1 000	450	1 000
<i>Macroptilium atropurpureum</i> (DC.) Urb.	20 000	350	35	350
<i>Macroptilium lathyroides</i> (L.) Urb.	20 000	200	20	200
<i>Macrotyloma axillare</i> (E.Mey.) Verdc.	20 000	250	25	250
<i>Macrotyloma uniflorum</i> (Lam.) Verdc.	20 000	800	80	800
<i>Medicago arabica</i> (L.) Huds. (in burr)	10 000	600	60	600
<i>Medicago arabica</i> (L.) Huds. (out of burr)	10 000	50	5	50
<i>Medicago italica</i> (Mill.) Fiori (includes <i>Medicago tornata</i> (L.) Mill.)	10 000	100	10	100
<i>Medicago littoralis</i> Rohde ex Loisel.	10 000	70	7	70
<i>Medicago lupulina</i> L.	10 000	50	5	50
<i>Medicago orbicularis</i> (L.) Bartal.	10 000	80	8	80
<i>Medicago polymorpha</i> L.	10 000	70	7	70
<i>Medicago rugosa</i> Desr.	10 000	180	18	180
<i>Medicago sativa</i> L.	10 000	50	5	50
<i>Medicago scutellata</i> (L.) Mill.	10 000	400	40	400
<i>Medicago truncatula</i> Gaertn.	10 000	100	10	100
<i>Megathyrsus maximus</i> (Jacq.) B.K.Simon & S.W.L.Jacobs (previously <i>Panicum maximum</i> Jacq.)	10 000	20	2	20
<i>Melilotus albus</i> Medik.	10 000	50	5	50
<i>Melilotus indicus</i> (L.) All.	10 000	50	5	50
<i>Melilotus officinalis</i> (L.) Lam.	10 000	50	5	50
<i>Melinis minutiflora</i> P.Beauv.	10 000	5	0.5	5
<i>Momordica charantia</i> L.	20 000	1 000	450	1 000
<i>Mucuna pruriens</i> (L.) DC.	20 000	1 000	1 000	1 000
<i>Nasturtium officinale</i> W.T.Aiton	10 000	5	0.5	5
<i>Neonotonia wightii</i> (Wight & Arn.) J.A.Lackey	10 000	150	15	150
<i>Neustanthus phaseoloides</i> (Roxb.) Benth. (previously <i>Pueraria phaseoloides</i> (Roxb.) Benth.)	20 000	300	30	300
<i>Nicotiana tabacum</i> L.	10 000	5	0.5	5
<i>Ocimum basilicum</i> L.	10 000	40	4	40
<i>Oenothera biennis</i> L.	10 000	10	1	10
<i>Oloptum miliaceum</i> (L.) Röser & Hamasha (previously <i>Piptatherum miliaceum</i> (L.) Coss.)	10 000	20	2	20
<i>Onobrychis viciifolia</i> Scop. (fruit)	10 000	600	60	600
<i>Onobrychis viciifolia</i> Scop. (seed)	10 000	400	40	400
<i>Origanum majorana</i> L.	10 000	5	0.5	5
<i>Origanum vulgare</i> L.	10 000	5	0.5	5
<i>Ornithopus compressus</i> L.	10 000	120	12	120
<i>Ornithopus sativus</i> Brot.	10 000	90	9	90
<i>Oryza sativa</i> L.	30 000	700	70	700
<i>Panicum antidotale</i> Retz.	10 000	20	2	20
<i>Panicum coloratum</i> L.	10 000	20	2	20
(<i>Panicum maximum</i> Jacq. see <i>Megathyrsus maximus</i> (Jacq.) B.K.Simon & S.W.L.Jacobs)				
<i>Panicum miliaceum</i> L.	10 000	150	15	150
<i>Panicum virgatum</i> L.	10 000	30	3	30
<i>Papaver somniferum</i> L.	10 000	10	1	10
<i>Paspopyrum smithii</i> (Rydb.) Barkworth & D.R.Dewey	10 000	150	15	150
<i>Paspalum dilatatum</i> Poir.	10 000	50	5	50
<i>Paspalum notatum</i> Flügge	10 000	70	7	70
<i>Paspalum plicatulum</i> Michx.	10 000	40	4	40

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>Paspalum scrobiculatum</i> L.	10 000	80	8	80
<i>Paspalum urvillei</i> Steud.	10 000	30	3	30
<i>Paspalum virgatum</i> L.	10 000	30	3	30
<i>Pastinaca sativa</i> L.	10 000	100	10	100
<i>Pennisetum clandestinum</i> Hochst. ex Chiov.	10 000	70	7	70
<i>Pennisetum glaucum</i> (L.) R.Br.	10 000	150	15	150
<i>Petroselinum crispum</i> (Mill.) Fuss	10 000	40	4	40
<i>Phacelia tanacetifolia</i> Benth.	10 000	50	5	50
<i>Phalaris aquatica</i> L.	10 000	40	4	40
<i>Phalaris arundinacea</i> L.	10 000	30	3	30
<i>Phalaris canariensis</i> L.	10 000	200	20	200
<i>Phaseolus coccineus</i> L.	30 000	1 000	1 000	1 000
<i>Phaseolus lunatus</i> L.	30 000	1 000	1 000	1 000
<i>Phaseolus vulgaris</i> L.	30 000	1 000	700	1 000
<i>Phleum nodosum</i> L.	10 000	10	1	10
<i>Phleum pratense</i> L.	10 000	10	1	10
<i>Physalis pubescens</i> L.	10 000	20	2	20
<i>Pimpinella anisum</i> L.	10 000	70	7	70
<i>(Piptatherum miliaceum</i> (L.) Coss. see <i>Oloptum miliaceum</i> (L.) Röser & Hamasha)				
<i>Pisum sativum</i> L. s.l.	30 000	1 000	900	1 000
<i>Plantago lanceolata</i> L.	10 000	60	6	60
<i>Poa annua</i> L.	10 000	10	1	10
<i>Poa bulbosa</i> L.	10 000	30	3	30
<i>Poa compressa</i> L.	10 000	5	0.5	5
<i>Poa nemoralis</i> L.	10 000	5	0.5	5
<i>Poa palustris</i> L.	10 000	5	0.5	5
<i>Poa pratensis</i> L.	10 000	5	1	5
<i>Poa secunda</i> J.Presl (includes <i>Poa ampla</i> Merr.)	10 000	15	1.5	15
<i>Poa trivialis</i> L.	10 000	5	1	5
<i>Portulaca oleracea</i> L.	10 000	5	0.5	5
<i>Psathyrostachys juncea</i> (Fisch.) Nevski	10 000	60	6	60
<i>Pseudoroegneria spicata</i> (Pursh) Å.Löve	10 000	80	8	80
<i>Psophocarpus tetragonolobus</i> (L.) DC.	20 000	1 000	1 000	1 000
<i>(Pueraria lobata</i> (Willd.) Ohwi see <i>Pueraria montana</i> (Lour.) Merr. var. <i>lobata</i> (Willd.) Maesen & S.M.Almeida ex Sanjappa & Predeep)				
<i>Pueraria montana</i> (Lour.) Merr. var. <i>lobata</i> (Willd.) Maesen & S.M.Almeida ex Sanjappa & Predeep (previously <i>Pueraria lobata</i> (Willd.) Ohwi)	10 000	350	35	350
<i>(Pueraria phaseoloides</i> (Roxb.) Benth. see <i>Neustanthus phaseoloides</i> (Roxb.) Benth.)				
<i>Raphanus sativus</i> L.	10 000	300	30	300
<i>Rheum ×rhabarbarum</i> auct., non L. (previously <i>Rheum rhaponticum</i> L.)	10 000	450	45	450
<i>(Rheum rhaponticum</i> L. see <i>Rheum ×rhabarbarum</i> auct., non L.)				
<i>Ricinus communis</i> L.	20 000	1 000	500	1 000
<i>Rosmarinus officinalis</i> L.	10 000	30	3	30
<i>Rumex acetosa</i> L.	10 000	30	3	30
<i>Salvia hispanica</i> L.	10 000	35	3.5	35
<i>Sanguisorba minor</i> Scop.	10 000	250	25	250

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>Satureja hortensis</i> L.	10 000	20	2	20
<i>Schizachyrium scoparium</i> (Michx.) Nash	10 000	50	5	50
<i>Scorzonera hispanica</i> L.	10 000	300	30	300
<i>Secale cereale</i> L.	30 000	1 000	120	1 000
<i>Securigera varia</i> (L.) Lassen	10 000	100	10	100
<i>Sesamum indicum</i> L.	10 000	70	7	70
<i>Setaria italica</i> (L.) P.Beauv.	10 000	90	9	90
<i>Setaria sphacelata</i> (Schumach.) Stapf & C.E.Hubb.	10 000	30	3	30
<i>Sinapis alba</i> L.	10 000	200	20	200
<i>Solanum</i> (sect. <i>Lycopersicon</i>) spp.	10 000	15	7	–
<i>Solanum</i> (sect. <i>Lycopersicon</i>) hybrids	10 000	15	7	–
<i>Solanum lycopersicum</i> L.	10 000	15	7	–
<i>Solanum melongena</i> L.	10 000	150	15	150
<i>Solanum nigrum</i> L.	10 000	25	2.5	25
<i>Solanum tuberosum</i> L.	10 000	25	10	–
<i>Sorghastrum nutans</i> (L.) Nash	10 000	70	7	70
<i>Sorghum xalimum</i> Parodi	30 000	200	20	200
<i>(Sorghum bicolor</i> (L.) Moench see <i>Sorghum bicolor</i> (L.) Moench subsp. <i>bicolor</i>)				
<i>Sorghum bicolor</i> (L.) Moench subsp. <i>bicolor</i> (previously <i>Sorghum bicolor</i> (L.) Moench)	30 000	900	90	900
<i>Sorghum bicolor</i> (L.) Moench subsp. <i>drummondii</i> (Steud.) de Wet ex Davidse (previously <i>Sorghum sudanense</i> (Piper) Stapf)	10 000	250	25	250
<i>Sorghum bicolor</i> (L.) Moench × <i>S. sudanense</i> (Piper) Stapf	30 000	300	30	300
<i>Sorghum halepense</i> (L.) Pers.	10 000	90	9	90
<i>(Sorghum sudanense</i> (Piper) Stapf see <i>Sorghum bicolor</i> (L.) Moench subsp. <i>drummondii</i> (Steud.) de Wet ex Davidse)				
<i>Spergula arvensis</i> L.	10 000	40	4	40
<i>Spinacia oleracea</i> L.	10 000	250	25	250
<i>Stylosanthes guianensis</i> (Aubl.) Sw.	10 000	70	7	70
<i>Stylosanthes hamata</i> (L.) Taub.	10 000	70	7	70
<i>Stylosanthes humilis</i> Kunth	10 000	70	7	70
<i>Stylosanthes scabra</i> Vogel	10 000	80	8	80
<i>Taraxacum officinale</i> F.H.Wigg., s.l.	10 000	30	3	30
<i>Tetragonia tetragonoides</i> (Pall.) Kuntze	20 000	1 000	200	1 000
<i>Thinopyrum elongatum</i> (Host) D.R.Dewey (previously <i>Elytrigia elongata</i> (Host) Nevs)	10 000	200	20	200
<i>Thinopyrum intermedium</i> (Host) Barkworth & D.R.Dewey (previously <i>Elytrigia intermedia</i> (Host) Nevski)	10 000	150	15	150
<i>Thymus vulgaris</i> L.	10 000	5	0.5	5
<i>Tragopogon porrifolius</i> L.	10 000	400	40	400
<i>Trifolium alexandrinum</i> L.	10 000	60	6	60
<i>Trifolium campestre</i> Schreb.	10 000	5	0.5	5
<i>Trifolium dubium</i> Sibth.	10 000	20	2	20
<i>Trifolium fragiferum</i> L.	10 000	40	4	40
<i>Trifolium glomeratum</i> L.	10 000	10	1	10
<i>Trifolium hirtum</i> All.	10 000	70	7	70
<i>Trifolium hybridum</i> L.	10 000	20	2	20
<i>Trifolium incarnatum</i> L.	10 000	80	8	80
<i>Trifolium lappaceum</i> L.	10 000	20	2	20

Table 2C Part 1. Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>Trifolium michelianum</i> Savi (includes <i>Trifolium balansae</i> Boiss.)	10 000	20	2	20
<i>Trifolium pratense</i> L.	10 000	50	5	50
<i>Trifolium repens</i> L.	10 000	20	2	20
<i>Trifolium resupinatum</i> L.	10 000	20	2	20
<i>Trifolium semipilosum</i> Fresen.	10 000	20	2	20
<i>Trifolium squarrosum</i> L.	10 000	150	15	150
<i>Trifolium subterraneum</i> L.	10 000	250	25	250
<i>Trifolium vesiculosum</i> Savi	10 000	30	3	30
<i>Trigonella foenum-graecum</i> L.	10 000	450	45	450
<i>Trisetum flavescens</i> (L.) P.Beauv.	10 000	5	0.5	5
✱ <i>Triticosecale</i> Wittm. ex A.Camus	30 000	1 000	120	1 000
(<i>Triticum aestivum</i> L. see <i>Triticum aestivum</i> L. subsp. <i>aestivum</i>)				
<i>Triticum aestivum</i> L. subsp. <i>aestivum</i> (previously <i>Triticum aestivum</i> L.)	30 000	1 000	120	1 000
<i>Triticum aestivum</i> L. subsp. <i>spelta</i> (L.) Thell. (previously <i>Triticum spelta</i> L.)	30 000	1 000	270	1 000
(<i>Triticum dicoccon</i> Schrank see <i>Triticum turgidum</i> L. subsp. <i>dicoccon</i> (Schrank) Thell.)				
(<i>Triticum durum</i> Desf. see <i>Triticum turgidum</i> L. subsp. <i>durum</i> (Desf.) van Slageren)				
(<i>Triticum spelta</i> L. see <i>Triticum aestivum</i> L. subsp. <i>spelta</i> (L.) Thell.)				
<i>Triticum turgidum</i> L. subsp. <i>dicoccon</i> (Schrank) Thell. (previously <i>Triticum dicoccon</i> Schrank)	30 000	1 000	270	1 000
<i>Triticum turgidum</i> L. subsp. <i>durum</i> (Desf.) van Slageren (previously <i>Triticum durum</i> Desf.)	30 000	1 000	120	1 000
<i>Urochloa brizantha</i> (Hochst. ex A.Rich.) R.D.Webster (previously <i>Brachiaria brizantha</i> (Hochst. ex A.Rich.) Stapf)	10 000	100	10	100
<i>Urochloa decumbens</i> (Stapf) R.D.Webster (previously <i>Brachiaria decumbens</i> Stapf)	10 000	100	10	100
<i>Urochloa humidicola</i> (Rendle) Morrone & Zuloaga (previously <i>Brachiaria humidicola</i> (Rendle) Schweick.)	10 000	100	10	100
<i>Urochloa mosambicensis</i> (Hack.) Dandy	10 000	30	3	30
<i>Urochloa mutica</i> (Forssk.) T.Q.Nguyen (previously <i>Brachiaria mutica</i> (Forssk.) Stapf)	10 000	30	3	30
<i>Urochloa ramosa</i> (L.) T.Q.Nguyen (previously <i>Brachiaria ramosa</i> (L.) Stapf)	10 000	90	9	90
<i>Urochloa ruziziensis</i> (R.Germ. & C.M.Evrard) Crins (previously <i>Brachiaria ruziziensis</i> R.Germ. & C.M.Evrard)	20 000	150	15	150
<i>Valerianella locusta</i> (L.) Laterr.	10 000	70	7	70
<i>Vicia benghalensis</i> L.	30 000	1 000	120	1 000
<i>Vicia ervilia</i> (L.) Willd.	30 000	1 000	120	1 000
<i>Vicia faba</i> L.	30 000	1 000	1 000	1 000
<i>Vicia narbonensis</i> L.	30 000	1 000	600	1 000
<i>Vicia pannonica</i> Crantz	30 000	1 000	120	1 000
<i>Vicia sativa</i> L. (includes <i>V. angustifolia</i> L.)	30 000	1 000	140	1 000
<i>Vicia villosa</i> Roth (includes <i>V. dasycarpa</i> Ten.)	30 000	1 000	100	1 000
<i>Vigna angularis</i> (Willd.) Ohwi & H.Ohashi	30 000	1 000	250	1 000
<i>Vigna marina</i> (Burm.) Merr.	30 000	800	80	800
<i>Vigna mungo</i> (L.) Hepper	30 000	1 000	700	1 000

**Table 2C Part 1.** Lot sizes and sample sizes: agricultural and vegetable seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working samples (g)	
			Purity analysis (3.5.1)	Other seeds by number (4.5.1)
1	2	3	4	5
<i>Vigna radiata</i> (L.) R.Wilczek	30 000	1 000	120	1 000
<i>Vigna subterranea</i> (L.) Verdc.	30 000	1 000	500	1 000
<i>Vigna unguiculata</i> (L.) Walp.	30 000	1 000	400	1 000
<i>Zea mays</i> L.	40 000	1 000	900	1 000
<i>Zoysia japonica</i> Steud.	10 000	10	1	10

Table 2C Part 2. Lot sizes and sample sizes: tree and shrub seeds

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Abies alba</i> Mill.	1 000	240	120
<i>Abies amabilis</i> Douglas ex J.Forbes	1 000	200	100
<i>Abies balsamea</i> (L.) Mill.	1 000	40	20
<i>Abies cephalonica</i> Loudon	1 000	360	180
<i>Abies cilicica</i> (Antoine & Kotschy) Carrière	1 000	1 000	500
<i>Abies concolor</i> (Gordon & Glend.) Lindl. ex Hildebr.	1 000	160	80
<i>Abies firma</i> Siebold & Zucc.	1 000	200	100
<i>Abies fraseri</i> (Pursh) Poir.	1 000	40	20
<i>Abies grandis</i> (Douglas ex D.Don) Lindl.	1 000	100	50
<i>Abies homolepis</i> Siebold & Zucc.	1 000	80	40
<i>Abies lasiocarpa</i> (Hook.) Nutt.	1 000	50	25
<i>Abies magnifica</i> A.Murray bis	1 000	400	200
<i>Abies nordmanniana</i> (Steven) Spach	1 000	360	180
<i>Abies numidica</i> de Lannoy ex Carrière	1 000	500	250
<i>Abies pinsapo</i> Boiss.	1 000	320	160
<i>Abies procera</i> Rehder	1 000	160	80
<i>Abies sachalinensis</i> (F.Schmidt) Mast.	1 000	60	30
<i>Abies veitchii</i> Lindl.	1 000	40	20
<i>Acacia</i> spp.	1 000	70	35
<i>Acer campestre</i> L.	1 000	400	200
<i>Acer negundo</i> L.	500	200	100
<i>Acer palmatum</i> Thunb.	500	100	50
<i>Acer platanoides</i> L.	500	700	350
<i>Acer pseudoplatanus</i> L.	500	600	300
<i>Acer rubrum</i> L.	500	100	50
<i>Acer saccharinum</i> L.	500	1 000	500
<i>Acer saccharum</i> Marshall	500	360	180
<i>Aesculus hippocastanum</i> L.	5 000	500 seeds	500 seeds
<i>Ailanthus altissima</i> (Mill.) Swingle	1 000	160	80
<i>Alnus cordata</i> (Loisel.) Duby	1 000	12	6
<i>Alnus glutinosa</i> (L.) Gaertn.	1 000	8	4
<i>Alnus incana</i> (L.) Moench	1 000	4	2
<i>Alnus rubra</i> Bong.	1 000	4	2
<i>Amorpha fruticosa</i> L.	1 000	1 000	150
<i>Berberis aquifolium</i> Pursh	1 000	60	30
<i>Betula papyrifera</i> Marshall	300	10	3

Table 2C Part 2. Lot sizes and sample sizes: tree and shrub seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Betula pendula</i> Roth	300	10	1
<i>Betula pubescens</i> Ehrh.	300	10	1
<i>Calocedrus decurrens</i> (Torr.) Florin	300	160	80
<i>Caragana arborescens</i> Lam.	1000	160	80
<i>Carica papaya</i> L.	1000	100	50
<i>Carpinus betulus</i> L.	1000	500	250
<i>Castanea sativa</i> Mill.	5000	500 seeds	500 seeds
<i>Catalpa</i> spp.*	1000	120	60
<i>Cedrela</i> spp.	1000	80	40
<i>Cedrus atlantica</i> (Endl.) G.Manetti ex Carrière	1000	400	200
<i>Cedrus deodara</i> (Roxb. ex D.Don) G.Don	1000	600	300
<i>Cedrus libani</i> A.Rich.	1000	400	200
<i>Chamaecyparis lawsoniana</i> A.Murray bis) Parl. (<i>Chamaecyparis nootkatensis</i> (D.Don) Spach see <i>Cupressus nootkatensis</i> D.Don)	1000	20	6
<i>Chamaecyparis obtusa</i> (Siebold & Zucc.) Endl.	1000	12	6
<i>Chamaecyparis pisifera</i> (Siebold & Zucc.) Endl.	1000	10	3
<i>Chamaecyparis thyoides</i> (L.) Britton <i>et al.</i>	1000	10	3
<i>Cornus mas</i> L.	1000	1000	600
<i>Cornus sanguinea</i> L.	1000	300	150
<i>Corylus avellana</i> L.	5000	500 fruits	500 fruits
<i>Corymbia citriodora</i> (Hook.) K.D.Hill & L.A.S.Johnson	1000	40	15
<i>Corymbia ficifolia</i> (F.Muell.) K.D.Hill & L.A.S.Johnson	1000	40	15
<i>Corymbia maculata</i> (Hook.) K.D.Hill & L.A.S.Johnson	1000	40	15
<i>Cotoneaster</i> spp.*	1000	40	20
<i>Crataegus monogyna</i> Jacq.	1000	400	200
<i>Cryptomeria japonica</i> (L. f.) D.Don	1000	20	10
<i>Cupressus arizonica</i> Greene	1000	60	30
<i>Cupressus macrocarpa</i> Hartw.	1000	40	20
<i>Cupressus nootkatensis</i> D.Don (previously <i>Chamaecyparis nootkatensis</i> (D.Don) Spach)	1000	20	10
<i>Cupressus sempervirens</i> L.	1000	40	20
<i>Cydonia oblonga</i> Mill.	1000	50	25
<i>Cytisus scoparius</i> (L.) Link	1000	40	20
<i>Elaeagnus angustifolia</i> L.	1000	800	400
<i>Eucalyptus astringens</i> (Maiden) Maiden	1000	40	15
<i>Eucalyptus botryoides</i> Sm.	1000	15	5
<i>Eucalyptus bridgesiana</i> R.T.Baker	1000	30	10
<i>Eucalyptus camaldulensis</i> Dehnh.	1000	15	5
<i>Eucalyptus cinerea</i> F.Muell. ex Benth.	1000	30	10
<i>Eucalyptus cladocalyx</i> F.Muell.	1000	40	15
<i>Eucalyptus cloeziana</i> F.Muell.	1000	40	15
<i>Eucalyptus cypellocarpa</i> L.A.S.Johnson	1000	30	10
<i>Eucalyptus dalrympleana</i> Maiden	1000	30	10
<i>Eucalyptus deanei</i> Maiden	1000	15	5
<i>Eucalyptus deglupta</i> Blume	1000	10	2
<i>Eucalyptus delegatensis</i> R.T.Baker	1000	40	15
<i>Eucalyptus elata</i> Dehnh.	1000	40	15
<i>Eucalyptus fastigata</i> H.Deane & Maiden	1000	40	15
<i>Eucalyptus glaucescens</i> Maiden & Blakely	1000	40	15
<i>Eucalyptus globulus</i> Labill. (includes <i>E. maidenii</i> F.Muell. and <i>E. saint-johnii</i> (R.T.Baker) R.T.Baker)	1000	60	20
<i>Eucalyptus grandis</i> W.Hill ex Maiden	1000	15	5
<i>Eucalyptus gunnii</i> Hook. f.	1000	15	5

Table 2C Part 2. Lot sizes and sample sizes: tree and shrub seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Eucalyptus largiflorens</i> F.Muell.	1 000	15	5
<i>Eucalyptus leucoxydon</i> F.Muell.	1 000	30	10
<i>Eucalyptus macrorhyncha</i> F.Muell. ex Benth.	1 000	40	15
<i>Eucalyptus mannifera</i> Mudie	1 000	15	5
<i>Eucalyptus melliodora</i> A.Cunn. ex Schauer	1 000	30	10
<i>Eucalyptus microtheca</i> F.Muell.	1 000	15	5
<i>Eucalyptus moluccana</i> Roxb.	1 000	30	10
<i>Eucalyptus muelleriana</i> A.W.Howitt	1 000	60	20
<i>Eucalyptus nitens</i> (H.Deane & Maiden) Maiden	1 000	30	10
<i>Eucalyptus pauciflora</i> Sieber ex Spreng. (includes <i>E. niphophila</i> Maiden & Blakely)	1 000	60	20
<i>Eucalyptus pilularis</i> Sm.	1 000	60	20
<i>Eucalyptus polybractea</i> R.T.Baker	1 000	60	20
<i>Eucalyptus radiata</i> Sieber ex DC.	1 000	40	15
<i>Eucalyptus regnans</i> F.Muell.	1 000	30	10
<i>Eucalyptus resinifera</i> Sm.	1 000	30	10
<i>Eucalyptus robusta</i> Sm.	1 000	15	5
<i>Eucalyptus rudis</i> Endl.	1 000	15	5
<i>Eucalyptus saligna</i> Sm.	1 000	15	5
<i>Eucalyptus sideroxylon</i> A.Cunn. ex Woolls	1 000	30	10
<i>Eucalyptus sieberi</i> L.A.S.Johnson	1 000	40	15
<i>Eucalyptus smithii</i> R.T.Baker	1 000	30	10
<i>Eucalyptus tereticornis</i> Sm.	1 000	15	5
<i>Eucalyptus viminalis</i> Labill.	1 000	30	10
<i>Euonymus europaeus</i> L.	1 000	200	100
<i>Fagus sylvatica</i> L.	5 000	1 000	600
<i>Fraxinus</i> spp.	1 000	400	200
<i>Ginkgo biloba</i> L.	5 000	500 seeds	500 seeds
<i>Gleditsia triacanthos</i> L.	1 000	800	400
<i>Ilex aquifolium</i> L.	1 000	200	90
<i>Juniperus communis</i> L. (berries)	1 000	300	150
<i>Juniperus communis</i> L. (seeds)	1 000	40	20
<i>Juniperus scopulorum</i> Sarg.	1 000	70	35
<i>Juniperus virginiana</i> L.	1 000	100	50
<i>Koeleria paniculata</i> Laxm.	1 000	800	400
<i>Laburnum alpinum</i> (Mill.) J.Presl	1 000	140	70
<i>Laburnum anagyroides</i> Medik.	1 000	140	70
<i>Larix decidua</i> Mill.	1 000	35	17
(Larix $\times$ eurolepis A.Henry see Larix $\times$ marschlinsii Coaz)			
<i>Larix gmelinii</i> (Rupr.) Rupr.	1 000	25	10
<i>Larix kaempferi</i> (Lamb.) Carrière	1 000	24	10
<i>Larix laricina</i> (D.Roi) K.Koch	1 000	25	10
<i>Larix $\times$ marschlinsii</i> Coaz (previously Larix $\times$ eurolepis A.Henry)	1 000	35	16
<i>Larix occidentalis</i> Nutt.	1 000	25	10
<i>Larix sibirica</i> Ledeb.	1 000	25	10
<i>Ligustrum vulgare</i> L.	1 000	100	50
<i>Liquidambar styraciflua</i> L.	300	30	15
<i>Liriodendron tulipifera</i> L.	1 000	180	90
<i>Malus</i> spp. (except <i>M. sargentii</i> , <i>M. sylvestris</i>)	1 000	50	25
<i>Malus sargentii</i> Rehder	1 000	24	12
<i>Malus sylvestris</i> (L.) Mill.	1 000	160	80
<i>Morus</i> spp.	1 000	20	5
<i>Nothofagus alpina</i> (Poepp. & Endl.) Oerst.	1 000	50	25

Table 2C Part 2. Lot sizes and sample sizes: tree and shrub seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Nothofagus obliqua</i> (Mirb.) Blume	1 000	60	30
<i>Picea abies</i> (L.) H.Karst.	1 000	40	20
<i>Picea engelmannii</i> Engelm.	1 000	16	8
<i>Picea glauca</i> (Moench) Voss	1 000	10	5
<i>Picea glehnii</i> (F.Schmidt) Mast.	1 000	25	9
<i>Picea jezoensis</i> (Siebold & Zucc.) Carrière	1 000	25	7
<i>Picea koyamae</i> Shiras.	1 000	25	9
<i>Picea mariana</i> (Mill.) Britton <i>et al.</i>	1 000	6	3
<i>Picea omorika</i> (Pančić) Purk.	1 000	25	8
<i>Picea orientalis</i> (L.) Link	1 000	30	15
<i>Picea polita</i> (Siebold & Zucc.) Carrière	1 000	80	40
<i>Picea pungens</i> Engelm.	1 000	30	15
<i>Picea rubens</i> Sarg.	1 000	25	9
<i>Picea sitchensis</i> (Bong.) Carrière	1 000	12	6
<i>Pinus albicaulis</i> Engelm.	1 000	700	350
<i>Pinus aristata</i> Engelm.	1 000	100	50
<i>Pinus banksiana</i> Lamb.	1 000	25	9
<i>Pinus brutia</i> Ten.	1 000	100	50
<i>Pinus canariensis</i> C.Sm.	1 000	60	30
<i>Pinus caribaea</i> Morelet	1 000	100	50
<i>Pinus cembra</i> L.	1 000	1 000	700
<i>Pinus cembroides</i> Zucc.	1 000	1 000	700
<i>Pinus clausa</i> (Chapm. ex Engelm.) Vasey ex Sarg.	1 000	40	20
<i>Pinus contorta</i> Douglas ex Loudon	1 000	25	9
<i>Pinus coulteri</i> D.Don	1 000	1 000	900
<i>Pinus densiflora</i> Siebold & Zucc.	1 000	60	30
<i>Pinus echinata</i> Mill.	1 000	50	25
<i>Pinus edulis</i> Engelm.	1 000	1 000	700
<i>Pinus elliotii</i> Engelm.	1 000	160	80
<i>Pinus flexilis</i> E.James	1 000	500	250
<i>Pinus glabra</i> Walter	1 000	80	40
<i>Pinus halepensis</i> Mill.	1 000	100	50
<i>Pinus heldreichii</i> Christ	1 000	120	60
<i>Pinus jeffreyi</i> A.Murray bis <i>et al.</i>	1 000	600	300
<i>Pinus kesiya</i> Royle ex Gordon ('khasya')	1 000	80	40
<i>Pinus koraiensis</i> Siebold & Zucc.	1 000	2 000	1 000
<i>Pinus lambertiana</i> Douglas	1 000	1 000	500
<i>Pinus merkusii</i> Jungh. & de Vriese	1 000	120	60
<i>Pinus monticola</i> Douglas ex D.Don	1 000	90	45
<i>Pinus mugo</i> Turra	1 000	40	20
<i>Pinus muricata</i> D.Don	1 000	50	25
<i>Pinus nigra</i> J.F.Arnold	1 000	100	50
<i>Pinus oocarpa</i> Schiede ex Schltdl.	1 000	70	35
<i>Pinus palustris</i> Mill.	1 000	500	250
<i>Pinus parviflora</i> Siebold & Zucc.	1 000	500	250
<i>Pinus patula</i> Schltdl. & Cham.	1 000	40	20
<i>Pinus peuce</i> Griseb.	1 000	240	120
<i>Pinus pinaster</i> Aiton	1 000	240	120
<i>Pinus pinea</i> L.	1 000	1 000	1 000
<i>Pinus ponderosa</i> P.Lawson & C.Lawson	1 000	200	100
<i>Pinus pumila</i> (Pall.) Regel	1 000	40	20
<i>Pinus radiata</i> D.Don	1 000	160	80
<i>Pinus resinosa</i> Aiton	1 000	50	25
<i>Pinus rigida</i> Mill.	1 000	40	20
<i>Pinus strobus</i> L.	1 000	90	45
<i>Pinus sylvestris</i> L.	1 000	40	20

Table 2C Part 2. Lot sizes and sample sizes: tree and shrub seeds (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Pinus tabuliformis</i> Carrière	1 000	100	50
<i>Pinus taeda</i> L.	1 000	140	70
<i>Pinus taiwanensis</i> Hayata	1 000	100	50
<i>Pinus thunbergii</i> Parl.	1 000	70	35
<i>Pinus virginiana</i> Mill.	1 000	50	25
<i>Pinus wallichiana</i> A.B.Jacks.	1 000	250	125
<i>Platanus</i> spp.	1 000	25	6
<i>Platycladus orientalis</i> (L.) Franco	1 000	120	60
<i>Populus</i> spp.	50	5	2
<i>Prunus avium</i> (L.) L.	1 000	900	450
<i>Prunus padus</i> L.	1 000	360	180
<i>Prunus persica</i> (L.) Batsch	5 000	500 seeds	500 seeds
<i>Prunus serotina</i> Ehrh.	1 000	500	250
<i>Prunus</i> spp. (TSW ≤ 200 g)	1 000	1 000	500
<i>Prunus</i> spp. (TSW > 200 g)	1 000	500 seeds	500 seeds
<i>Pseudotsuga menziesii</i> (Mirb.) Franco	1 000	60	30
<i>Pyrus</i> spp.	1 000	180	90
<i>Quercus</i> spp.	5 000	500 seeds	500 seeds
<i>Robinia pseudoacacia</i> L.	1 000	100	50
<i>Rosa</i> spp.	1 000	50	25
<i>Salix</i> spp.	50	5	2
<i>Senegalia</i> spp.	1 000	70	35
<i>Sequoia sempervirens</i> (D.Don) Endl.	1 000	25	12
<i>Sequoiadendron giganteum</i> (Lindl.) J.Buchholz	1 000	25	12
<i>Sorbus</i> spp.	1 000	25	10
<i>Spartium junceum</i> L.	1 000	40	20
<i>Styphnolobium japonicum</i> (L.) Schott	1 000	100	50
<i>Syringa</i> spp.	1 000	30	15
<i>Taxodium distichum</i> (L.) Rich.	300	500	250
<i>Taxus</i> spp.	1 000	320	160
<i>Tectona grandis</i> L. f.	1 000	2 000	1 000
<i>Thuja occidentalis</i> L.	1 000	25	4
<i>Thuja plicata</i> Donn ex D.Don	1 000	10	3
<i>Tilia cordata</i> Mill.	1 000	180	90
<i>Tilia platyphyllos</i> Scop.	1 000	500	250
<i>Tsuga canadensis</i> (L.) Carrière	1 000	25	7
<i>Tsuga heterophylla</i> (Raf.) Sarg.	1 000	10	4
<i>Ulmus americana</i> L.	1 000	30	15
<i>Ulmus parvifolia</i> Jacq.	1 000	20	8
<i>Ulmus pumila</i> L.	1 000	30	15
<i>Vachellia</i> spp.	1 000	70	35
<i>Viburnum opulus</i> L.	1 000	160	80
<i>Zelkova serrata</i> (Thunb.) Makino	1 000	60	30

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Abutilon</i> × <i>hybridum</i> hort. ex Voss	5 000	40	10
<i>Achillea clavennae</i> L.	5 000	5	0.5
<i>Achillea filipendulina</i> Lam.	5 000	5	0.5
<i>Achillea ptarmica</i> L.	5 000	5	0.5
<i>Achillea umbellata</i> Sm.	5 000	5	0.5
<i>Adonis vernalis</i> L.	5 000	20	5
<i>Ageratum houstonianum</i> Mill.	5 000	5	0.5
<i>Agrimonia eupatoria</i> L.	5 000	200	50
<i>Alcea rosea</i> L.	5 000	80	20
<i>Althaea</i> hybrids	5 000	80	20
<i>Althaea officinalis</i> L.	5 000	80	20
<i>Alyssum argenteum</i> All.	5 000	10	3
<i>Alyssum montanum</i> L.	5 000	10	3
<i>Amaranthus caudatus</i> L.	5 000	10	2
<i>Amaranthus cruentus</i> L.	5 000	10	2
<i>Amaranthus hybridus</i> L.	5 000	10	2
<i>Amaranthus tricolor</i> L.	5 000	10	2
<i>Amberboa moschata</i> (L.) DC.	5 000	40	10
<i>Ammobium alatum</i> R.Br.	5 000	5	1
(<i>Anagallis arvensis</i> L. see <i>Lysimachia arvensis</i> (L.) U.Manns & Anderb.)			
<i>Anchusa azurea</i> Mill.	5 000	100	25
<i>Anchusa capensis</i> Thunb.	5 000	40	10
<i>Anemone coronaria</i> L.	5 000	10	3
<i>Anemone pulsatilla</i> L.	5 000	10	3
<i>Anemone sylvestris</i> L.	5 000	10	3
<i>Angelica archangelica</i> L.	5 000	40	10
<i>Antirrhinum majus</i> L.	5 000	5	0.5
<i>Aquilegia alpina</i> L.	5 000	20	4
<i>Aquilegia canadensis</i> L.	5 000	20	4
<i>Aquilegia chrysantha</i> A.Gray	5 000	20	4
<i>Aquilegia</i> × <i>cultorum</i> Bergmans	5 000	20	4
<i>Aquilegia vulgaris</i> L.	5 000	20	4
(<i>Arabis alpina</i> L. see <i>Arabis alpina</i> L. subsp. <i>alpina</i>)			
<i>Arabis alpina</i> L. subsp. <i>alpina</i> (previously <i>Arabis alpina</i> L.)	5 000	10	2
<i>Arabis alpina</i> L. subsp. <i>caucasica</i> (Willd.) Briq. (previously <i>Arabis caucasica</i> Willd.)	5 000	10	2
<i>Arabis</i> × <i>arendsii</i> H.R.Wehrh.	5 000	10	2
<i>Arabis blepharophylla</i> Hook. & Arn.	5 000	10	2
(<i>Arabis caucasica</i> Willd. see <i>Arabis alpina</i> L. subsp. <i>caucasica</i> (Willd.) Briq.)			
<i>Arabis procurrens</i> Waldst. & Kit.	5 000	10	2
<i>Arabis scopoliana</i> Boiss.	5 000	10	2
(<i>Arctotis stoechadifolia</i> P.J.Bergius see <i>Arctotis venusta</i> Norl.)			
<i>Arctotis venusta</i> Norl. (previously <i>Arctotis stoechadifolia</i> P.J.Bergius)	5 000	20	4
<i>Armeria maritima</i> (Mill.) Willd.	5 000	20	5
<i>Artemisia absinthium</i> L.	5 000	5	0.5
<i>Artemisia dracunculus</i> L.	5 000	5	0.5
<i>Artemisia maritima</i> L.	5 000	5	0.5
<i>Artemisia vulgaris</i> L.	5 000	5	0.5
<i>Asclepias tuberosa</i> L.	5 000	130	13
<i>Asparagus aethiopicus</i> L.	10 000	200	60

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Asparagus plumosus</i> L.	10 000	200	50
<i>Aster alpinus</i> L.	5 000	20	5
<i>Aster amellus</i> L.	5 000	20	5
(<i>Aster dumosus</i> L. see <i>Symphytotrichum dumosum</i> (L.) G.L.Nesom)			
<i>Aubrieta deltoidea</i> (L.) DC. (includes <i>A. graeca</i> Griseb.)	5 000	5	1
<i>Aurinia saxatilis</i> (L.) Desv.	5 000	10	3
<i>Bassia scoparia</i> (L.) A.J.Scott	5 000	10	3
<i>Begonia</i> Semperflorens-Cultorum Group	5 000	5	0.1
<i>Begonia</i> × <i>tuberhybrida</i> Voss	5 000	5	0.1
<i>Bellis perennis</i> L.	5 000	5	0.5
<i>Betonica macrantha</i> K.Koch (previously <i>Stachys macrantha</i> (K.Koch) Stearn)	5 000	20	5
<i>Brachyscome iberidifolia</i> Benth.	5 000	5	0.3
<i>Briza maxima</i> L.	5 000	40	10
<i>Browallia viscosa</i> Kunth	5 000	5	0.5
<i>Brunnera macrophylla</i> (Adams) I.M.Johnst.	5 000	40	10
<i>Calceolaria</i> × <i>herbeohybrida</i> Voss	5 000	5	0.1
<i>Calceolaria polyantha</i> Cav.	5 000	5	0.1
<i>Calendula officinalis</i> L.	5 000	80	20
<i>Callistephus chinensis</i> (L.) Nees	5 000	20	6
<i>Campanula carpatia</i> Jacq.	5 000	5	0.2
<i>Campanula fragilis</i> Cirillo	5 000	5	1
<i>Campanula garganica</i> Ten.	5 000	5	0.5
<i>Campanula glomerata</i> L.	5 000	5	0.2
<i>Campanula lactiflora</i> M.Bieb.	5 000	5	1
<i>Campanula medium</i> L.	5 000	5	0.6
<i>Campanula persicifolia</i> L.	5 000	5	0.2
<i>Campanula portenschlagiana</i> Schult.	5 000	5	0.5
<i>Campanula pyramidalis</i> L.	5 000	5	1
<i>Campanula rapunculus</i> L.	5 000	5	1
<i>Celosia argentea</i> L.	5 000	10	2
<i>Centaurea benedicta</i> (L.) L.	5 000	300	75
<i>Centaurea cyanus</i> L.	5 000	40	10
<i>Centaurea gymnocarpa</i> Moris & D.Not.	5 000	40	10
<i>Centaurea imperialis</i> Hausskn. ex Bornm.	5 000	40	10
<i>Centaurea macrocephala</i> Muss. Puschk. ex Willd.	5 000	40	10
<i>Centaurea montana</i> L.	5 000	40	10
<i>Centaurea ragusina</i> L.	5 000	40	10
<i>Cerastium tomentosum</i> L.	5 000	10	2
<i>Chelidonium majus</i> L.	5 000	5	1
<i>Chrysanthemum indicum</i> L.	5 000	30	8
<i>Clarkia amoena</i> (Lehm.) A.Nelson & J.F.Macbr.	5 000	5	1
<i>Clarkia pulchella</i> Pursh	5 000	5	1
<i>Clarkia unguiculata</i> Lindl.	5 000	5	1
<i>Cleome hassleriana</i> Chodat	5 000	20	5
<i>Cleretum bellidiforme</i> (Burm. f.) G.D.Rowley (previously <i>Dorotheanthus bellidiformis</i> (Burm. f.) N.E.Br.)	5 000	5	0.5
<i>Cobaea scandens</i> Cav.	5 000	200	50
<i>Coix lacryma-jobi</i> L.	5 000	600	150
<i>Coleostephus multicaulis</i> (Desf.) Durieu	5 000	30	8
<i>Consolida ajacis</i> (L.) Schur	5 000	30	8
<i>Consolida regalis</i> Gray	5 000	30	8

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Convolvulus tricolor</i> L.	5 000	100	25
<i>Coreopsis basalis</i> (A.Dietr.) S.F.Blake (includes <i>C. drummondii</i> (D.Don) Torr. & A.Gray)	5 000	20	5
<i>Coreopsis lanceolata</i> L.	5 000	20	5
<i>Coreopsis maritima</i> (Nutt.) Hook. f.	5 000	5	1
<i>Coreopsis tinctoria</i> Nutt.	5 000	5	1
<i>Cosmos bipinnatus</i> Cav.	5 000	80	20
<i>Cosmos sulphureus</i> Cav.	5 000	80	20
<i>Cyclamen persicum</i> Mill.	5 000	100	30
<i>Cymbalaria muralis</i> G.Gaertn. et al.	5 000	5	0.2
<i>Cynoglossum amabile</i> Stapf & J.R.Drumm.	5 000	40	10
<i>Dahlia pinnata</i> Cav.	5 000	80	20
<i>Datura metel</i> L.	5 000	100	25
<i>Datura stramonium</i> L.	5 000	100	25
<i>Delphinium ×belladonna</i> hort. ex Bergmans	5 000	20	4
<i>Delphinium cardinale</i> Hook.	5 000	20	4
<i>Delphinium ×cultorum</i> Voss	5 000	20	4
<i>Delphinium formosum</i> Boiss. & A.Huet	5 000	20	4
<i>Delphinium grandiflorum</i> L.	5 000	20	4
<i>Dianthus barbatus</i> L.	5 000	10	3
<i>Dianthus caryophyllus</i> L.	5 000	20	5
<i>Dianthus chinensis</i> L.	5 000	10	3
<i>Dianthus deltoides</i> L.	5 000	20	0.5
<i>Dianthus plumarius</i> L.	5 000	20	5
<i>Digitalis lanata</i> Ehrh.	5 000	5	1
<i>Digitalis purpurea</i> L.	5 000	5	0.2
<i>Dimorphotheca ecklonis</i> DC. (previously <i>Osteospermum ecklonis</i> (DC.) Norl.)	5 000	40	10
<i>Dimorphotheca pluvialis</i> (L.) Moench	5 000	40	10
<i>Dimorphotheca tragus</i> (Aiton) DC.	5 000	40	10
<i>Doronicum orientale</i> Hoffm.	5 000	10	2
(<i>Dorotheanthus bellidiformis</i> (Burm. f.) N.E.Br. see <i>Cleretum bellidiforme</i> (Burm. f.) G.D.Rowley)			
<i>Echinacea purpurea</i> (L.) Moench	5 000	20	5
<i>Echinops ritro</i> L.	5 000	80	20
<i>Echium candicans</i> L. f.	5 000	40	10
<i>Echium plantagineum</i> L.	5 000	40	10
<i>Erigeron speciosus</i> (Lindl.) DC.	5 000	5	0.5
<i>Erysimum cheiri</i> (L.) Crantz	5 000	10	3
<i>Erysimum ×marshallii</i> (Henfr.) Bois	5 000	10	3
<i>Eschscholzia californica</i> Cham.	5 000	20	5
<i>Eustoma exaltatum</i> (L.) Salisb. ex G.Don	5 000	5	0.2
<i>Fatsia japonica</i> (Thunb.) Decne. & Planch.	5 000	60	15
<i>Felicia heterophylla</i> (Cass.) Grau	5 000	20	5
<i>Freesia refracta</i> (Jacq.) Klatt	5 000	100	25
<i>Gaillardia aristata</i> Pursh	5 000	30	8
<i>Gaillardia pulchella</i> Foug.	5 000	20	6
<i>Galega officinalis</i> L.	5 000	80	20
<i>Galeopsis segetum</i> Neck.	5 000	20	4
<i>Gazania rigens</i> (L.) Gaertn.	5 000	20	5
<i>Gentiana acaulis</i> L.	5 000	5	0.7
<i>Geranium hybrids</i>	5 000	40	10

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Gerbera jamesonii</i> Adlam	5 000	40	10
<i>Geum coccineum</i> Sm.	5 000	20	5
<i>Geum quellyon</i> Sweet	5 000	20	5
<i>Gilia tricolor</i> Benth.	5 000	5	1
<i>Glandularia canadensis</i> (L.) Nutt.	5 000	20	6
<i>Glandularia ×hybrida</i> (hort. ex Groenl. & Rümpler) G.L.Nesom & Pruski (previously <i>Verbena</i> Hybrida Group)	5 000	20	6
<i>Glebionis carinata</i> (Schousb.) Tzvelev	5 000	30	8
<i>Glebionis coronaria</i> (L.) Cass. ex Spach	5 000	30	8
<i>Glebionis segetum</i> (L.) Fourr.	5 000	30	8
<i>Gomphrena globosa</i> L.	5 000	40	10
<i>Goniolimon tataricum</i> (L.) Boiss.	5 000	20	5
<i>Grevillea robusta</i> A.Cunn. ex R.Br.	5 000	80	20
<i>Gypsophila elegans</i> M.Bieb.	5 000	10	2
<i>Gypsophila paniculata</i> L.	5 000	10	2
<i>Gypsophila repens</i> L.	5 000	10	2
<i>Helenium autumnale</i> L.	5 000	5	0.9
<i>Helianthemum nummularium</i> (L.) Mill.	5 000	20	5
<i>Helianthus debilis</i> Nutt.	10 000	150	40
<i>Heliopsis helianthoides</i> (L.) Sweet	5 000	40	10
<i>Heliotropium arborescens</i> L.	5 000	5	1
<i>Hesperis matronalis</i> L.	5 000	20	5
<i>Heteranthemis viscidiflora</i> Schott	5 000	30	8
<i>Heuchera sanguinea</i> Engelm.	5 000	5	0.1
<i>Hibiscus trionum</i> L.	5 000	40	10
<i>Hippeastrum hybrids</i>	5 000	80	20
<i>Hypericum perforatum</i> L.	5 000	5	0.3
<i>Hyssopus officinalis</i> L.	5 000	10	3
<i>Iberis amara</i> L.	5 000	20	6
<i>Iberis gibraltarica</i> L.	5 000	10	3
<i>Iberis sempervirens</i> L.	5 000	10	3
<i>Iberis umbellata</i> L.	5 000	10	3
<i>Impatiens balsamina</i> L.	5 000	100	25
<i>Impatiens walleriana</i> Hook. f.	5 000	10	2
<i>Inula helenium</i> L.	5 000	20	4
<i>Ipomoea alba</i> L.	10 000	400	100
<i>Ipomoea purpurea</i> (L.) Roth	10 000	400	100
<i>Ipomoea quamoclit</i> L.	10 000	200	50
<i>Ipomoea tricolor</i> Cav.	10 000	400	100
<i>Jacobaea maritima</i> (L.) Pelser & Meijden	5 000	5	0.5
<i>Kalanchoe blossfeldiana</i> Poelln.	5 000	5	0.1
<i>Kalanchoe crenata</i> (Andrews) Haw.	5 000	5	0.1
<i>Kalanchoe globulifera</i> H.Perrier	5 000	5	0.1
<i>Kniphofia uvaria</i> (L.) Oken	5 000	10	3
<i>Lathyrus latifolius</i> L.	10 000	400	100
<i>Lathyrus odoratus</i> L.	10 000	600	150
<i>Lavandula angustifolia</i> Mill.	5 000	10	2
<i>Lavatera trimestris</i> L.	5 000	40	10
<i>Legousia speculum-veneris</i> (L.) Chaix	5 000	5	1
<i>Leontopodium nivale</i> (Ten.) Hand.-Mazz.	5 000	5	0.1
<i>Leonurus cardiaca</i> L.	5 000	10	2
<i>Leucanthemum maximum</i> (Ramond) DC.	5 000	20	5

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Leucanthemum vulgare</i> Lam.	5 000	20	5
<i>Levisticum officinale</i> W.D.J.Koch	5 000	30	8
<i>Liatris pycnostachya</i> Michx.	5 000	30	8
<i>Liatris spicata</i> (L.) Willd.	5 000	30	8
<i>Lilium regale</i> E.H.Wilson	5 000	40	10
<i>Limonium bellidifolium</i> (Gouan) Dumort.	5 000	20	5
(<i>Limonium bonduellei</i> (T.Lestib.) Kuntze see <i>Limonium sinuatum</i> (L.) Mill. subsp. <i>bonduellei</i> (T.Lestib.) Sauvage & Vindt)			
<i>Limonium gerberi</i> Soldano	5 000	20	5
(<i>Limonium sinuatum</i> (L.) Mill. see <i>Limonium sinuatum</i> (L.) Mill. subsp. <i>sinuatum</i>)			
<i>Limonium sinuatum</i> (L.) Mill. subsp. <i>bonduellei</i> (T.Lestib.) Sauvage & Vindt (previously <i>Limonium bonduellei</i> (T.Lestib.) Kuntze)	5 000	200	50
<i>Limonium sinuatum</i> (L.) Mill. subsp. <i>sinuatum</i> (previously <i>Limonium sinuatum</i> (L.) Mill.) (heads)	5 000	200	50
<i>Limonium sinuatum</i> (L.) Mill. subsp. <i>sinuatum</i> (previously <i>Limonium sinuatum</i> (L.) Mill.) (seeds)	5 000	20	6
<i>Linaria bipartita</i> (Vent.) Willd.	5 000	5	0.2
<i>Linaria maroccana</i> Hook. f.	5 000	5	0.4
<i>Linaria vulgaris</i> Mill.	5 000	5	0.2
<i>Linum flavum</i> L.	5 000	20	5
<i>Linum grandiflorum</i> Desf.	5 000	40	10
<i>Linum narbonense</i> L.	5 000	20	5
<i>Linum perenne</i> L.	5 000	20	5
<i>Lobelia cardinalis</i> L. (includes <i>L. fulgens</i> Humb. & Bonpl. ex Willd.)	5 000	5	0.1
<i>Lobelia erinus</i> L.	5 000	5	0.2
<i>Lobularia maritima</i> (L.) Desv.	5 000	5	1
<i>Lomelosia caucasica</i> (M.Bieb.) Greuter & Burdet	5 000	80	20
<i>Lonas annua</i> (L.) Vines & Druce	5 000	5	0.6
<i>Lunaria annua</i> L.	5 000	80	20
(<i>Lupinus hartwegii</i> Lindl. see <i>Lupinus mexicanus</i> Cerv. ex Lag.)			
<i>Lupinus</i> hybrids	10 000	200	60
<i>Lupinus mexicanus</i> Cerv. ex Lag. (previously <i>Lupinus hartwegii</i> Lindl.)	10 000	200	60
<i>Lupinus nanus</i> Douglas ex Benth.	10 000	200	60
<i>Lupinus polyphyllus</i> Lindl.	10 000	200	60
<i>Lysimachia arvensis</i> (L.) U.Manns & Anderb. (previously <i>Anagallis arvensis</i> L.)	5 000	10	2
<i>Malcolmia maritima</i> (L.) W.T.Aiton	5 000	10	3
<i>Malope trifida</i> Cav.	5 000	20	5
<i>Malva sylvestris</i> L.	5 000	30	15
<i>Marrubium vulgare</i> L.	5 000	10	2
<i>Matricaria chamomilla</i> L.	5 000	5	0.5
<i>Matthiola incana</i> (L.) W.T.Aiton	5 000	20	4
<i>Matthiola longipetala</i> (Vent.) DC.	5 000	10	2
<i>Melissa officinalis</i> L.	5 000	10	2
<i>Mentha ×piperita</i> L.	5 000	5	0.5
<i>Mimosa pudica</i> L.	5 000	40	10
<i>Mimulus cardinalis</i> Douglas ex Benth.	5 000	5	0.2
<i>Mimulus cupreus</i> hort. ex Dombrain	5 000	5	0.2
<i>Mimulus ×hybridus</i> hort. ex Voss	5 000	5	0.2

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Mimulus luteus</i> L.	5 000	5	0.2
<i>Mirabilis jalapa</i> L.	10 000	800	200
<i>Moluccella laevis</i> L.	5 000	100	25
<i>Myosotis</i> hybrids	5 000	10	2
<i>Myosotis scorpioides</i> L.	5 000	10	2
<i>Myosotis sylvatica</i> Hoffm.	5 000	10	2
<i>Nemesia strumosa</i> Benth.	5 000	5	1
<i>Nemesia versicolor</i> E.Mey. ex Benth.	5 000	5	1
<i>Nemophila maculata</i> Benth. ex Lindl.	5 000	20	5
<i>Nemophila menziesii</i> Hook. & Arn.	5 000	20	5
<i>Nepeta cataria</i> L.	5 000	10	2
<i>Nicotiana alata</i> Link & Otto	5 000	5	0.2
<i>Nicotiana</i> * <i>sanderi</i> W.Watson	5 000	5	0.2
<i>Nicotiana suaveolens</i> Lehm.	5 000	5	0.5
<i>Nierembergia hippomanica</i> Miers	5 000	5	0.5
<i>Nigella damascena</i> L.	5 000	20	6
<i>Nigella hispanica</i> L.	5 000	20	6
<i>Nigella sativa</i> L.	5 000	40	10
<i>Oenothera macrocarpa</i> Nutt.	5 000	40	10
(<i>Osteospermum ecklonis</i> (DC.) Norl. see <i>Dimorphotheca ecklonis</i> DC.)			
<i>Papaver alpinum</i> L.	5 000	5	0.5
<i>Papaver glaucum</i> Boiss. & Hausskn.	5 000	5	0.5
<i>Papaver nudicaule</i> L.	5 000	5	0.5
<i>Papaver orientale</i> L.	5 000	5	1
<i>Papaver rhoeas</i> L.	5 000	5	0.5
<i>Pelargonium</i> Zonale Group	5 000	80	20
<i>Penstemon barbatus</i> (Cav.) Roth	5 000	10	2
<i>Penstemon hartwegii</i> Benth.	5 000	10	2
<i>Penstemon</i> hybrids	5 000	10	2
<i>Pericallis cruenta</i> (Masson ex L'Hér.) Bolle	5 000	5	0.5
<i>Perilla frutescens</i> (L.) Britton	5 000	10	3
<i>Petunia</i> * <i>atkinsiana</i> (Sweet) D.Don ex W.H.Baxter	5 000	5	0.2
<i>Phacelia campanularia</i> A.Gray	5 000	10	2
<i>Phlox drummondii</i> Hook.	5 000	20	5
<i>Phlox paniculata</i> L.	5 000	20	5
<i>Phlox subulata</i> L.	5 000	20	5
<i>Pholistoma auritum</i> (Lindl.) Lilja	5 000	20	5
<i>Physalis alkekengi</i> L.	5 000	20	4
<i>Pimpinella major</i> (L.) Huds.	5 000	20	5
<i>Pimpinella saxifraga</i> L.	5 000	20	5
<i>Plectocephalus americana</i> (Nutt.) D.Don	5 000	100	35
<i>Plectranthus scutellarioides</i> (L.) R.Br.	5 000	10	2
<i>Portulaca grandiflora</i> Hook.	5 000	5	0.3
<i>Primula auricula</i> L.	5 000	5	1
<i>Primula denticulata</i> Sm.	5 000	5	0.5
<i>Primula elatior</i> (L.) Hill	5 000	10	2
<i>Primula japonica</i> A.Gray	5 000	5	1
<i>Primula</i> * <i>kewensis</i> W.Watson	5 000	5	0.5
<i>Primula malacoides</i> Franch.	5 000	5	0.5
<i>Primula obconica</i> Hance	5 000	5	0.5
<i>Primula praeinitens</i> Ker Gawl.	5 000	5	1

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submit- ted sample (g)	Minimum working sam- ple for purity analysis (3.5.1) (g)
1	2	3	4
<i>Primula veris</i> L.	5 000	5	1
<i>Primula vulgaris</i> Huds.	5 000	5	1
<i>Psephellus dealbatus</i> (Willd.) K.Koch	5 000	40	10
<i>Psylliostachys suworowii</i> (Regel) Roshkova	5 000	20	5
<i>Ranunculus asiaticus</i> L.	5 000	5	1
<i>Reseda odorata</i> L.	5 000	10	3
<i>Rheum palmatum</i> L.	5 000	100	30
<i>Rhodanthe humboldtiana</i> (Gaudich.) Paul G.Wilson	5 000	30	8
<i>Rhodanthe manglesii</i> Lindl.	5 000	30	8
<i>Rhodanthe chlorocephala</i> (Turcz.) Paul G.Wilson (includes <i>Helipterum roseum</i> (Hook.) Benth.)	5 000	30	8
<i>Rudbeckia fulgida</i> Aiton	5 000	10	2
<i>Rudbeckia hirta</i> L.	5 000	5	1
<i>Ruta graveolens</i> L.	5 000	20	6
<i>Saintpaulia ionantha</i> H.Wendl.	5 000	5	0.1
<i>Salpiglossis sinuata</i> Ruiz & Pav.	5 000	5	1
<i>Salvia coccinea</i> Buc'hoz ex Etl.	5 000	30	8
<i>Salvia farinacea</i> Benth.	5 000	20	5
<i>Salvia officinalis</i> L.	5 000	30	20
<i>Salvia patens</i> Cav.	5 000	30	8
<i>Salvia pratensis</i> L.	5 000	30	8
<i>Salvia sclarea</i> L.	5 000	80	20
<i>Salvia splendens</i> Sellow ex Nees	5 000	30	8
<i>Salvia viridis</i> L.	5 000	20	5
<i>Sanvitalia procumbens</i> Lam.	5 000	10	2
<i>Saponaria calabrica</i> Guss.	5 000	20	5
<i>Saponaria ocymoides</i> L.	5 000	20	5
<i>Saponaria officinalis</i> L.	5 000	20	5
<i>Scabiosa atropurpurea</i> L.	5 000	60	15
<i>Schefflera elegantissima</i> (hort. Veitch ex Mast.) Lowry & Frodin	5 000	20	6
<i>Schizanthus pinnatus</i> Ruiz & Pav.	5 000	10	2
<i>Senecio elegans</i> L.	5 000	5	0.5
<i>Silene chalcedonica</i> (L.) E.H.L.Krause	5 000	5	1
<i>Silene coronaria</i> (L.) Clairv.	5 000	20	5
<i>Silene pendula</i> L.	5 000	10	2
<i>Silybum marianum</i> (L.) Gaertn.	5 000	200	50
<i>Sinningia speciosa</i> (Lodd. et al. ex Ker Gawl.) Hiern	5 000	5	0.2
<i>Solanum giganteum</i> Jacq.	5 000	20	5
<i>Solanum laciniatum</i> Aiton	5 000	20	5
<i>Solanum marginatum</i> L. f.	5 000	20	5
<i>Solanum pseudocapsicum</i> L.	5 000	20	5
(<i>Stachys macrantha</i> (K.Koch) Stearn see <i>Betonica macrantha</i> K.Koch)			
<i>Symphotrichum dumosum</i> (L.) G.L.Nesom (previously <i>Aster</i> <i>dumosus</i> L.)	5 000	20	5
<i>Tagetes erecta</i> L.	5 000	40	10
<i>Tagetes patula</i> L.	5 000	40	10
<i>Tagetes tenuifolia</i> Cav.	5 000	20	5
<i>Tanacetum achilleifolium</i> (M.Bieb.) Sch. Bip.	5 000	30	8
<i>Tanacetum cinerariifolium</i> (Trevir.) Sch. Bip.	5 000	10	3
<i>Tanacetum coccineum</i> (Willd.) Grierson	5 000	30	8
<i>Tanacetum parthenium</i> (L.) Sch. Bip.	5 000	20	5

Table 2C Part 3. Lot sizes and sample sizes: flower, spice, herb and medicinal species (continued)

Species	Maximum weight of lot (kg) (except see 2.8 Note 2)	Minimum submitted sample (g)	Minimum working sample for purity analysis (3.5.1) (g)
1	2	3	4
<i>Thunbergia alata</i> Bojer ex Sims	5 000	200	50
<i>Thymus serpyllum</i> L.	5 000	5	0.5
<i>Torenia fournieri</i> Linden ex E.Fourn.	5 000	5	0.2
<i>Tripleurospermum inodorum</i> (L.) Sch. Bip.	5 000	5	0.5
<i>Tripleurospermum maritimum</i> (L.) W.D.J.Koch	5 000	5	0.5
<i>Tropaeolum majus</i> L.	10 000	1 000	350
<i>Tropaeolum peltophorum</i> Benth.	10 000	1 000	350
<i>Tropaeolum peregrinum</i> L.	10 000	1 000	350
<i>Vaccaria hispanica</i> (Mill.) Rauschert	5 000	20	5
<i>Valeriana officinalis</i> L.	5 000	10	2
<i>Verbascum densiflorum</i> Bertol.	5 000	5	0.3
<i>Verbascum phlomoides</i> L.	5 000	5	0.5
<i>Verbascum thapsus</i> L.	5 000	5	0.5
<i>Verbena bonariensis</i> L.	5 000	20	6
(Verbena Hybrida Group see <i>Glandularia ×hybrida</i> (hort. ex Groenl. & Rümpler) G.L.Nesom & Pruski)			
<i>Verbena rigida</i> Spreng.	5 000	10	2
<i>Vinca minor</i> L.	5 000	20	5
<i>Viola cornuta</i> L.	5 000	10	3
<i>Viola odorata</i> L.	5 000	10	3
<i>Viola tricolor</i> L.	5 000	10	3
<i>Xeranthemum annuum</i> L.	5 000	10	3
<i>Xerochrysum bracteatum</i> (Vent.) Tzvelev	5 000	10	2
<i>Zinnia elegans</i> Jacq.	5 000	80	20
<i>Zinnia haageana</i> Regel	5 000	20	6

Table 2D Part 1. Sample sizes (numbers of seeds) for pelleted seeds, encrusted seed and seed granules

Determinations	Minimum submitted sample	Minimum working sample
Purity analysis (including verification of species)	2 500	2 500
Thousand-seed weight	2 500	Pure pellet fraction
Germination	2 500	400
Determination of other seeds	10 000	7 500
Determination of other seeds (encrusted seeds and seed granules)	25 000	25 000
Size grading	5 000	1 000

Table 2D Part 2. Sample sizes (number of seeds) for seed tapes and mats

Determinations	Minimum submitted sample	Minimum working sample
Verification of species	300	100
Germination	2 000	400
Purity analysis (if required)	2 500	2 500
Determination of other seeds	10 000	7 500

2.9 Heterogeneity testing for seed lots in multiple containers

The object of heterogeneity testing is to detect the presence of heterogeneity which makes the seed lot technically unacceptable for sampling according to the object as defined in 2.1.

2.9.1 The H value test

2.9.1.1 Definitions of terms and symbols

The testing of predominantly in-range heterogeneity of an attribute adopted as an indicator involves a comparison between the observed variance and the acceptable variance of that attribute. The container-samples of a seed lot are samples drawn independently of each other from different containers. The examinations of container-samples for the indicating attribute must also be mutually independent. Since there is only one source of information for each container, heterogeneity within containers is not directly involved. The acceptable variance is calculated by multiplying the theoretical variance caused by random variation with a factor f for additional variation, taking into account the level of heterogeneity which is achievable in good seed production practice. The theoretical variance can be calculated from the respective probability distributions, which is the binomial distribution in the case of purity and germination, and the Poisson distribution in the case of the other seed count.

N number of containers in the lot

N number of independent container-samples

n number of seeds tested from each container-sample (1000 for purity, 100 for germination and 2500 for other seed count, see 2.9.1.3)

X test result of the adopted attribute in a container-sample

$\sum$ symbol for sum of all values

f factor for multiplying the theoretical variance to obtain the acceptable variance (see Table 2E)

Mean of all X values determined for the lot in respect of the adopted attribute:

$$\bar{X} = \frac{\sum X}{N}$$

Acceptable variance of independent container-samples in respect of purity or germination percentages:

$$W = \frac{\bar{X} \times (100 - \bar{X})}{n} \times f$$

Acceptable variance of independent container-samples in respect of number of other seeds:

$$W = \bar{X} \times f$$

Observed variance of independent container-samples based on all X values in respect of the adopted attribute:

$$V = \frac{N \sum X^2 - (\sum X)^2}{N(N-1)}$$

H value:

$$H = \frac{V}{W} - f$$

Negative H values are reported as zero.

Table 2E. Factors for additional variation in seed lots to be used for calculating W and finally the H value

Attributes	Non-chaffy seeds	Chaffy seeds
Purity	1.1	1.2
Other seed count	1.4	2.2
Germination	1.1	1.2

Remarks:

- For purity and germination calculate to two decimal places if N is less than 10 and to three decimal places if N is 10 or more.
- For the number of other seeds, calculate to one decimal place if N is less than 10, and to two decimal places if N is 10 or more.
- For definition of non-chaffy and chaffy seeds see 3.6.6 of the ISTA Rules. The chaffiness of various genera is listed in Table 3B Part 1.

Table 2F. Sampling intensity and critical H values. Number of independent container samples to be drawn as depending on the number of containers in the lot and critical H values for seed lot heterogeneity at a significance level of 1 % probability

Number of containers in the lot	Number of independent container samples	Critical H value for purity and germination attributes		Critical H value for other seed count attributes	
		non-chaffy seeds	chaffy seeds	non-chaffy seeds	chaffy seeds
5	5	2.55	2.78	3.25	5.10
6	6	2.22	2.42	2.83	4.44
7	7	1.98	2.17	2.52	3.98
8	8	1.80	1.97	2.30	3.61
9	9	1.66	1.81	2.11	3.32
10	10	1.55	1.69	1.97	3.10
11–15	11	1.45	1.58	1.85	2.90
16–25	15	1.19	1.31	1.51	2.40
26–35	17	1.10	1.20	1.40	2.20
36–49	18	1.07	1.16	1.36	2.13
50 or more	20	0.99	1.09	1.26	2.00

2.9.1.2 Sampling the lot

The number of independent container samples must be not less than presented in Table 2F.

Sampling intensity has been chosen such that in a lot containing about 10 % deviating containers, at least one deviating container is selected with a probability of $p = 90\%$. Since the detection of a deviating container is conditional on selection, the power of both tests to detect heterogeneity is at best close to equal, but usually lower than the chosen selection probability. (Reference: Steiner, A. M. and Meyer, U. (1990), H value and R value heterogeneity testing of seed lots; properties, sampling intensity and precision. *Agrobiological Research* **43**, 103–114.)

The containers to be sampled are chosen strictly at random. The sample taken from the container must adequately represent the whole contents, e.g. the top, middle and bottom of a bag. The weight of each container-sample must be not less than half that specified in Table 2C, column 3.

2.9.1.3 Testing procedure

The attribute adopted to indicate heterogeneity may be:

- percentage by weight of any purity component,
- percentage of any germination test component, or
- the total number seeds or the number of any single species in the determination of other seeds by number.

In the laboratory, a working sample is drawn from each container-sample and tested independently of any other sample for the chosen attribute.

- The percentage by weight of any component may be used, provided it can be separated as in the purity analysis, e.g. pure seed, other seeds, or empty seeds of grasses. The working sample should be of such weight as is estimated to contain 1000 seeds counted from each container-sample. Each working sample is separated into two fractions: the selected component and the remainder.
- Any kind of seed or seedling determinable in a standard germination test may be used, e.g. normal seedlings, abnormal seedlings or hard seeds. From each container-sample a germination test of 100 seeds is set up simultaneously and completed in accordance with conditions specified in Table 5A.
- The seed count may be of any component that can be counted, e.g. a specified seed species, or all other seeds together. Each working sample must be of a weight estimated to contain about 2500 seeds and a count is made in it of the number of seeds of the kind selected (i.e. other seed count).

2.9.1.4 Use of Table 2F

Table 2F shows the critical H values which would be exceeded in only 1 % of tests from seed lots with an acceptable distribution of the attribute adopted as indicator. If the calculated H value exceeds the critical H value belonging to the sample number N , the attribute and the chaffiness in Table 2F, then the lot is considered to show significant heterogeneity in the in-range, or possibly also the off-range sense. If, however, the calculated H value is less than or equal to the tabulated critical H value, then the lot is considered to show no heterogeneity in the in-range, or possibly off-range sense with respect to the attribute being tested.

2.9.1.5 Reporting results

The result of the H value heterogeneity test for seed lots in multiple containers must be reported under 'Other determinations', as follows:

- $\bar{X}$: mean of all X values determined for the lot in respect of the adopted attribute;
- N : number of independent container samples;
- No : number of containers in the lot;
- the calculated H value;
- the statement: 'This H value does/does not indicate significant heterogeneity.'

Note: the H value must not be calculated or reported if $\bar{X}$ is outside the following limits:

- purity components: above 99.8 % or below 0.2 %;
- germination: above 99.0 % or below 1.0 %;
- number of specified seeds: below two per sample.

2.9.2 The R value test

The object of this test is to detect off-range heterogeneity of the seed lot using the attribute adopted as an indicator. The test for off-range heterogeneity involves comparing the maximum difference found between samples of similar size drawn from the lot with a tolerated range. This tolerated range is based on the acceptable standard deviation, which is achievable in good seed production practice.

Each independent container-sample is taken from a different container, so that heterogeneity within containers is not directly involved. Information about heterogeneity within containers is contained, however, in the acceptable standard deviation which is in fact incorporated into the tabulation of tolerated ranges. The acceptable standard deviation was calculated by the standard deviation

due to random variation according to the binomial distribution in the case of purity and germination, and to the Poisson distribution in the case of the other seed count, multiplied by the square root of the factor f given in Table 2E, respectively. The spread between containers is characterised by the calculated range to be compared with the corresponding tolerated range.

2.9.2.1 Definitions of terms and symbols

No number of containers in the lot

N number of independent container-samples

n number of seeds tested from each container-sample (1000 for purity, 100 for germination and 2500 for other seed count, see 2.9.1.3)

X test result of the adopted attribute in a container-sample

Σ symbol for sum of all values

Mean of all X values determined for the lot in respect of the adopted attribute:

$$\bar{X} = \frac{\Sigma X}{N}$$

Range found as maximum difference between independent container samples of the lot in respect of the adopted attribute:

$$R = X_{\max} - X_{\min}$$

Note: for precision of X for the R value test, see 2.9.1.1 'Remarks' to the H value test.

2.9.2.2 Sampling the lot

Sampling for the R value test is the same as for the H value test (see 2.9.1.2); the same samples must be used.

2.9.2.3 Testing procedure

The same testing procedures of purity, germination and the other seed count are used for the R value test as are used for the H value test (see 2.9.1.3). For calculations, the same set of data must be used.

2.9.2.4 Use of tables

Seed lot off-range heterogeneity is tested by using the appropriate table for tolerated, i.e. critical range:

- Table 2G for components of pure seed analyses,
- Table 2H for germination determinations, and
- Table 2I for numbers of other seeds.

Find the value $\bar{X}$ in the 'Average' columns of the appropriate table. When entering the table, round averages following the usual procedure; read off the tolerated range which would be exceeded in only 1 % of tests from seed lots with an acceptable distribution of the attribute:

- in columns 5–9 for cases when $N = 5$ to 9,
- in columns 10–19 for cases when $N = 10$ to 19, or
- in column 20 when $N = 20$.

If the calculated R value exceeds this tolerated range, then the lot is considered to show significant heterogeneity in the off-range sense. If, however, the calculated R value is less than or equal to the tabulated tolerated range, then the lot is considered to show no heterogeneity in the off-range sense with respect to the attribute being tested.

When using the tables, round averages to the next tabulated value (if in the middle, then downwards).

2.9.2.5 Reporting results

The result of the R value heterogeneity test for seed lots in multiple containers must be reported under 'Other determinations', as follows:

- $\bar{X}$: mean of all X values determined for the lot in respect of the adopted attribute;
- N : number of independent container samples;
- No : number of containers in the lot;
- the calculated R value;
- the statement: 'This R value does/does not indicate significant heterogeneity.'

2.9.3 Interpretation of results

Whenever either of the two tests, the H value test or the R value test, indicates significant heterogeneity, then the lot must be declared heterogeneous. When, however, neither of the two tests indicates significant heterogeneity, then the lot must be adopted as non-heterogeneous, having a non-significant level of heterogeneity.

Table 2G Part 1. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of purity analyses as the indicating attribute in non-chaffy seeds

Average % of the component and its complement		Tolerated range for number of independent samples (N)		
		5–9	10–19	20
99.9	0.1	0.5	0.5	0.6
99.8	0.2	0.7	0.8	0.8
99.7	0.3	0.8	0.9	1.0
99.6	0.4	1.0	1.1	1.2
99.5	0.5	1.1	1.2	1.3
99.4	0.6	1.2	1.3	1.4
99.3	0.7	1.3	1.4	1.6
99.2	0.8	1.4	1.5	1.7
99.1	0.9	1.4	1.6	1.8
99.0	1.0	1.5	1.7	1.9
98.5	1.5	1.9	2.1	2.3
98.0	2.0	2.1	2.4	2.6
97.5	2.5	2.4	2.7	2.9
97.0	3.0	2.6	2.9	3.2
96.5	3.5	2.8	3.1	3.4
96.0	4.0	3.0	3.4	3.7
95.5	4.5	3.2	3.5	3.9
95.0	5.0	3.3	3.7	4.1
94.0	6.0	3.6	4.1	4.5
93.0	7.0	3.9	4.4	4.8
92.0	8.0	4.1	4.6	5.1
91.0	9.0	4.4	4.9	5.4
90.0	10.0	4.6	5.1	5.6
89.0	11.0	4.8	5.4	5.9
88.0	12.0	5.0	5.6	6.1
87.0	13.0	5.1	5.8	6.3
86.0	14.0	5.3	5.9	6.5
85.0	15.0	5.4	6.1	6.7
84.0	16.0	5.6	6.3	6.9
83.0	17.0	5.7	6.4	7.0
82.0	18.0	5.9	6.6	7.2
81.0	19.0	6.0	6.7	7.4
80.0	20.0	6.1	6.8	7.5
78.0	22.0	6.3	7.1	7.8
76.0	24.0	6.5	7.3	8.0
74.0	26.0	6.7	7.5	8.2
72.0	28.0	6.9	7.7	8.4
70.0	30.0	7.0	7.8	8.6
68.0	32.0	7.1	8.0	8.7
66.0	34.0	7.2	8.1	8.9
64.0	36.0	7.3	8.2	9.0
62.0	38.0	7.4	8.3	9.1
60.0	40.0	7.5	8.4	9.2
58.0	42.0	7.5	8.4	9.2
56.0	44.0	7.6	8.5	9.3
54.0	46.0	7.6	8.5	9.3
52.0	48.0	7.6	8.6	9.4
50.0	50.0	7.6	8.6	9.4

Table 2G Part 2. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of purity analyses as the indicating attribute in chaffy seeds

Average % of the component and its complement		Tolerated range for number of independent samples (N)		
		5–9	10–19	20
99.9	0.1	0.5	0.6	0.6
99.8	0.2	0.7	0.8	0.9
99.7	0.3	0.9	1.0	1.1
99.6	0.4	1.0	1.1	1.2
99.5	0.5	1.1	1.3	1.4
99.4	0.6	1.2	1.4	1.5
99.3	0.7	1.3	1.5	1.6
99.2	0.8	1.4	1.6	1.7
99.1	0.9	1.5	1.7	1.8
99.0	1.0	1.6	1.8	1.9
98.5	1.5	1.9	2.2	2.4
98.0	2.0	2.2	2.5	2.7
97.5	2.5	2.5	2.8	3.1
97.0	3.0	2.7	3.0	3.3
96.5	3.5	2.9	3.3	3.6
96.0	4.0	3.1	3.5	3.8
95.5	4.5	3.3	3.7	4.1
95.0	5.0	3.5	3.9	4.3
94.0	6.0	3.8	4.2	4.6
93.0	7.0	4.1	4.6	5.0
92.0	8.0	4.3	4.8	5.3
91.0	9.0	4.6	5.1	5.6
90.0	10.0	4.8	5.4	5.9
89.0	11.0	5.0	5.6	6.1
88.0	12.0	5.2	5.8	6.4
87.0	13.0	5.4	6.0	6.6
86.0	14.0	5.5	6.2	6.8
85.0	15.0	5.7	6.4	7.0
84.0	16.0	5.8	6.6	7.2
83.0	17.0	6.0	6.7	7.4
82.0	18.0	6.1	6.9	7.5
81.0	19.0	6.3	7.0	7.7
80.0	20.0	6.4	7.1	7.8
78.0	22.0	6.6	7.4	8.1
76.0	24.0	6.8	7.6	8.4
74.0	26.0	7.0	7.8	8.6
72.0	28.0	7.2	8.0	8.8
70.0	30.0	7.3	8.2	9.0
68.0	32.0	7.4	8.3	9.1
66.0	34.0	7.5	8.5	9.3
64.0	36.0	7.6	8.6	9.4
62.0	38.0	7.7	8.7	9.5
60.0	40.0	7.8	8.8	9.6
58.0	42.0	7.9	8.8	9.7
56.0	44.0	7.9	8.9	9.7
54.0	46.0	7.9	8.9	9.8
52.0	48.0	8.0	8.9	9.8
50.0	50.0	8.0	8.9	9.8

Table 2H Part 1. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of germination tests as the indicating attribute in non-chaffy seeds

Average % of the component and its complement		Tolerated range for number of independent samples (N)		
		5–9	10–19	20
99	1	5	6	6
98	2	7	8	9
97	3	9	10	11
96	4	10	11	12
95	5	11	12	13
94	6	12	13	15
93	7	13	14	16
92	8	14	15	17
91	9	14	16	17
90	10	15	17	18
89	11	16	17	19
88	12	16	18	20
87	13	17	19	20
86	14	17	19	21
85	15	18	20	22
84	16	18	20	22
83	17	19	21	23
82	18	19	21	23
81	19	19	22	24
80	20	20	22	24
79	21	20	23	25
78	22	20	23	25
77	23	21	23	25
76	24	21	24	26
75	25	21	24	26
74	26	22	24	26
73	27	22	25	27
72	28	22	25	27
71	29	22	25	27
70	30	23	25	28
69	31	23	26	28
68	32	23	26	28
67	33	23	26	28
66	34	23	26	29
65	35	24	26	29
64	36	24	26	29
63	37	24	27	29
62	38	24	27	29
61	39	24	27	29
60	40	24	27	30
59	41	24	27	30
58	42	24	27	30
57	43	24	27	30
56	44	24	27	30
55	45	25	27	30
54	46	25	27	30
53	47	25	28	30
52	48	25	28	30
51	49	25	28	30
50	50	25	28	30

Table 2H Part 2. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of germination tests as the indicating attribute in chaffy seeds

Average % of the component and its complement		Tolerated range for number of independent samples (N)		
		5–9	10–19	20
99	1	6	6	7
98	2	8	8	9
97	3	9	10	11
96	4	10	12	13
95	5	11	13	14
94	6	12	14	15
93	7	13	15	16
92	8	14	16	17
91	9	15	17	18
90	10	16	17	19
89	11	16	18	20
88	12	17	19	21
87	13	17	20	21
86	14	18	20	22
85	15	18	21	23
84	16	19	21	23
83	17	19	22	24
82	18	20	22	24
81	19	20	23	25
80	20	21	23	25
79	21	21	24	26
78	22	21	24	26
77	23	22	24	27
76	24	22	25	27
75	25	22	25	27
74	26	23	25	28
73	27	23	26	28
72	28	23	26	28
71	29	23	26	29
70	30	24	26	29
69	31	24	27	29
68	32	24	27	29
67	33	24	27	30
66	34	24	27	30
65	35	25	27	30
64	36	25	28	30
63	37	25	28	30
62	38	25	28	31
61	39	25	28	31
60	40	25	28	31
59	41	25	28	31
58	42	25	28	31
57	43	25	28	31
56	44	26	29	31
55	45	26	29	31
54	46	26	29	31
53	47	26	29	31
52	48	26	29	31
51	49	26	29	31
50	50	26	29	31

Table 2I Part 1. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of other seed count analyses as the indicating attribute in non-chaffy seeds

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
1	6	7	7
2	8	9	10
3	10	11	12
4	11	13	14
5	13	14	15
6	14	15	17
7	15	17	18
8	16	18	19
9	17	19	21
10	18	20	22
11	19	21	23
12	19	22	24
13	20	23	25
14	21	23	26
15	22	24	26
16	22	25	27
17	23	26	28
18	24	26	29
19	24	27	30
20	25	28	30
21	25	28	31
22	26	29	32
23	27	30	33
24	27	30	33
25	28	31	34
26	28	32	35
27	29	32	35
28	29	33	36
29	30	33	37
30	30	34	37
31	31	34	38
32	31	35	38
33	32	36	39
34	32	36	39
35	33	37	40
36	33	37	41
37	34	38	41
38	34	38	42
39	34	39	42
40	35	39	43
41	35	40	43
42	36	40	44
43	36	41	44
44	37	41	45
45	37	41	45
46	37	42	46
47	38	42	46
48	38	43	47
49	39	43	47
50	39	44	48

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
51	39	44	48
52	40	45	49
53	40	45	49
54	40	45	50
55	41	46	50
56	41	46	51
57	42	47	51
58	42	47	51
59	42	47	52
60	43	48	52
61	43	48	53
62	43	49	53
63	44	49	54
64	44	49	54
65	44	50	54
66	45	50	55
67	45	50	55
68	45	51	56
69	46	51	56
70	46	52	56
71	46	52	57
72	47	52	57
73	47	53	58
74	47	53	58
75	48	53	58
76	48	54	59
77	48	54	59
78	49	54	60
79	49	55	60
80	49	55	60
81	49	55	61
82	50	56	61
83	50	56	61
84	50	56	62
85	51	57	62
86	51	57	62
87	51	57	63
88	52	58	63
89	52	58	64
90	52	58	64
91	52	59	64
92	53	59	65
93	53	59	65
94	53	60	65
95	54	60	66
96	54	60	66
97	54	61	66
98	54	61	67
99	55	61	67
100	55	62	67

Table 2I Part 1. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of other seed count analyses as the indicating attribute in non-chaffy seeds (continued)

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
101	55	62	68
102	55	62	68
103	56	62	68
104	56	63	69
105	56	63	69
106	57	63	69
107	57	64	70
108	57	64	70
109	57	64	70
110	58	65	71
111	58	65	71
112	58	65	71
113	58	65	72
114	59	66	72
115	59	66	72
116	59	66	73
117	59	67	73
118	60	67	73
119	60	67	73
120	60	67	74

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
121	60	68	74
122	61	68	74
123	61	68	75
124	61	68	75
125	61	69	75
126	62	69	76
127	62	69	76
128	62	70	76
129	62	70	76
130	63	70	77
131	63	70	77
132	63	71	77
133	63	71	78
134	64	71	78
135	64	71	78
136	64	72	78
137	64	72	79
138	64	72	79

For higher other seed counts, tolerances (R) are calculated by using the following formula and rounding up to the next whole number:

For N = 5–9: $R = \sqrt{(\text{average count of other seed}) \times 5.44}$

For N = 10–19: $R = \sqrt{(\text{average count of other seed}) \times 6.11}$

For N = 20: $R = \sqrt{(\text{average count of other seed}) \times 6.69}$

Table 2I Part 2. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of other seed count analyses as the indicating attribute in chaffy seeds

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
1	7	8	9
2	10	11	12
3	12	14	15
4	14	16	17
5	16	18	19
6	17	19	21
7	19	21	23
8	20	22	24
9	21	23	26
10	22	25	27
11	23	26	28
12	24	27	30
13	25	28	31
14	26	29	32
15	27	30	33
16	28	31	34
17	29	32	35
18	29	33	36
19	30	34	37
20	31	35	38
21	32	36	39
22	33	36	40
23	33	37	41
24	34	38	42
25	35	39	42
26	35	40	43
27	36	40	44
28	37	41	45
29	37	42	46
30	38	42	46
31	38	43	47
32	39	44	48
33	40	44	49
34	40	45	49
35	41	46	50
36	41	46	51
37	42	47	51
38	43	48	52
39	43	48	53
40	44	49	54
41	44	50	54
42	45	50	55
43	45	51	55
44	46	51	56
45	46	52	57
46	47	52	57
47	47	53	58
48	48	54	59
49	48	54	59
50	49	55	60

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
51	49	55	60
52	50	56	61
53	50	56	62
54	51	57	62
55	51	57	63
56	52	58	63
57	52	58	64
58	52	59	64
59	53	59	65
60	53	60	65
61	54	60	66
62	54	61	66
63	55	61	67
64	55	62	68
65	56	62	68
66	56	63	69
67	56	63	69
68	57	64	70
69	57	64	70
70	58	65	71
71	58	65	71
72	58	65	72
73	59	66	72
74	59	66	73
75	60	67	73
76	60	67	74
77	60	68	74
78	61	68	75
79	61	69	75
80	62	69	75
81	62	69	76
82	62	70	76
83	63	70	77
84	63	71	77
85	63	71	78
86	64	71	78
87	64	72	79
88	65	72	79
89	65	73	80
90	65	73	80
91	66	74	80
92	66	74	81
93	66	74	81
94	67	75	82
95	67	75	82
96	67	75	83
97	68	76	83
98	68	76	83
99	68	77	84
100	69	77	84

Table 2I Part 2. Maximum tolerated ranges for the R value test at a significance level of 1 % probability using components of other seed count analyses as the indicating attribute in chaffy seeds (continued)

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
101	69	77	85
102	69	78	85
103	70	78	86
104	70	79	86
105	70	79	86
106	71	79	87
107	71	80	87
108	71	80	88
109	72	80	88
110	72	81	88
111	72	81	89
112	73	81	89
113	73	82	90
114	73	82	90
115	74	83	90
116	74	83	91
117	74	83	91
118	75	84	92
119	75	84	92
120	75	84	92

Average count of other seeds	Tolerated range for number of independent samples (N)		
	5–9	10–19	20
121	76	85	93
122	76	85	93
123	76	85	93
124	76	86	94
125	77	86	94
126	77	86	95
127	77	87	95
128	78	87	95
129	78	87	96
130	78	88	96
131	79	88	96
132	79	88	97
133	79	89	97
134	79	89	98
135	80	89	98
136	80	90	98
137	80	90	99
138	81	90	99

For higher other seed counts, tolerances (R) are calculated by using the following formula and rounding up to the next whole number:

For N = 5–9: $R = \sqrt{(\text{average count of other seed}) \times 6.82}$

For N = 10–19: $R = \sqrt{(\text{average count of other seed}) \times 7.65}$

For N = 20: $R = \sqrt{(\text{average count of other seed}) \times 8.38}$

